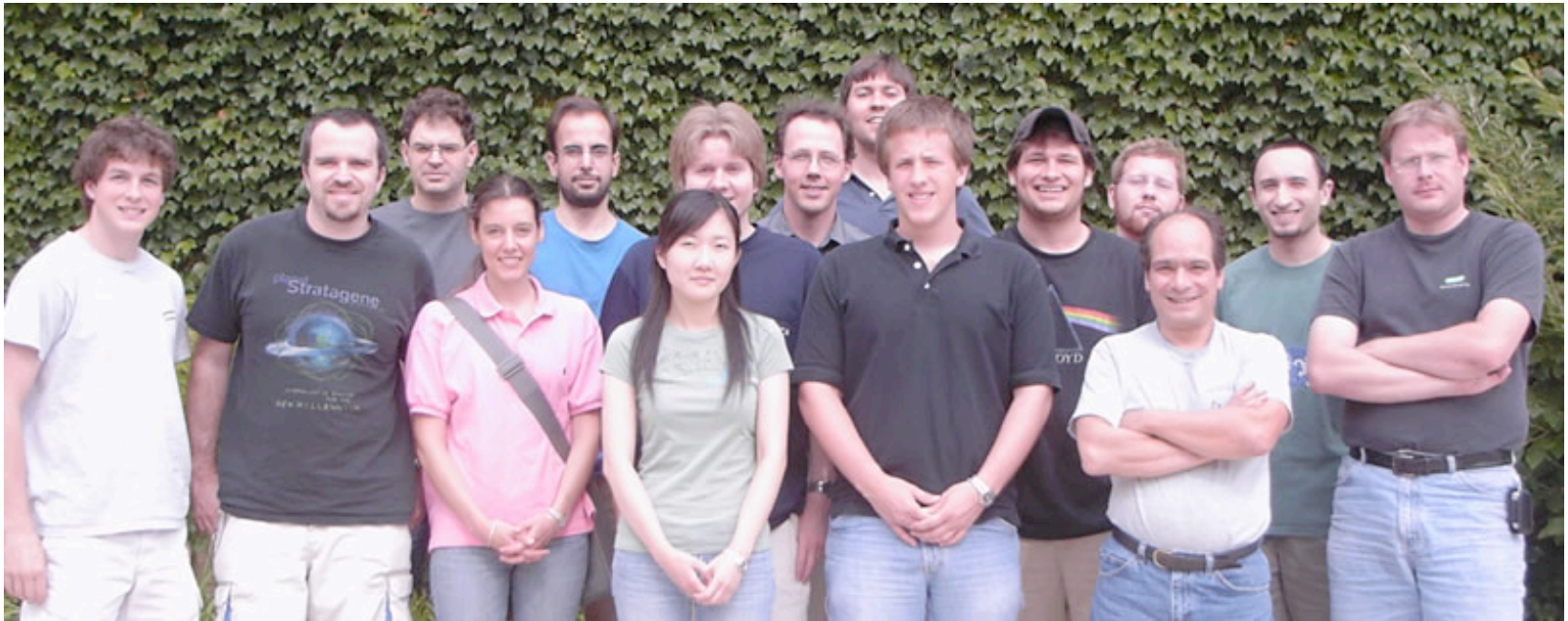
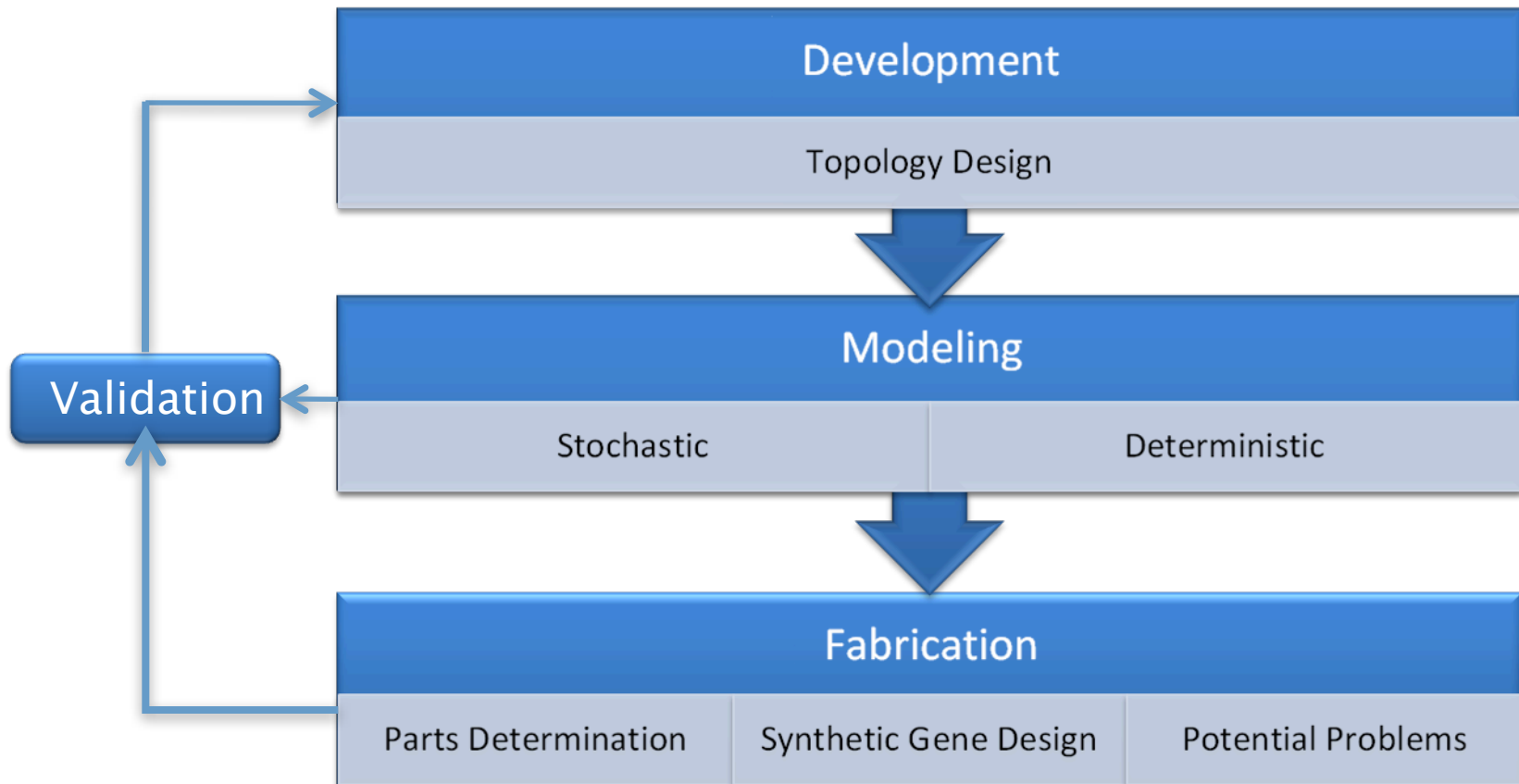


# University of Michigan iGem Team

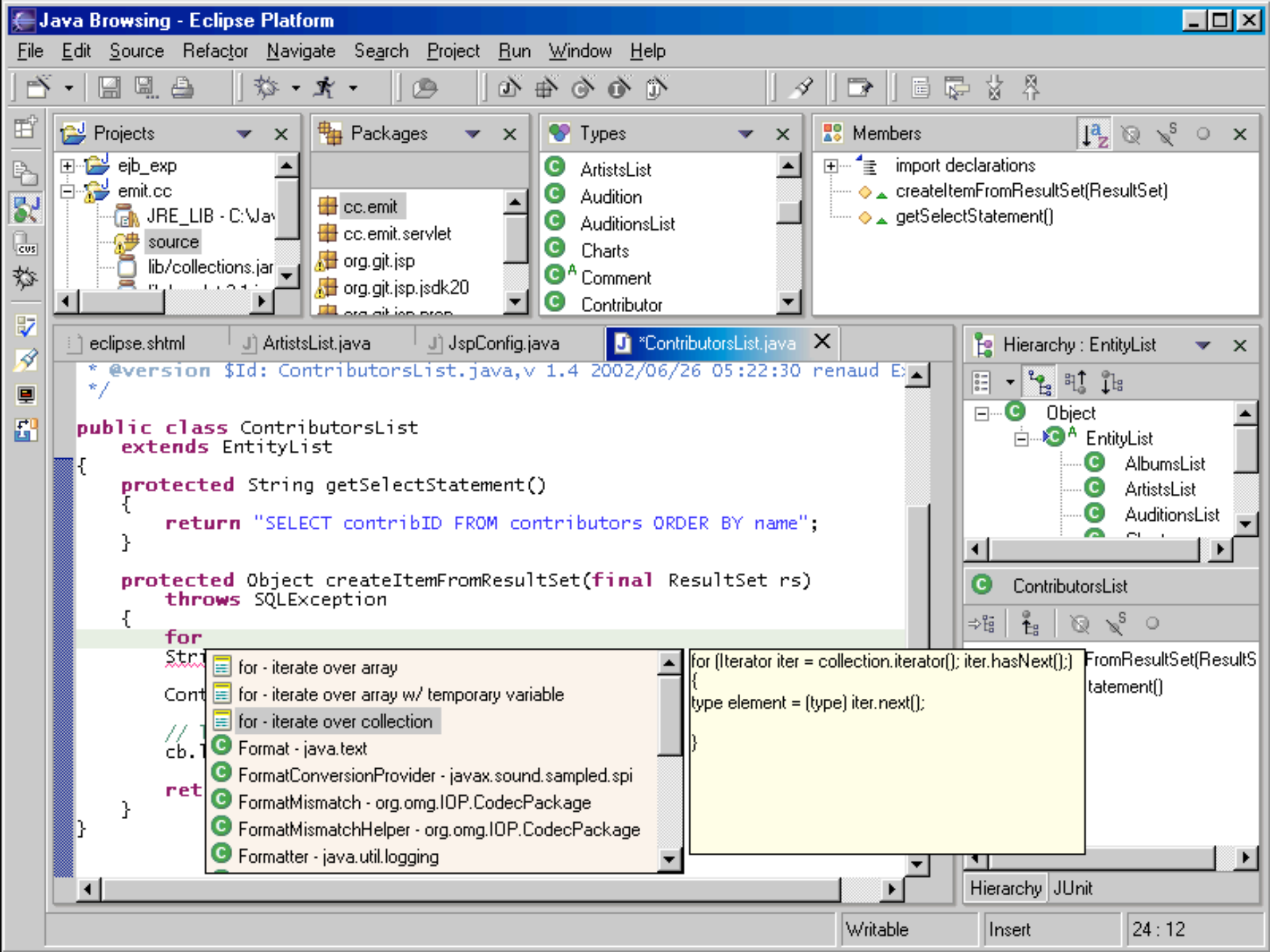


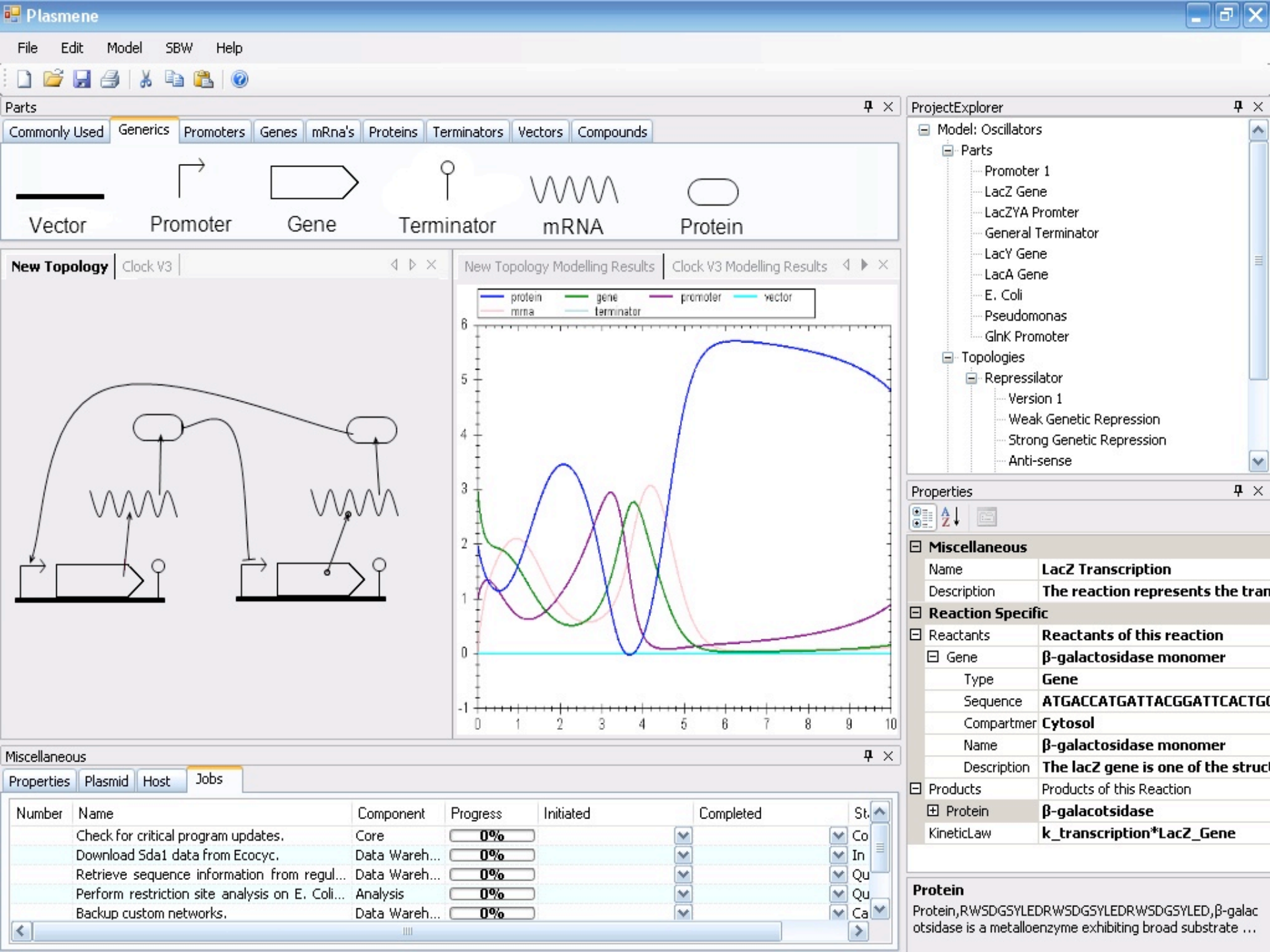
# Synthetic Biology Pipeline

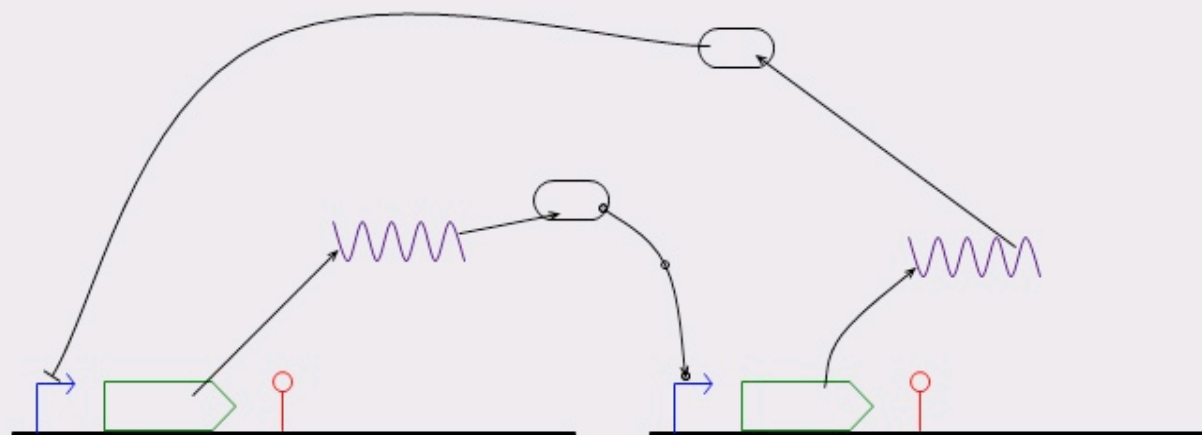


# **Plasmene**

Rapid, Computer-Aided Biological Network  
Design





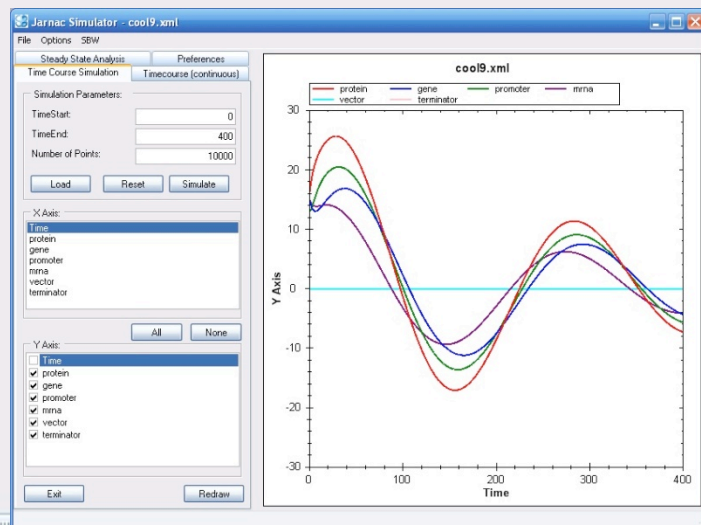
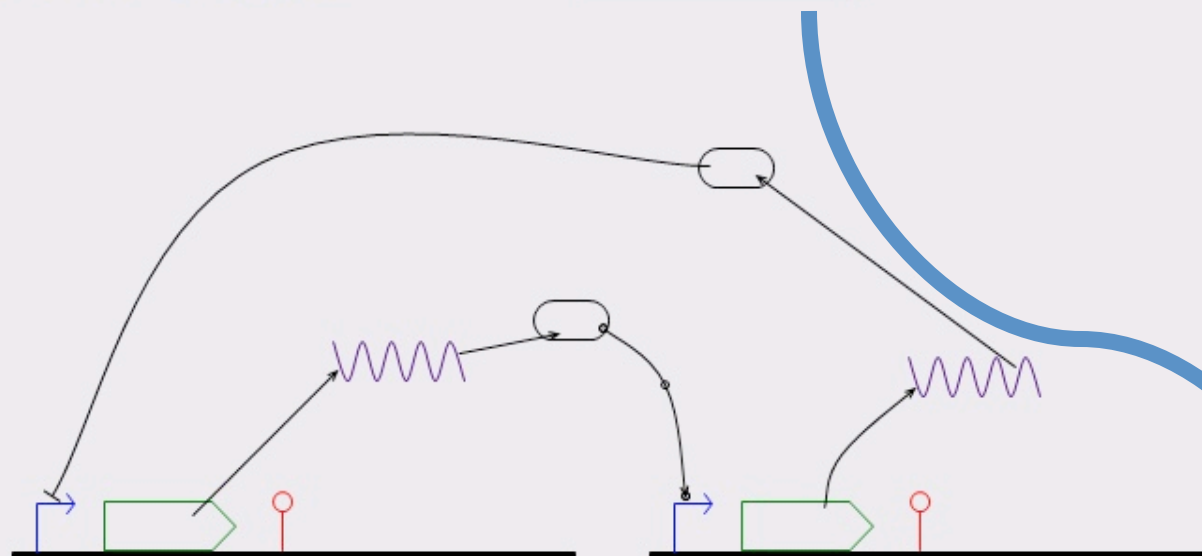
**Misc**

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CurrentArrowSettingState	
CurvePoints	PointF[] Array
DefiningPoints	
EndPoint	{X=385, Y=255}
EndWorkspaceItem	Plasmene.Diagramming.Type
IsHovered	False
IsReversible	False
IsSelected	True
KineticLaw	Plasmene.Diagramming.Type
MouseHitInformation	Plasmene.Diagramming.Hit
Name	
Points	PointF[] Array
Products	(Collection)
RateEquation	
Reactants	(Collection)
ShowValidity	False
StartPoint	{X=341, Y=169}
StartWorkspaceItem	Plasmene.Diagramming.Type
Workspace	
WorkspaceLayer	

Name



To SBML L1V1



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<xml>
  <species>
    <gene>
      <LacZ>
    </gene>
  </species>
  <reactions>
  </reactions>
```

# Technical Details

Work on Version 2 has begun.

- Developed in C#
- Uses a variation of the model view presenter design pattern
- Currently composed of two independent projects – a diagramming component and a biology class library
- Communication project is planned to allow easy integration with websites, web services, and databases

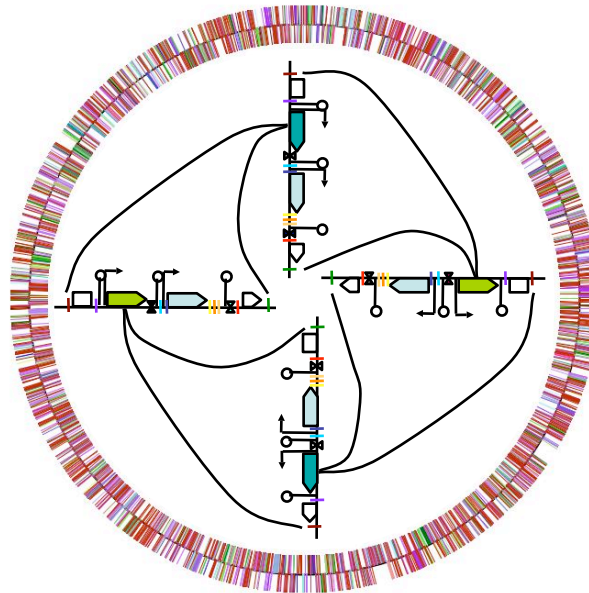
Proof-of-concept version available for download from Michigan's page on the iGEM wiki.

Interested?

Contact Paul Rogero  
[ptrog@umich.edu](mailto:ptrog@umich.edu)



# A System for Fabrication of Chromosomal Genetic Circuits by Nested Landing Pads



# Why Use Our Chromosomal Landing Pads?

- Limit Noise
- Genetically Stable
- Simple phenotypic screening and genetic confirmation
- User-friendly construction

# Landing Pad Construction



Polylinker: EcoRI, SacI, NotI, NdeI, ClaI, BamHI, SalI, XhoI, HindIII

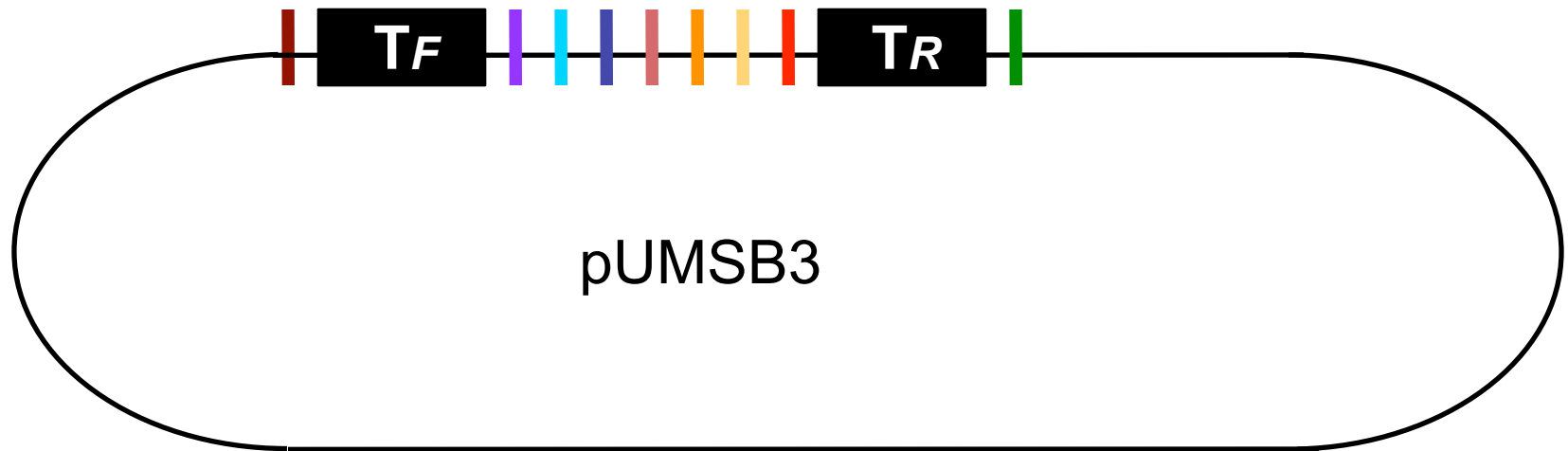
# Landing Pad Construction



Polylinker: **EcoRI**, **SacI**, **NotI**, **NdeI**, **Clal**, **BamHI**, **Sall**, **XhoI**, **HindIII**

TF - Front end of target gene

# Landing Pad Construction

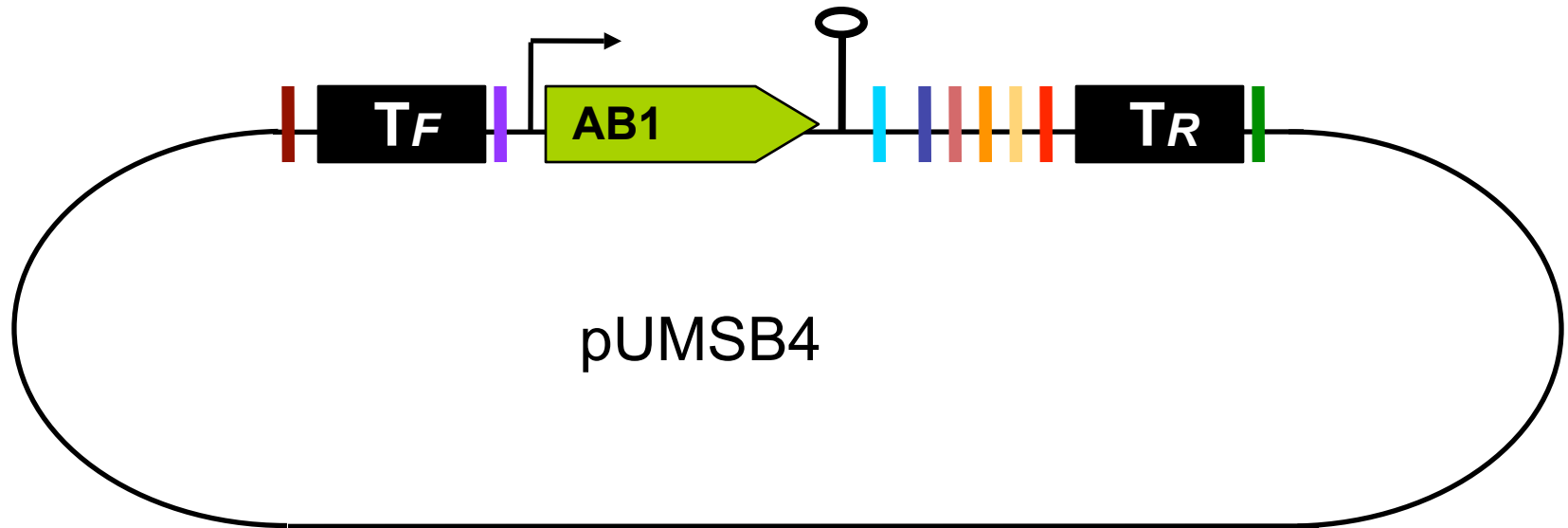


Polylinker: **EcoRI**, **SacI**, **NotI**, **NdeI**, **ClaI**, **BamHI**, **Sall**, **XhoI**, **HindIII**

TR - Rear end of target gene



# Landing Pad Construction

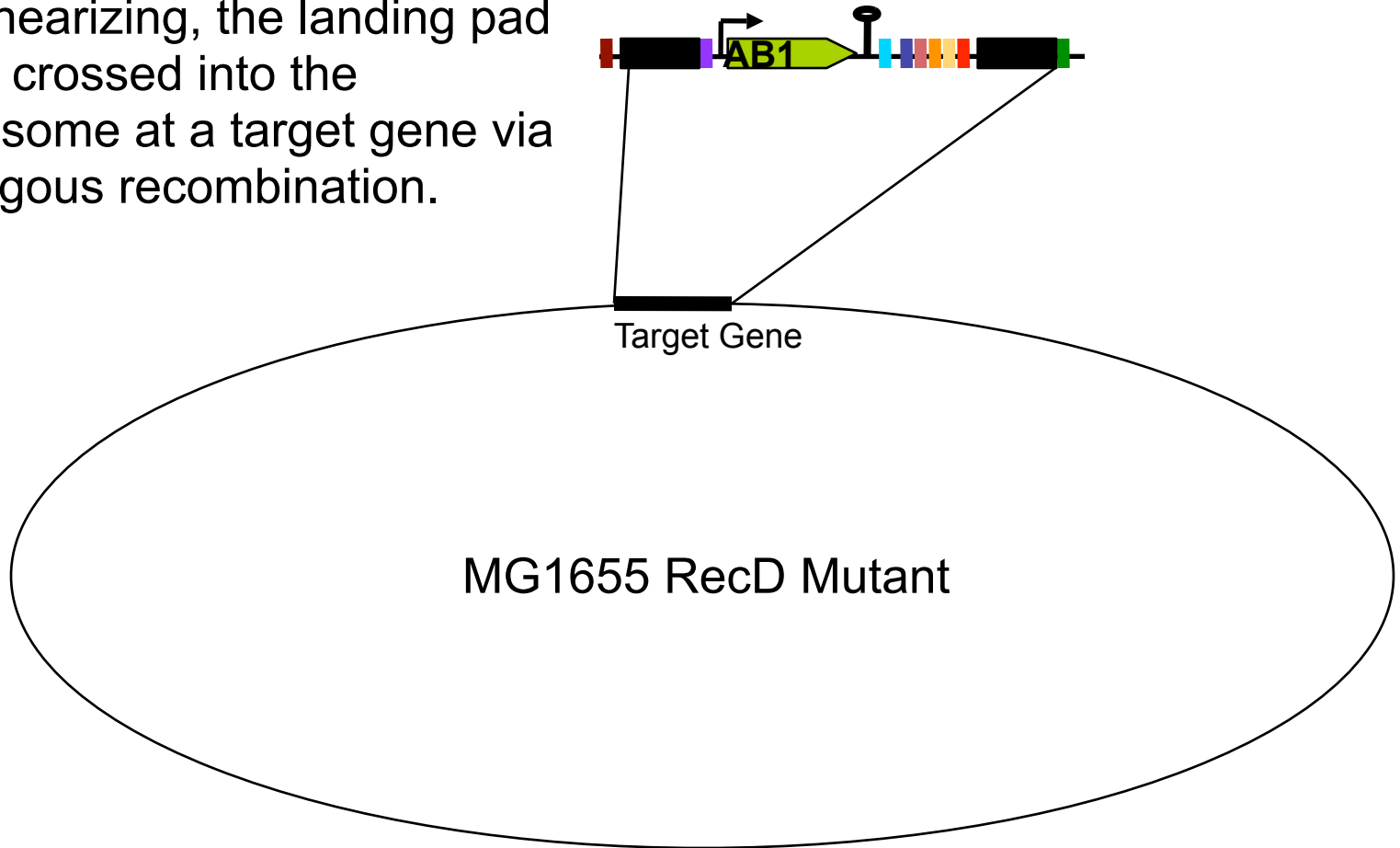


Polylinker: EcoRI, SacI, NotI, NdeI, ClaI, BamHI, SalI, XhoI, HindIII

AB1 - Antibiotic Resistance

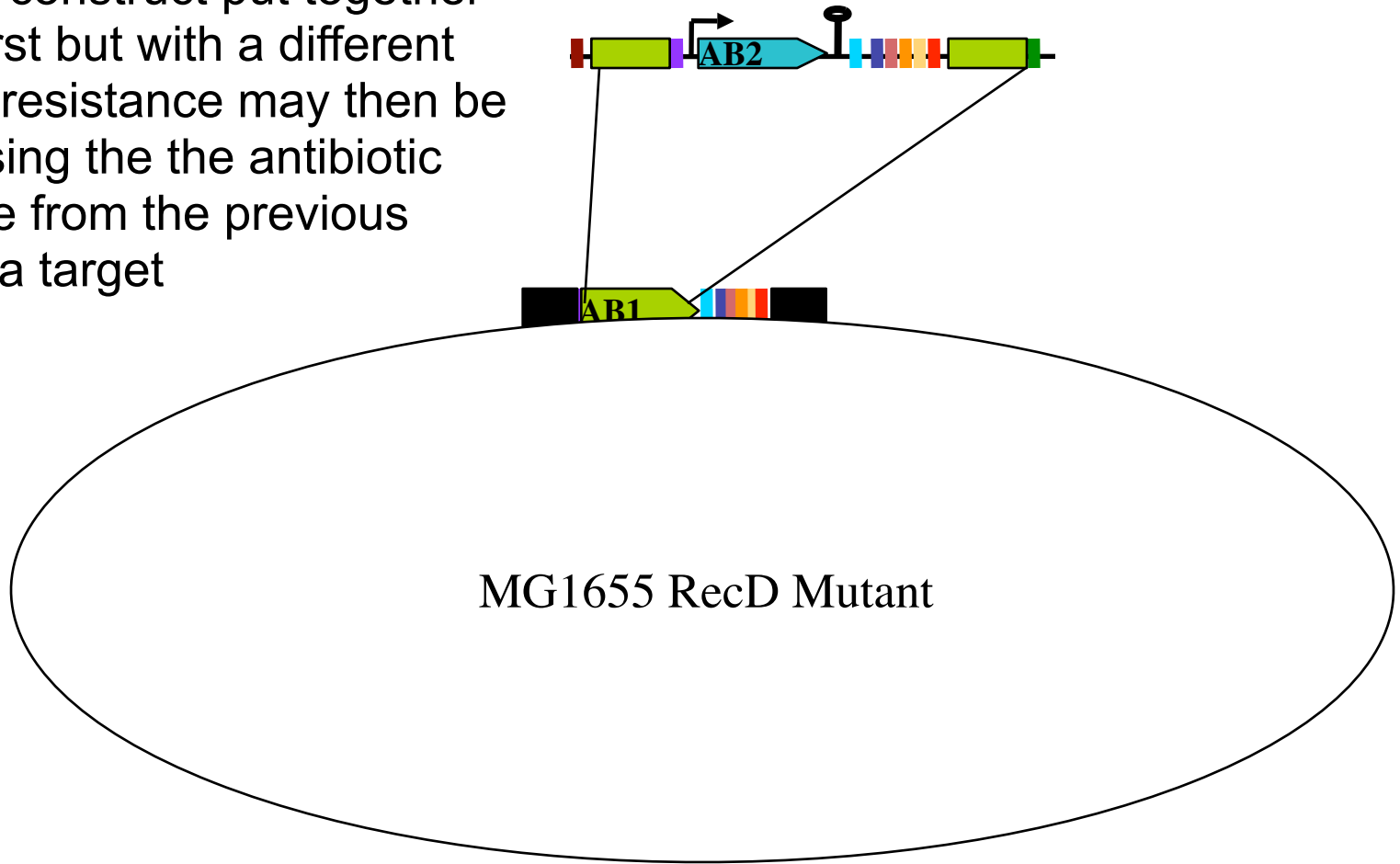
# Homologous Recombination onto the Chromosome

Upon linearizing, the landing pad may be crossed into the chromosome at a target gene via homologous recombination.



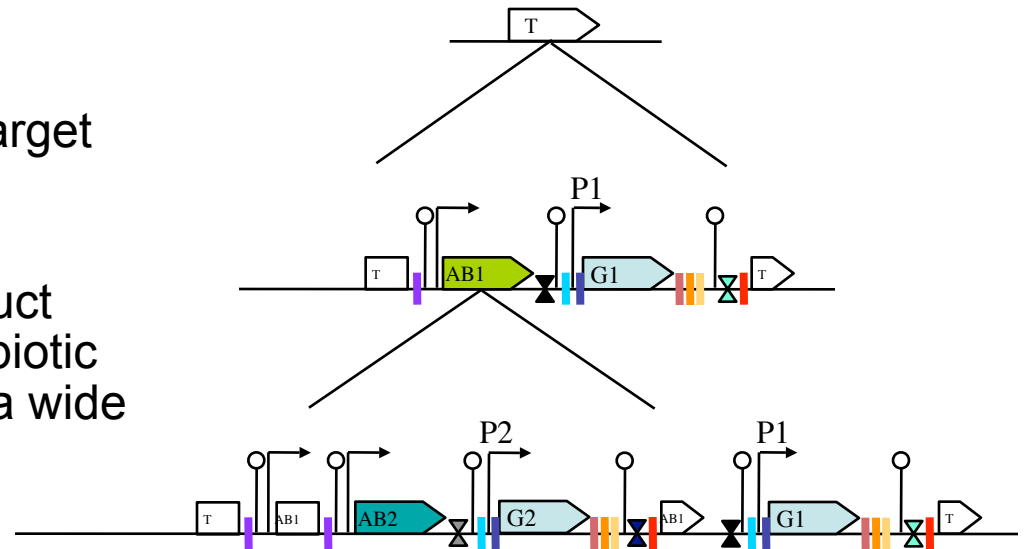
# Homologous Recombination onto the Chromosome

A second construct put together like the first but with a different antibiotic resistance may then be nested using the the antibiotic resistance from the previous round as a target



# Unique Features

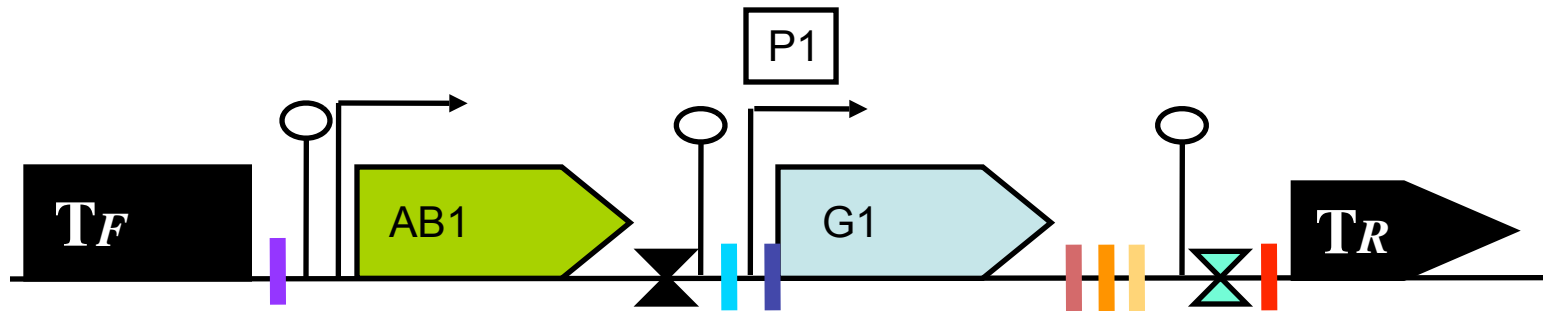
- Nesting
  - AB1 may be used as a target for future rounds of recombination.
  - By targeting AB1, construct essentially switches antibiotic sensitivities allowing for a wide range of constructs.



- Bar Codes (X)

- Unique 21 bp sequences placed at the end of each landing pad
- Allows for easy sequencing to check fidelity of recombinant techniques

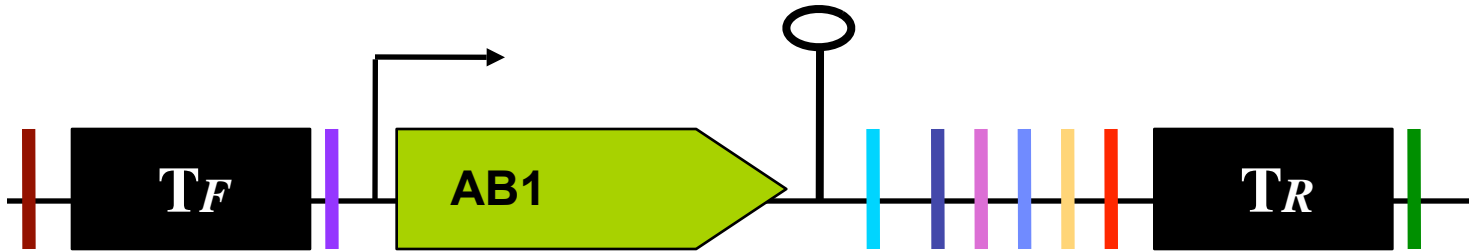
# Custom Construction



- The design allows the experimentalist to mix and match custom promoter / gene / terminator combinations.
- A series of extra restriction sites within the polylinker gives flexibility to those parts which may be already constructed.



# BioBrick Compatibility



- To make the current design BioBrick compatible, we reverse engineered a “patch” between the **NotI** and **BamHI** which contains both the **XbaI** and a **SpeI** site.
- Thus BioBricks may be built to designers desire as they normally would and then transferred into the landing pad.

# Tuning Kinetic Order Through The Use Of Repressor Sinks

# What is Kinetic Order?

Consider:

$$\frac{dy}{dx} = k_1 \frac{x^n}{k_2 + x^n}$$

# What is Kinetic Order?

Consider:

$$\frac{d[\text{RNA}]}{dt} = k_1 \frac{[\text{PIG}]^n}{k_2 + [\text{PIG}]^n}$$

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Hill coefficient  
↓



# What is Kinetic Order?

Consider:

$$\frac{d[\text{RNA}]}{dt} = \frac{k_1 [\text{PIG}]^n}{k_2 + [\text{PIG}]^n}$$

Hill coefficient  
↓

(Hill coefficient describes kinetic order)

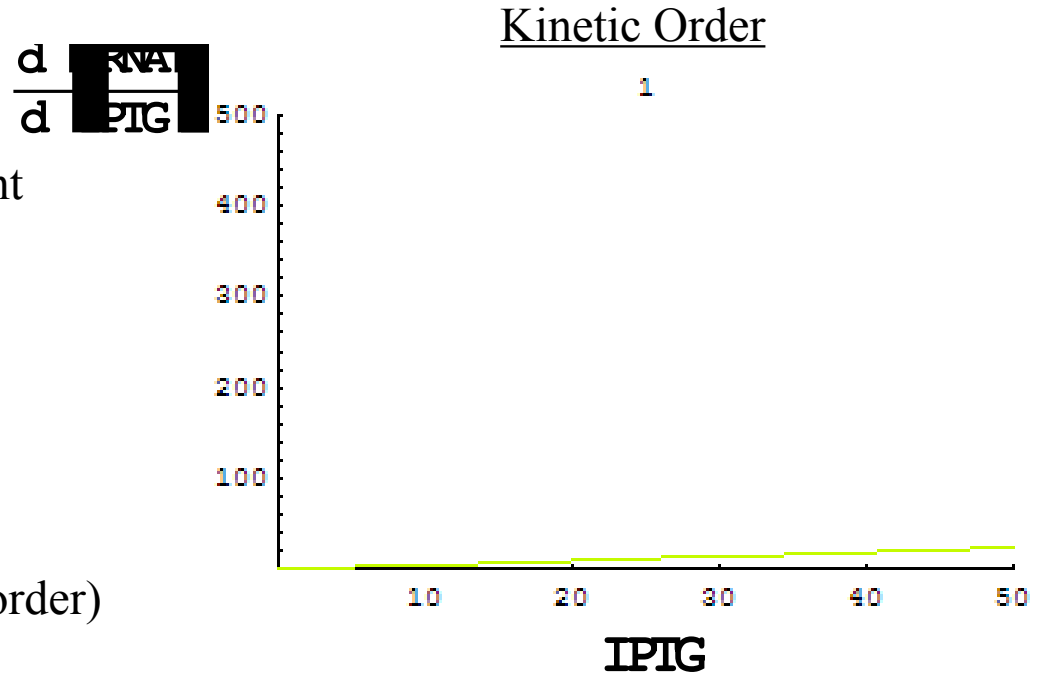
# What is Kinetic Order?

Consider:

$$\frac{d[\text{RNA}]}{dt} = \frac{k_1 [\text{IPTG}]^n}{k_2 + [\text{IPTG}]^n}$$

Hill coefficient  
↓  
 $n$

(Hill coefficient describes kinetic order)

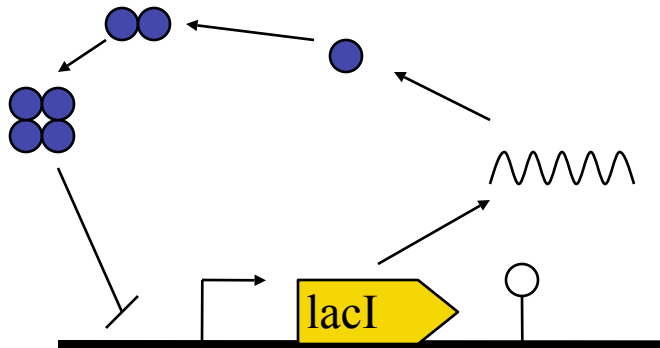


# Why Increase Kinetic Order?

- Increase Switch-like Behavior
- Increase Oscillations

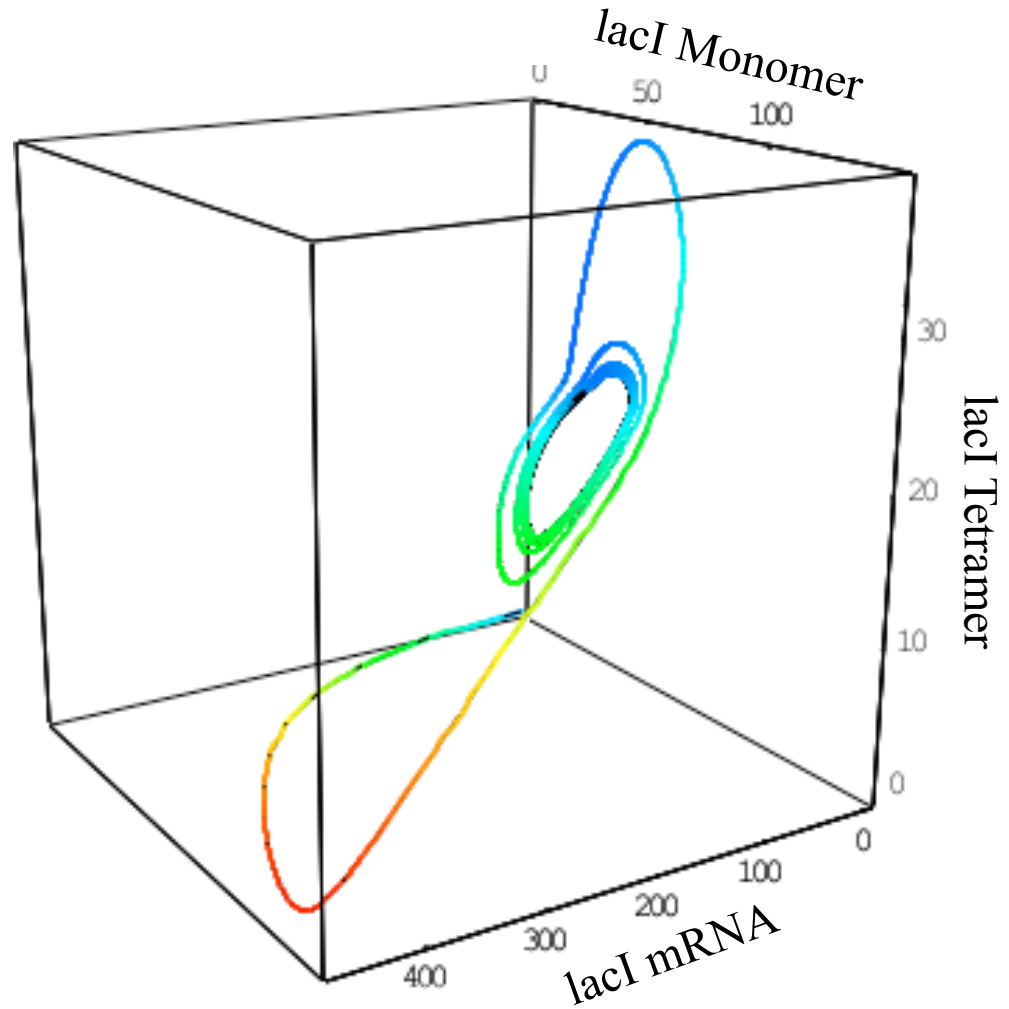
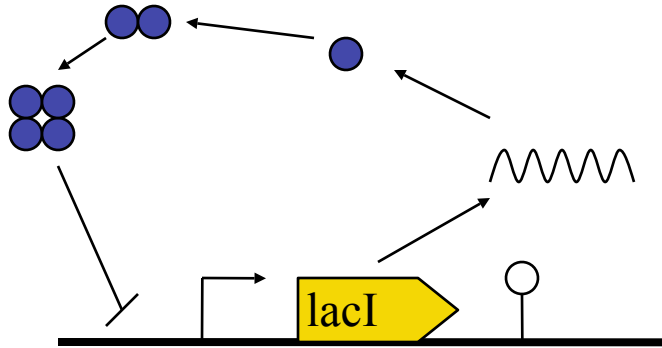
# Why Increase Kinetic Order?

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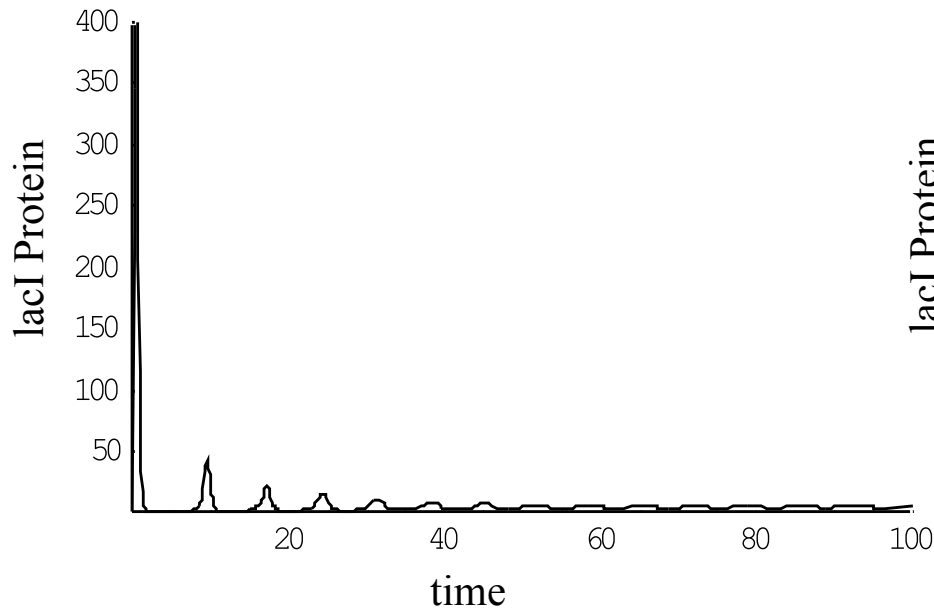
# Why Increase Kinetic Order?

- Increase Switch-like Behavior
- Increase Oscillations

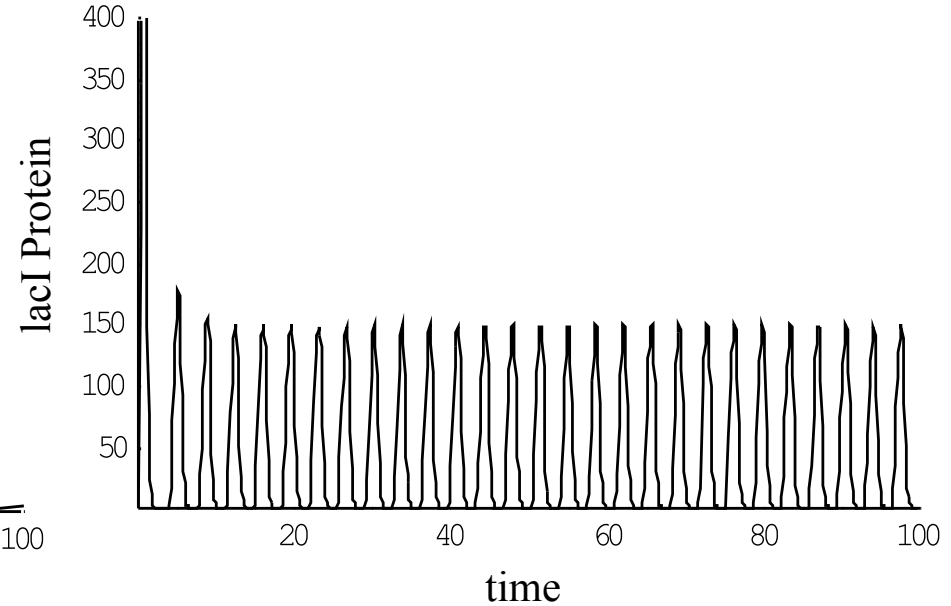


# lacI Oscillator: Sinks vs. No Sinks

# of Sinks: 0



# of Sinks: 10



# The Model

## Definitions

$S_c$  = Chromosomal sink,

$S_s$  = Sink sites,

$S_z$  = *LacZYA* promoter,

$IP$  = IPTG,

$m$  = Number of sink sites,

$M$  = Number of chromosomal sink sites,

$R_{Sc}$  = Repressor bound to non-specific binding site

$R_{Ss}$  = Repressor bound to sink site,

$R_{Sz}$  = Repressor bound to promoter

$R_{IP}$  = Repressor bound to IPTG (inactivated)

$R_f$  = Free Repressor,

$R_t$  = Total Repressor (constant)

$R$  = Active Repressor (all repressor in system not bound to IPTG)

$k_c = k_{eq}$  for Non-specific binding site,

$k_s = k_{eq}$  for sink site,

$k_z = k_{eq}$  for promoter

## Assumptions

① Constant amount of total repressor,

② Quasi-equilibria repressor-IPTG,

Repressor-Sink, Repressor-promoter,

③  $k_s = k_z$ ,

④  $k_c$  is very small ( $10^{-6}$ ),  $R_f \ll 1$ ,  $k_c \cdot M \sim 1$ , so  $\frac{k_c M}{1 + k_c R_f} \sim 1$

## The Model

$R_t = R_{IP} + R_{Sc} + R_{Ss} + R_{Sz} + R_f$

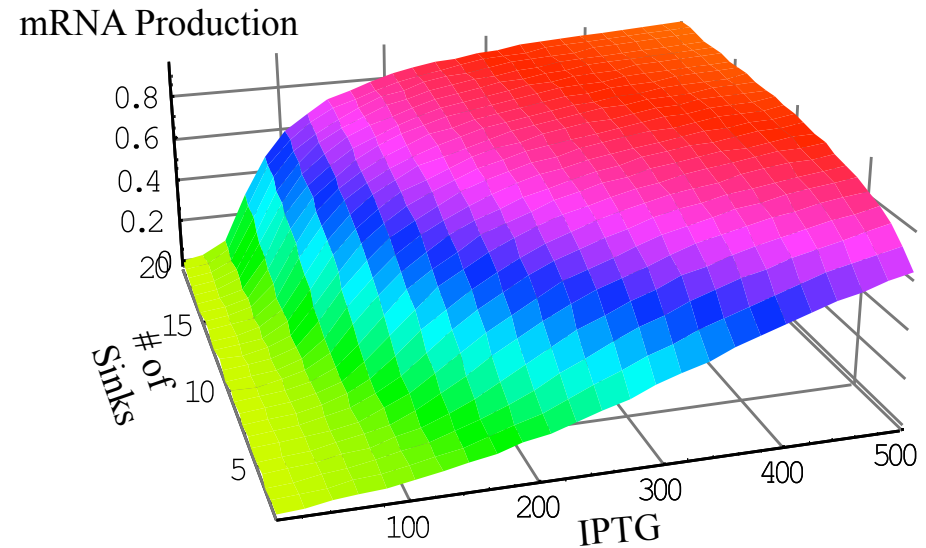
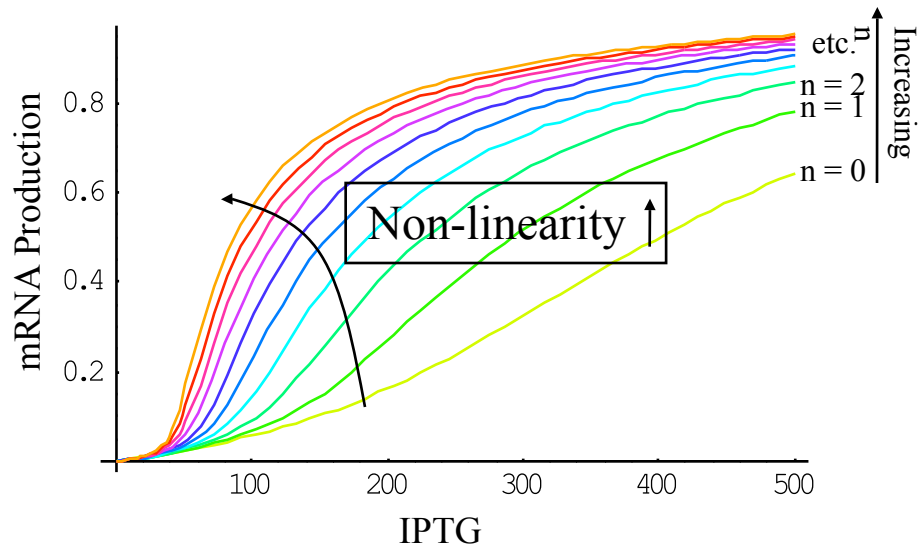
$M = \frac{R_{Sc}}{k_c R_f} + \frac{R_{Ss}}{k_s R_f} + \frac{R_{Sz}}{k_z R_f} + \frac{R_f}{k_c R_f}$

$m = \frac{R_{Sc}}{k_c R_f} + \frac{R_{Ss}}{k_s R_f} + \frac{R_{Sz}}{k_z R_f} + \frac{R_f}{k_c R_f}$

$1 = \frac{R_t}{R_f} + \frac{k_c M}{1 + k_c R_f} + \frac{k_s m}{1 + k_s R_f} + \frac{k_z}{1 + k_z R_f}$

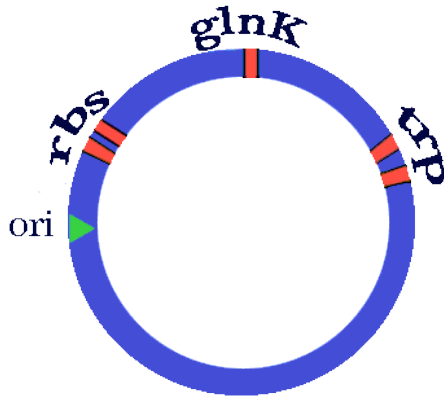
$$\frac{R_t}{1 + IP \cdot k_{IP}} = R_f \left( 1 + \frac{k_c M}{1 + k_c R_f} + \frac{k_s m}{1 + k_s R_f} + \frac{k_z}{1 + k_z R_f} \right)$$

# Increase in Kinetic Order with Sinks



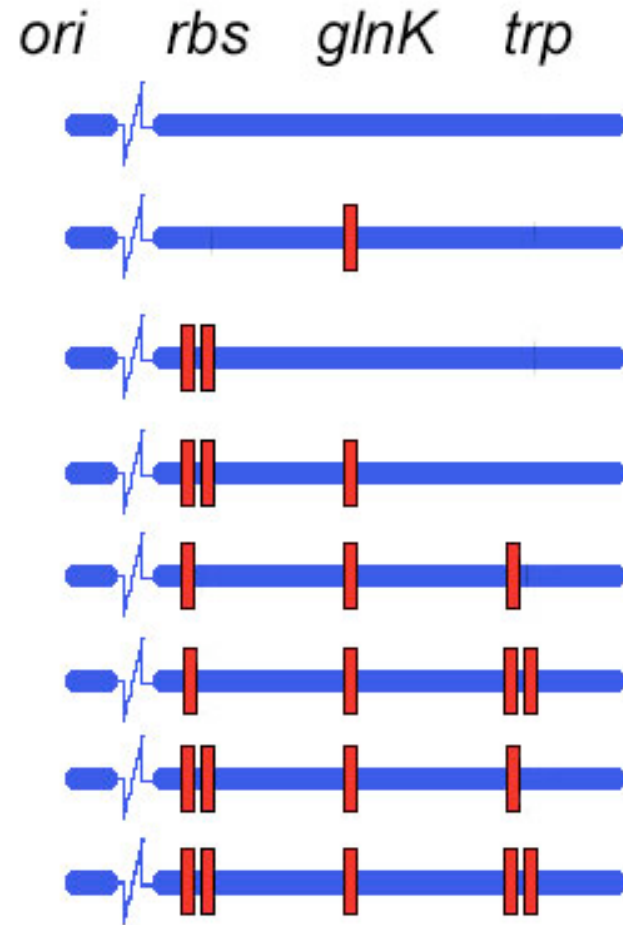


# Experimental Manipulation of Lac Operon Kinetic Order

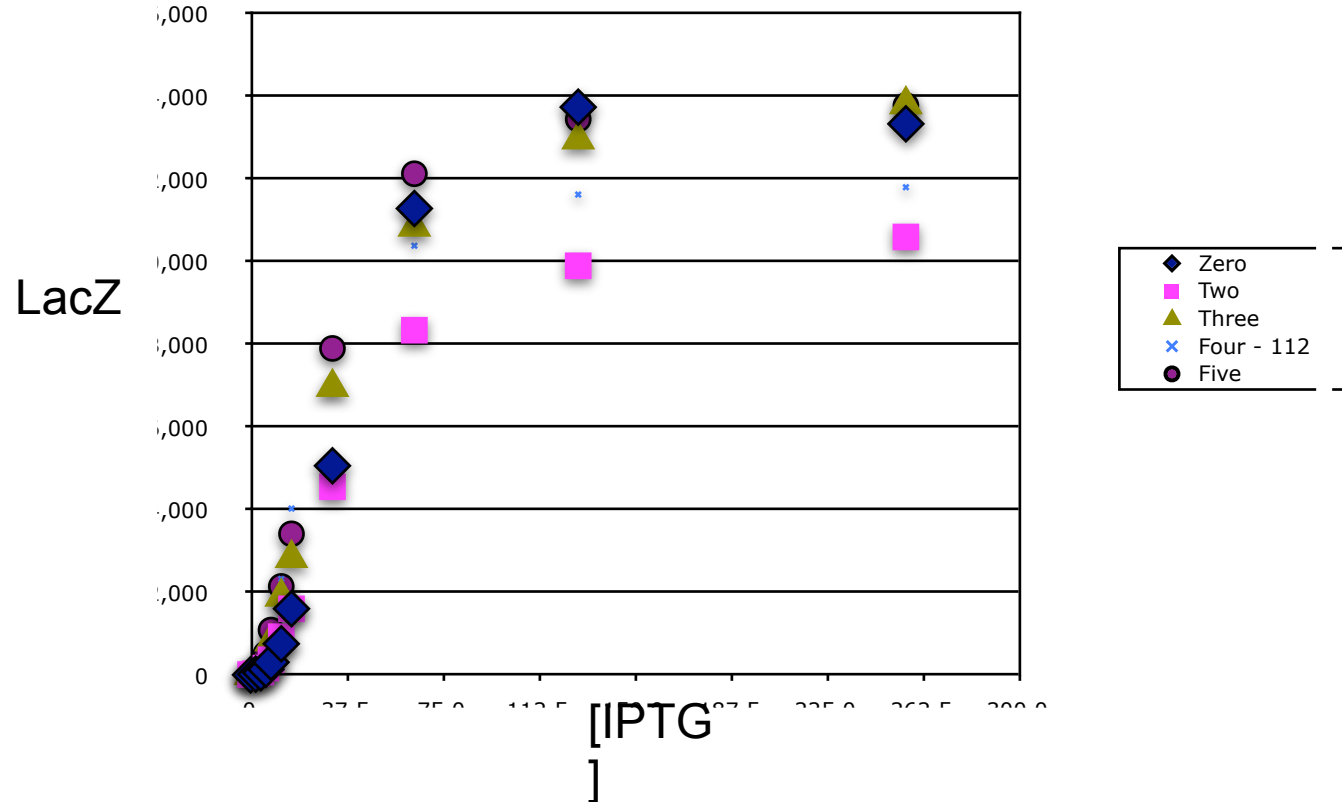


# Sinks on Chromosome

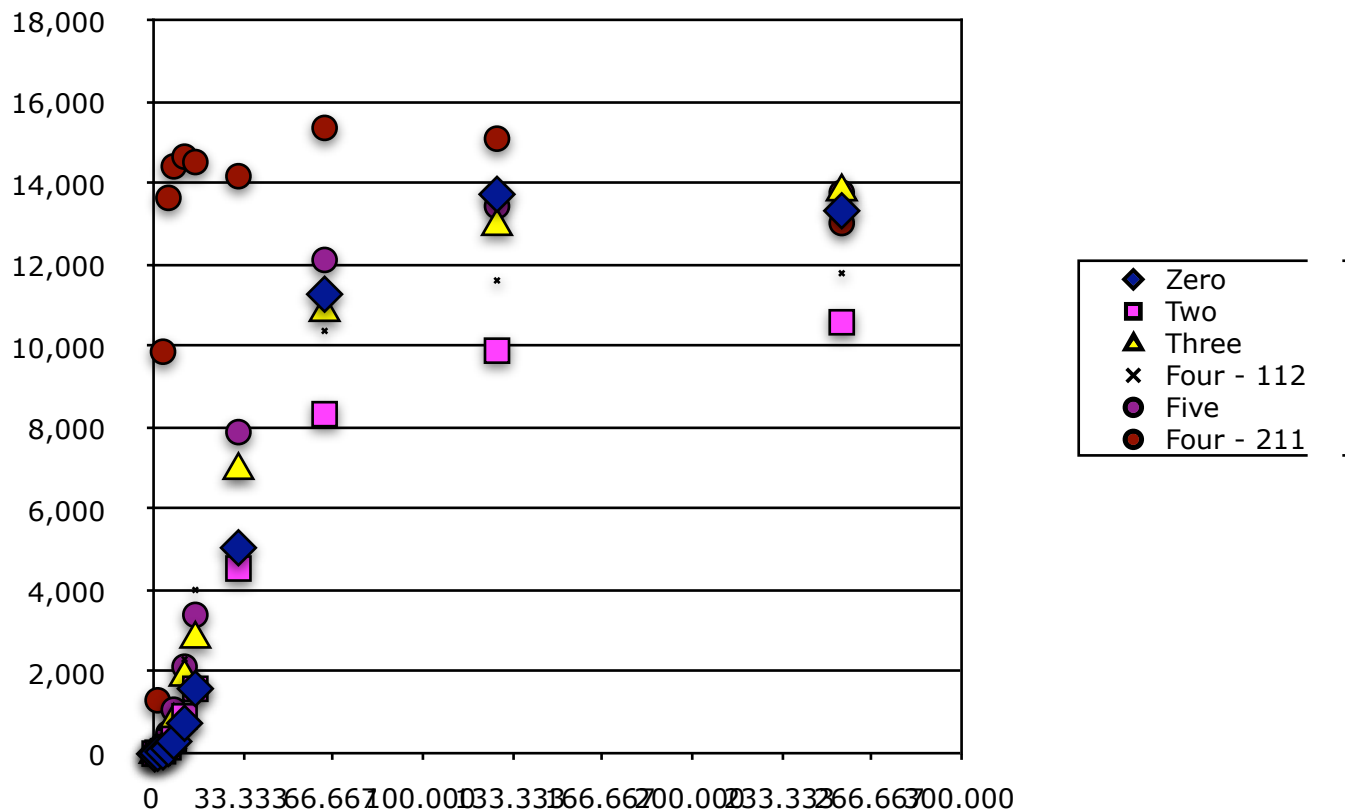
- $O_p$  (red bars) inserted into chromosome using standard recombination methods
  - Landing pads
  - P1 vir transduction



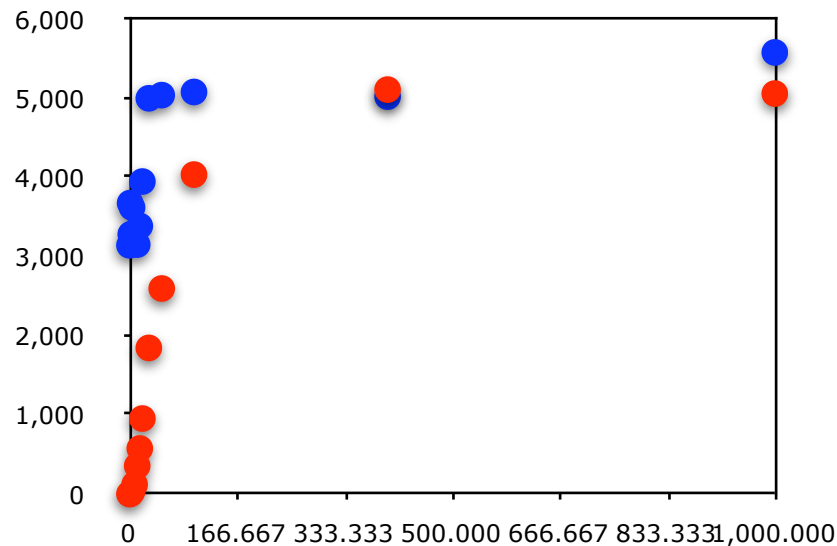
# Increasing Number of Sinks on Chromosome Raises



# Strain with 4 Sinks Shows Switch-like Behavior

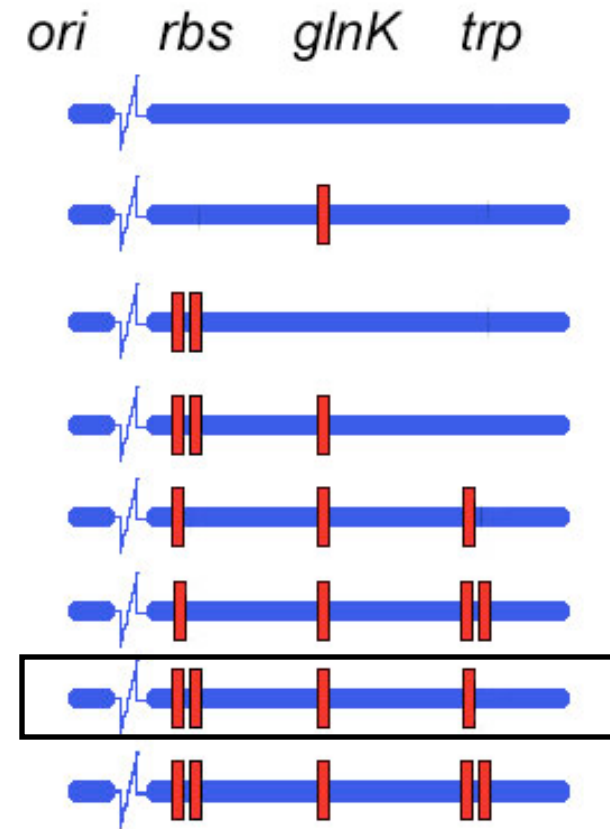
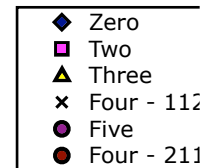
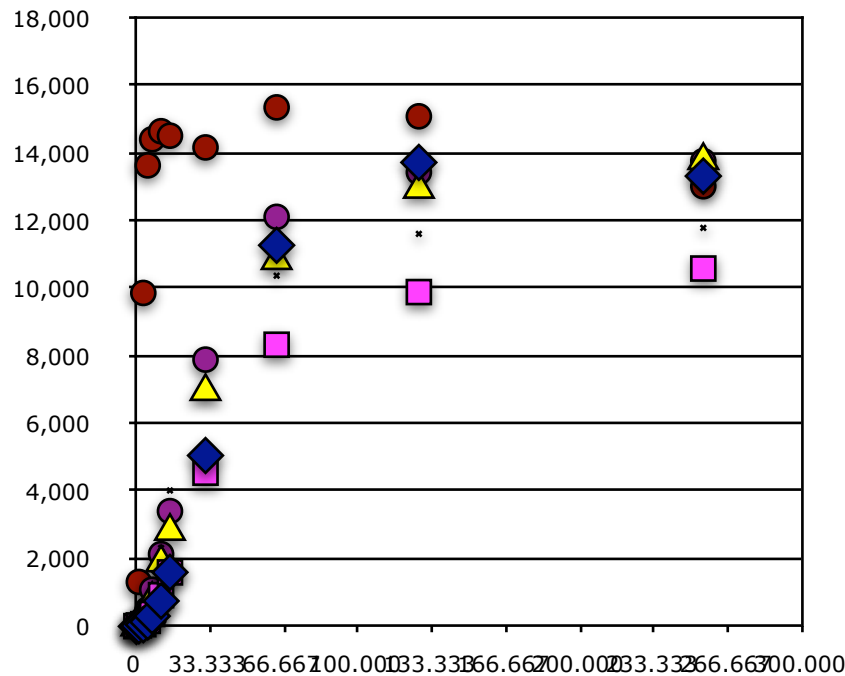


# Sinks on Plasmid Raise Basal Level of LacZ Expression



— Without sinks  
With sinks

# Strain with 4 Sinks Shows Switch-like Behavior



# Discussion

- Ongoing Work
  - Rebuild strains in different ways to see if we can repeat results
  - Examine effects of position and proximity
  - Examine effects of more weakly binding operators ( $O_1$ ,  $O_2$ , etc.)
- Adding sinks to chromosome may allow for “tuning” of kinetic order

# Overall Conclusion

- Work presented here was accomplished in a team effort
- Thanks go out to the many people who helped on the team's projects
- Go Blue!



# Overall Conclusion

- Work presented here was accomplished in a team effort
- Thanks go out to the many people who helped on the team's projects
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