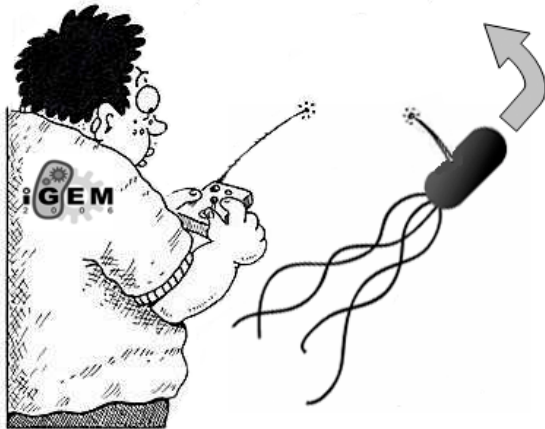


# Remote Control of Bacterial Chemotaxis



UCSF iGEM Team 2006



Patrick Visperas

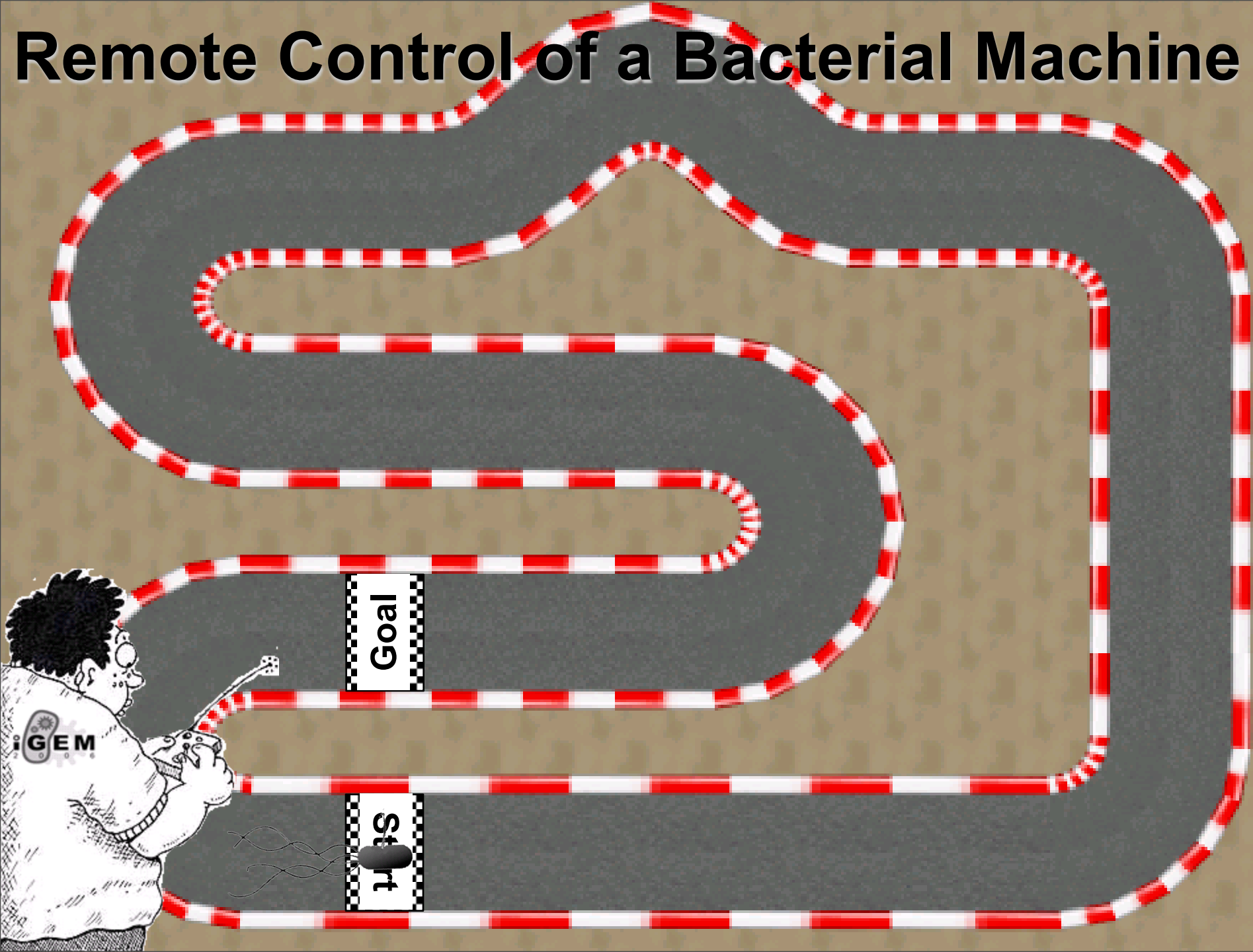
Matthew Eames

Eli Groban

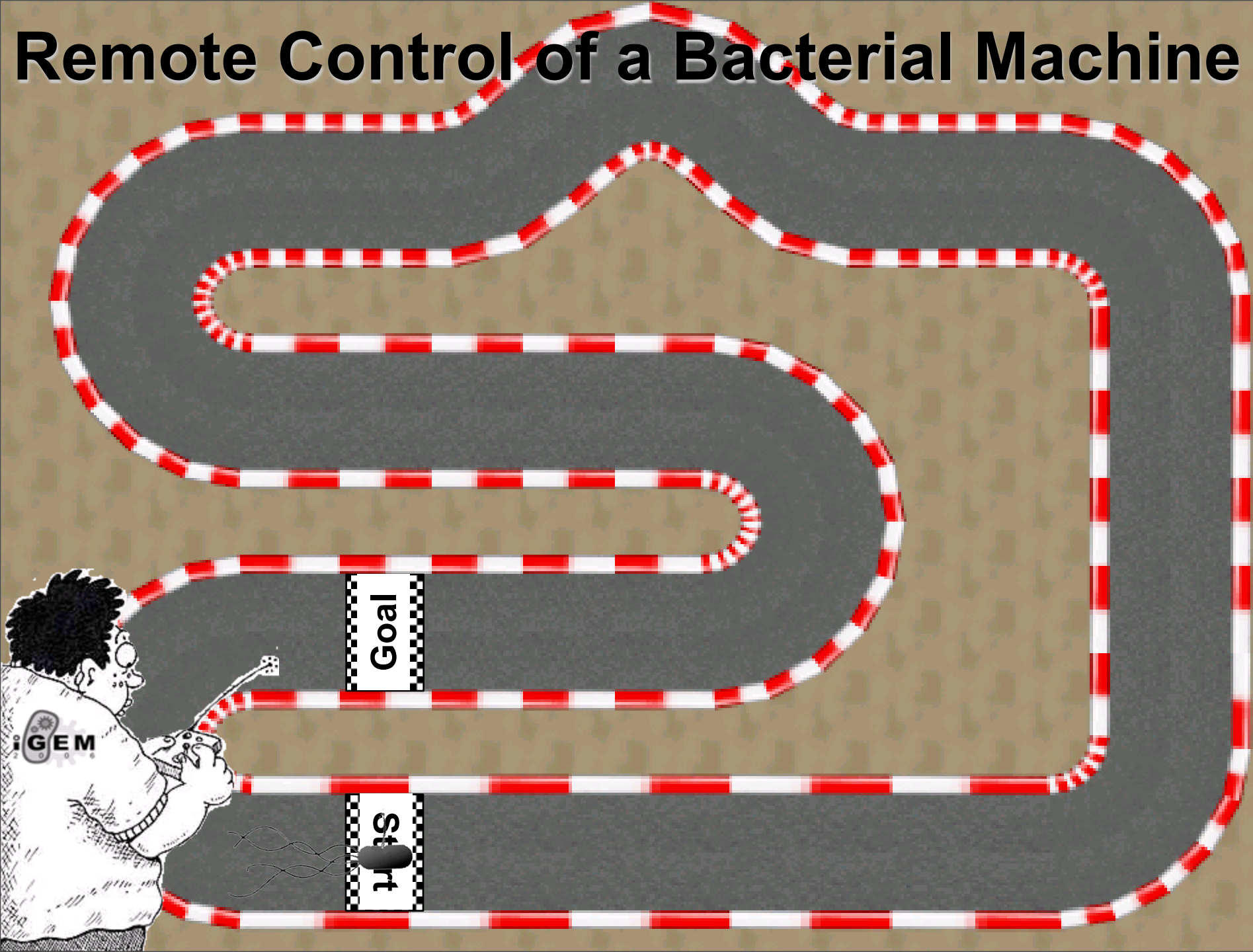
Ala Trusina

Christopher Voigt, Tanja Kortemme, Chao Tang, Chris Rao

# Remote Control of a Bacterial Machine

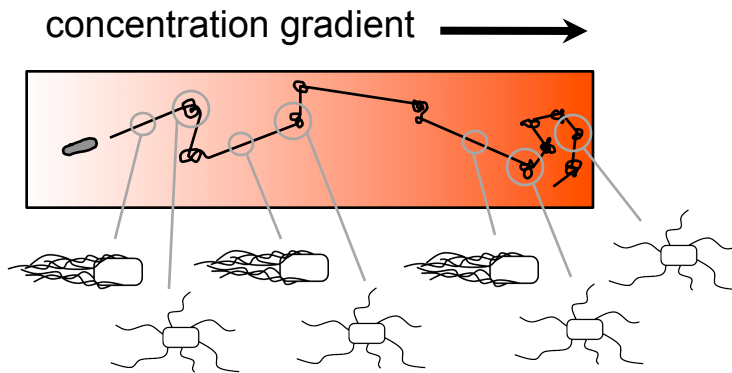
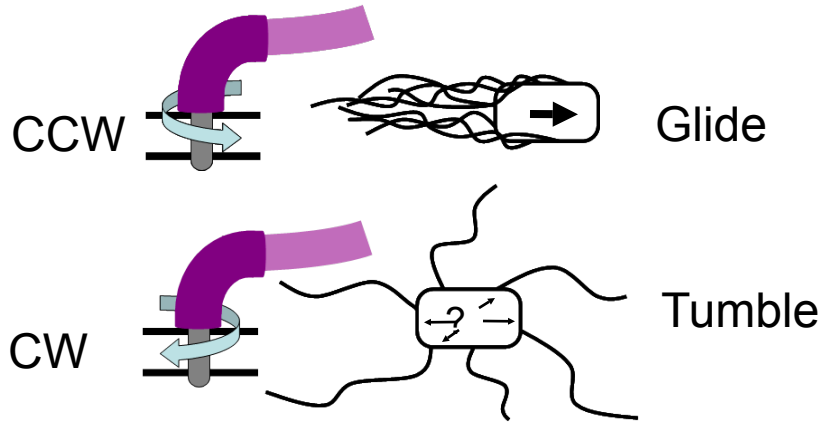


# Remote Control of a Bacterial Machine

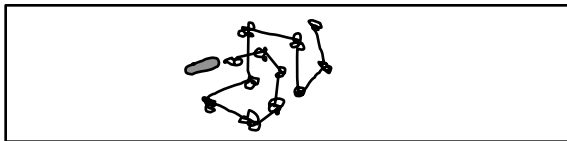


# How Bacterial Chemotaxis Works

## Bacteria Swim Up Gradients

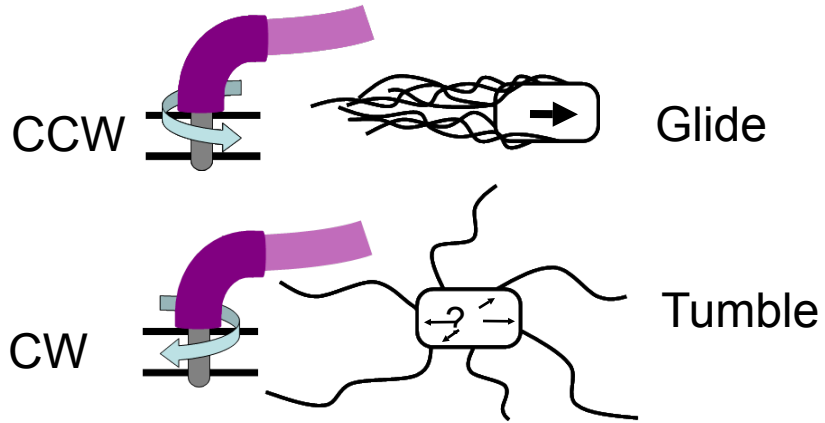


No gradient

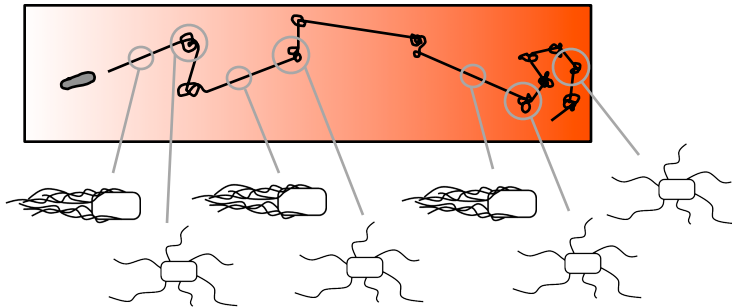


# How Bacterial Chemotaxis Works

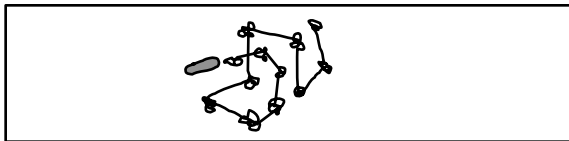
## Bacteria Swim Up Gradients



concentration gradient →

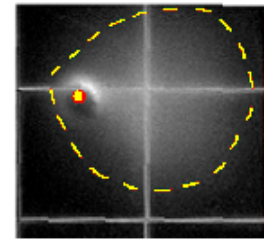
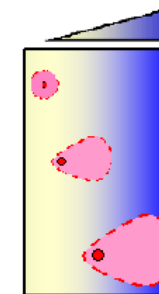
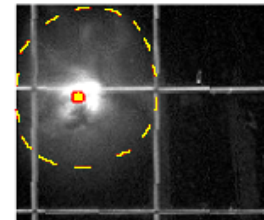
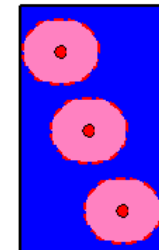
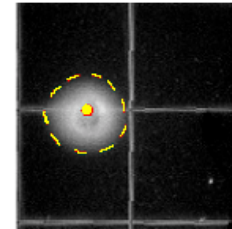
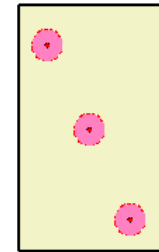


No gradient

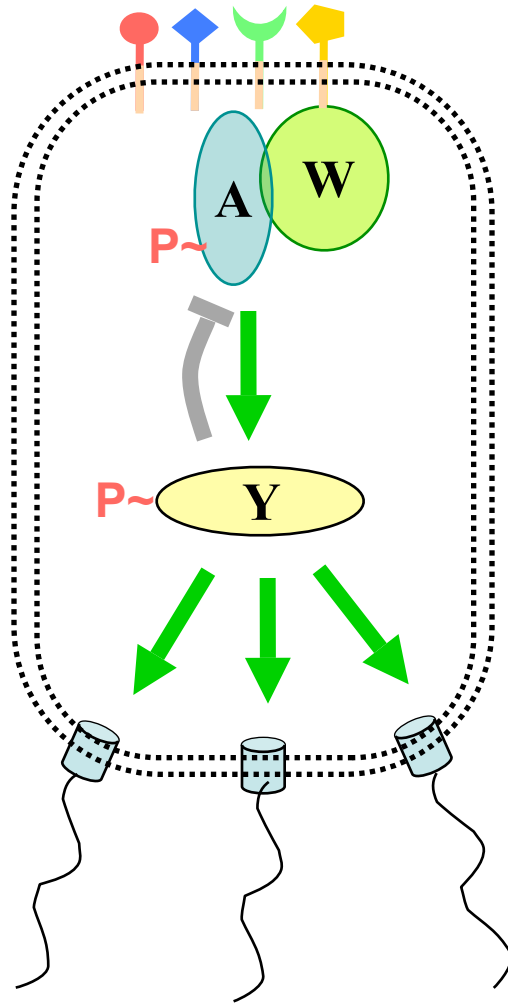


## How Chemotaxis is Observed

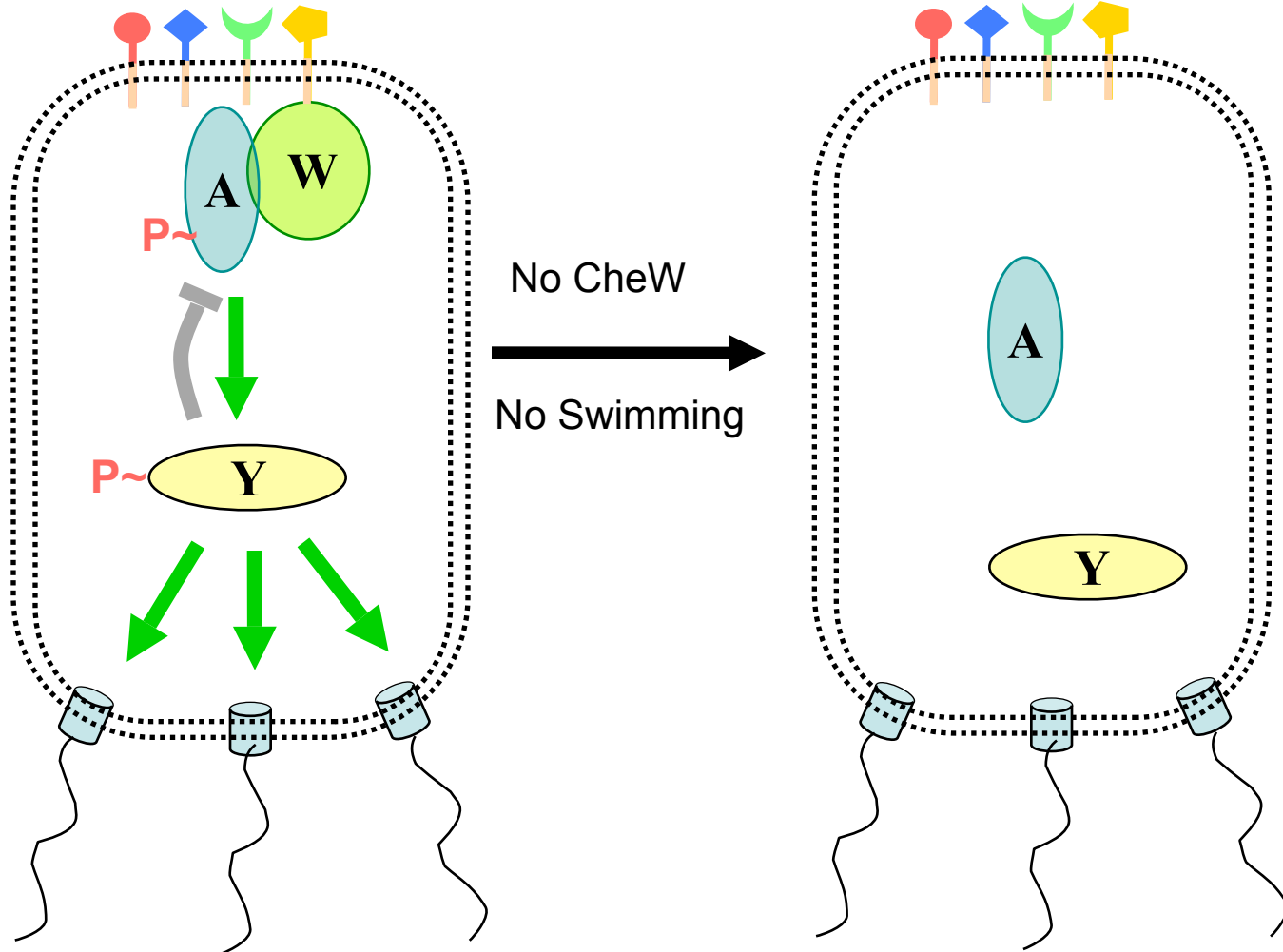
Goulian Motility Assay (U Penn)



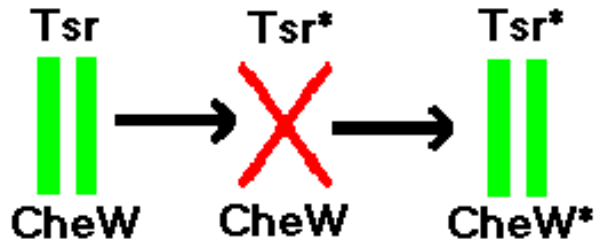
# The Regulatory Network



# The Regulatory Network

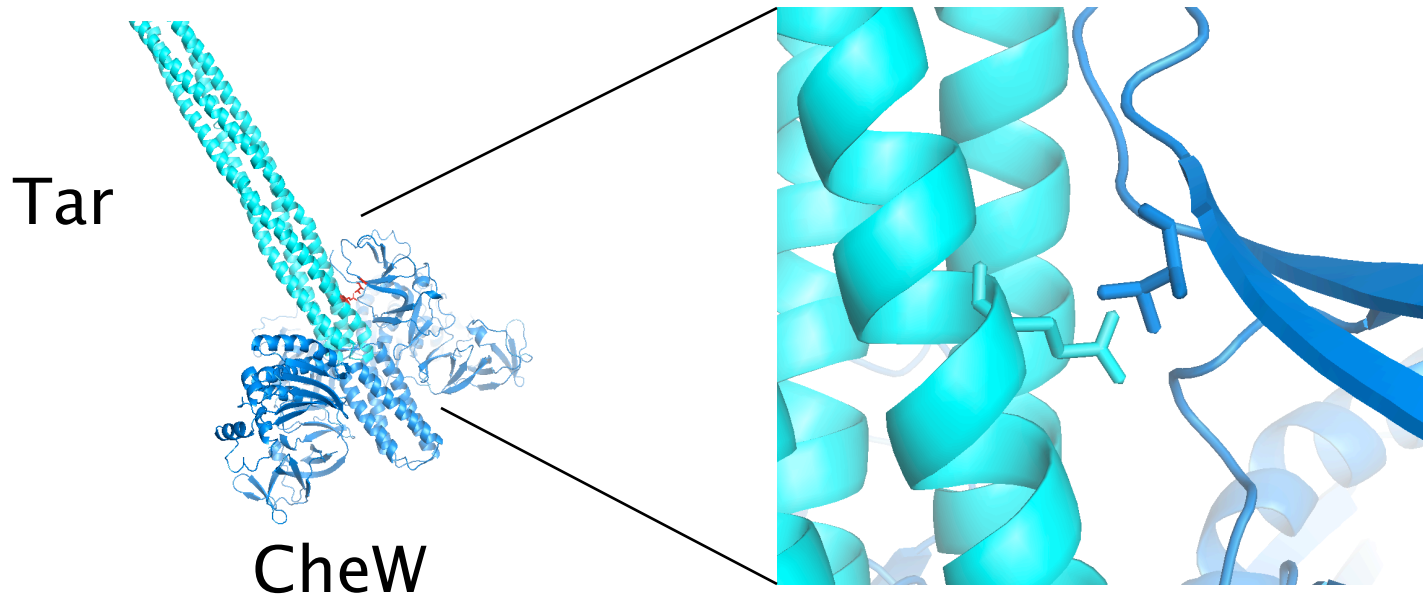
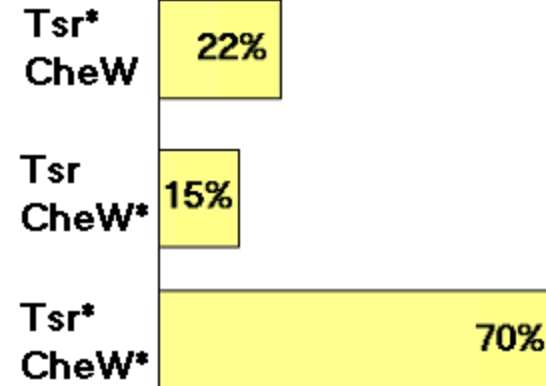


# Binding Partners



- Orthogonal binding pair Tsr-CheW (Liu et al, 1991)
- We mapped these mutations onto the Tar-CheW complex

Swarm rate (% of w.t.)

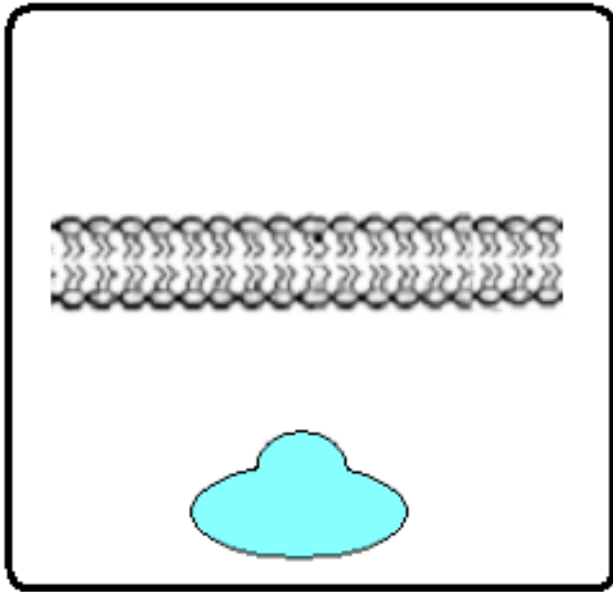
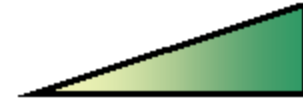




# Tsr mutation can be mapped to Tar

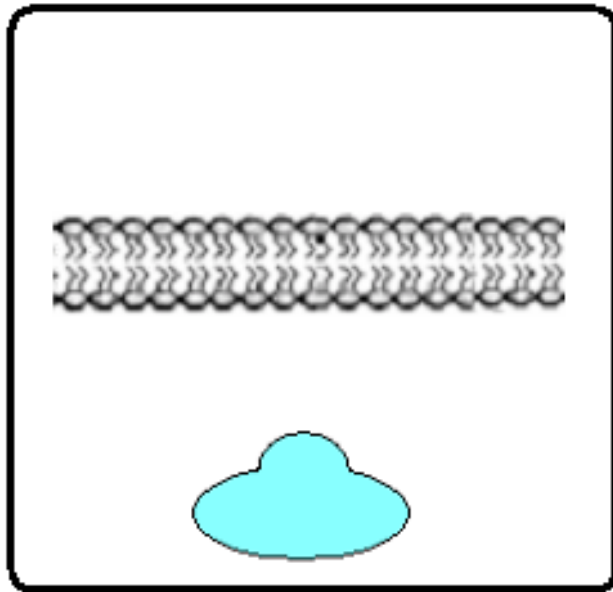
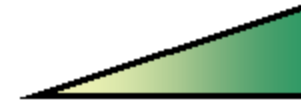
Gradient

Aspartate

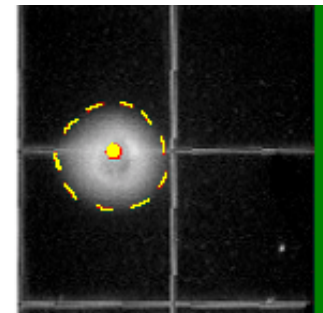


# Tsr mutation can be mapped to Tar

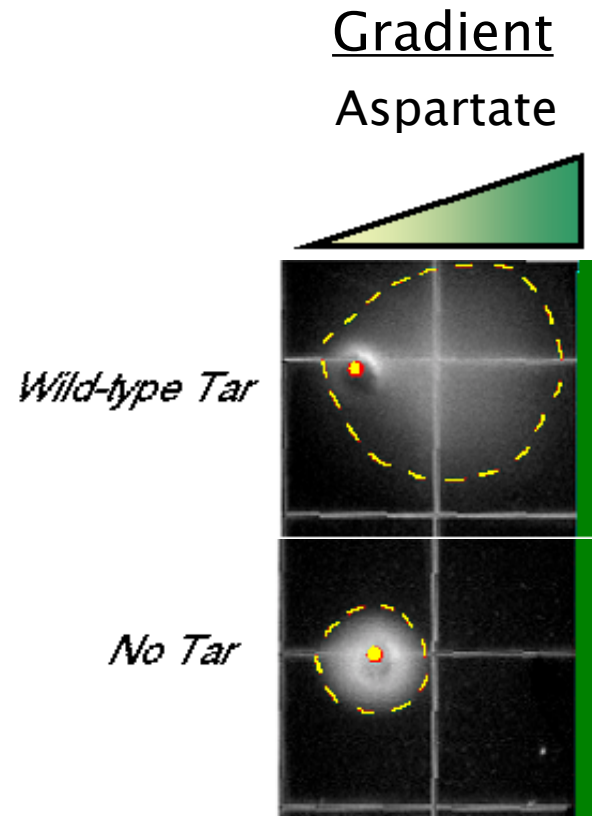
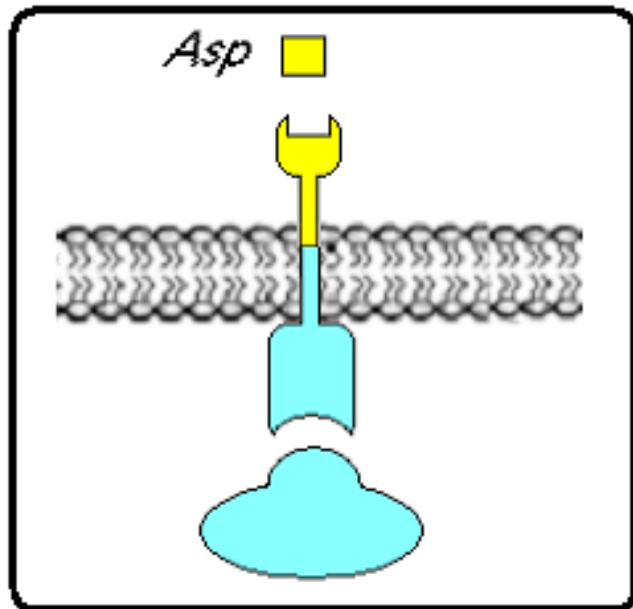
Gradient  
Aspartate



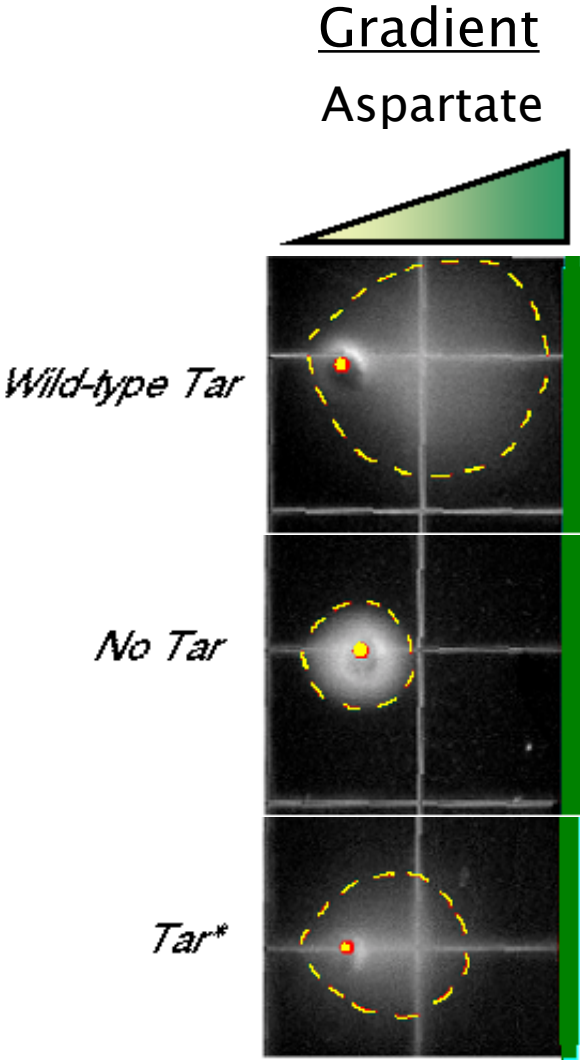
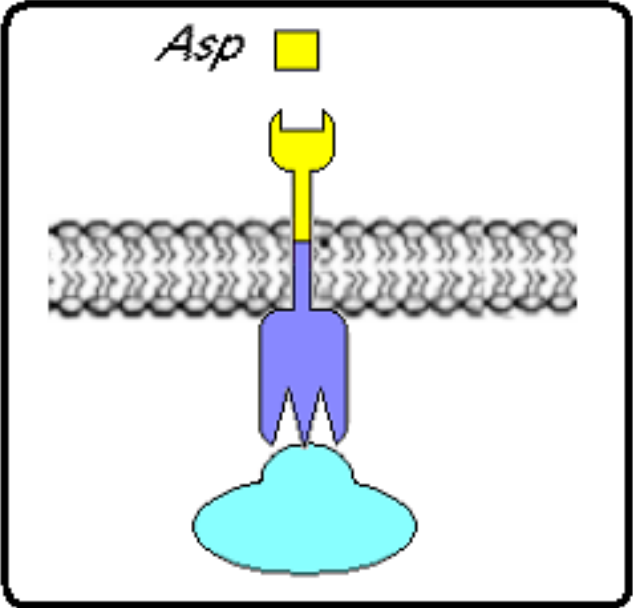
*No Tar*



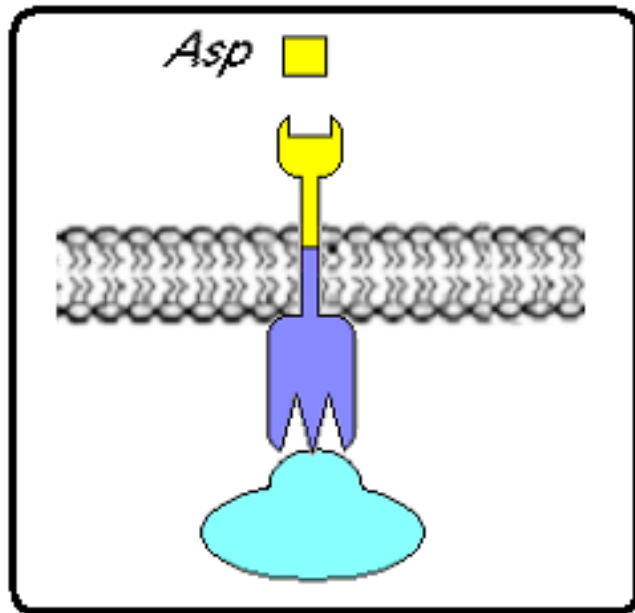
# Tsr mutation can be mapped to Tar



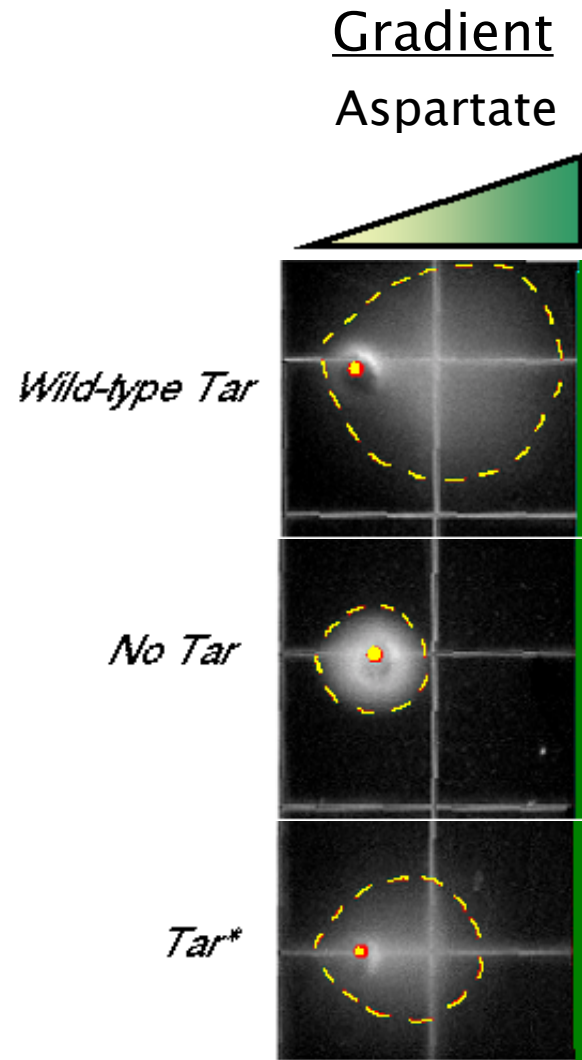
# Tsr mutation can be mapped to Tar



# Tsr mutation can be mapped to Tar

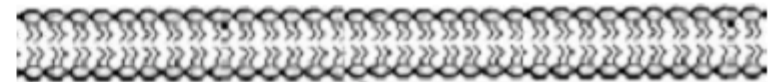
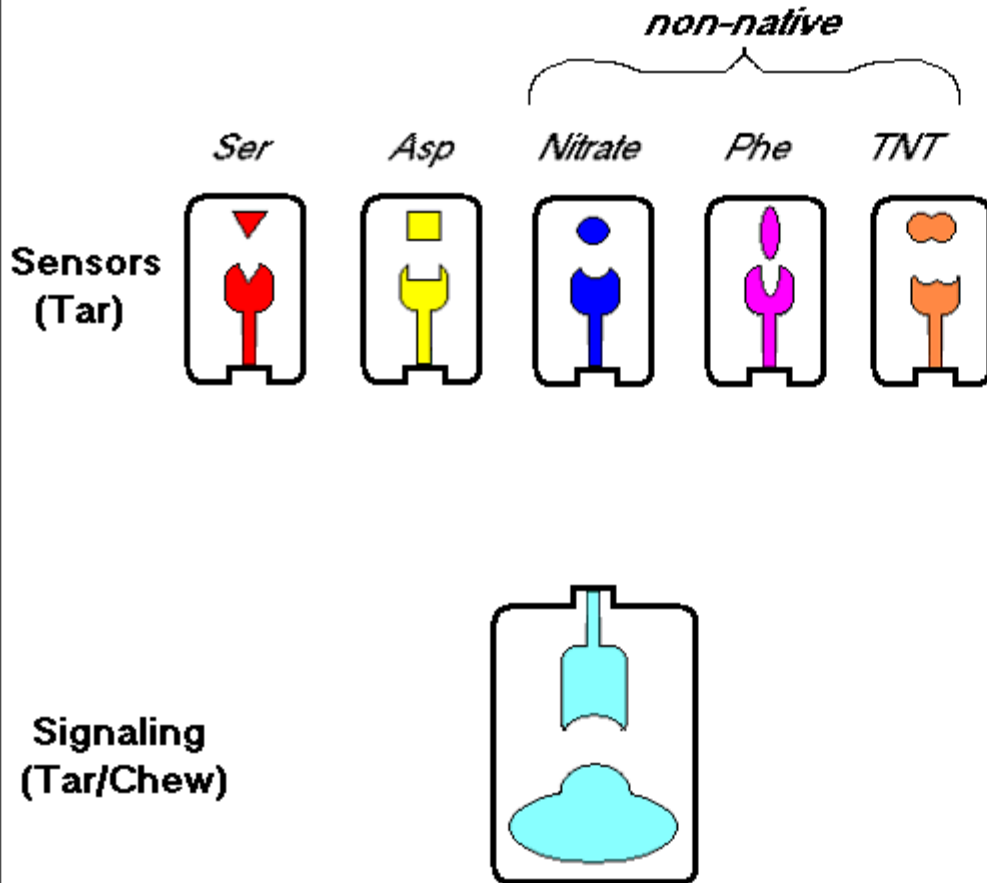


- point mutation in Tar\* reduces motility to approximately 40% that of wild-type



# Part Design

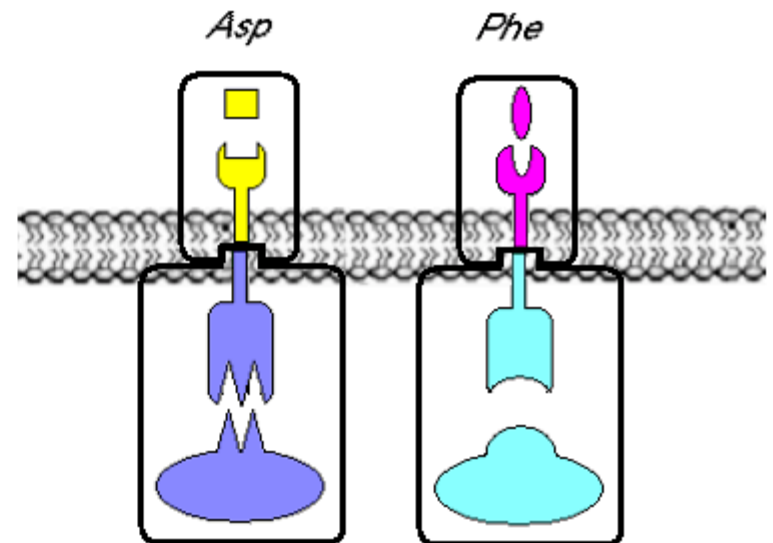
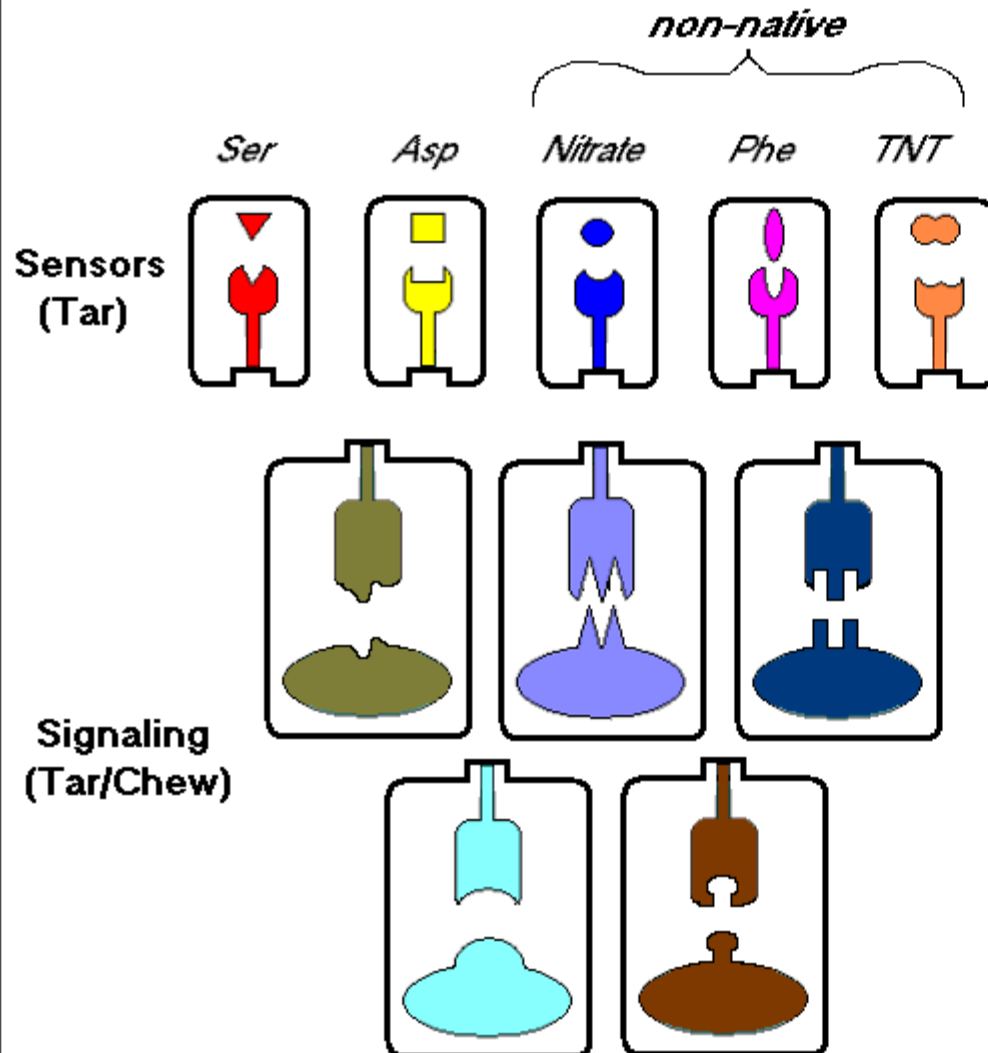
- New orthogonal signaling pairs



“Codon randomization” is used to reuse scaffolds without fear of recombination

# Part Design

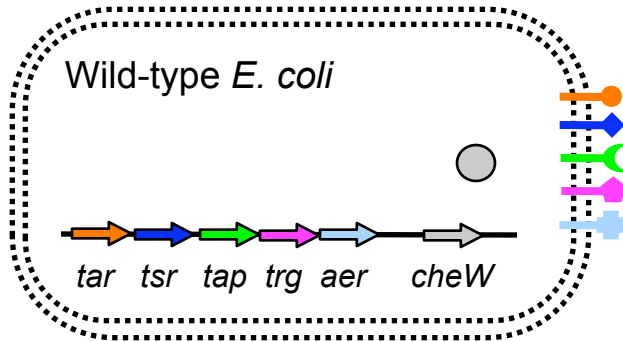
- New orthogonal signaling pairs



“Codon randomization” is used to reuse scaffolds without fear of recombination

# Chassis Engineering

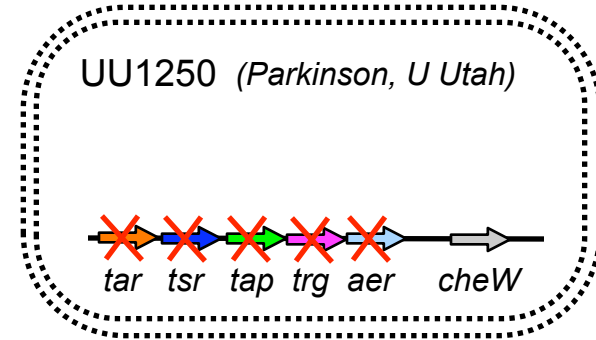
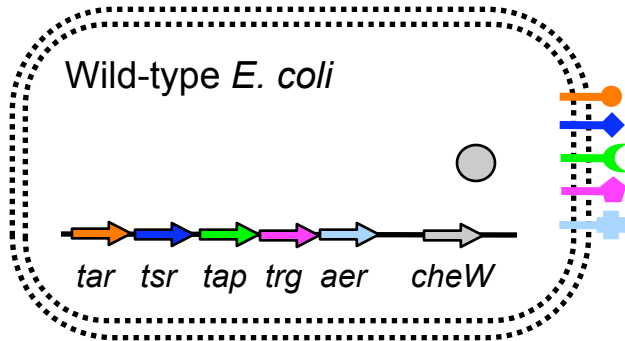
- Build *E. coli* strain lacking all sensors and CheW, but retaining the other chemotaxis proteins





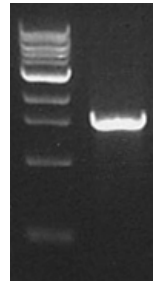
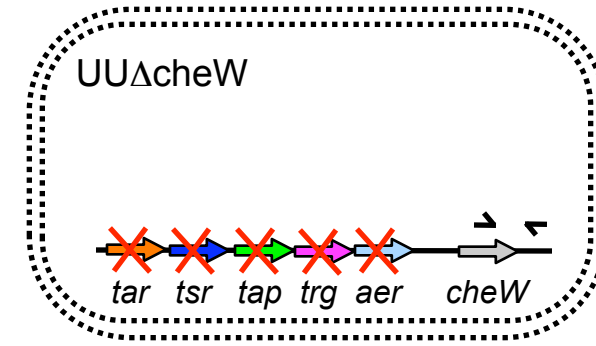
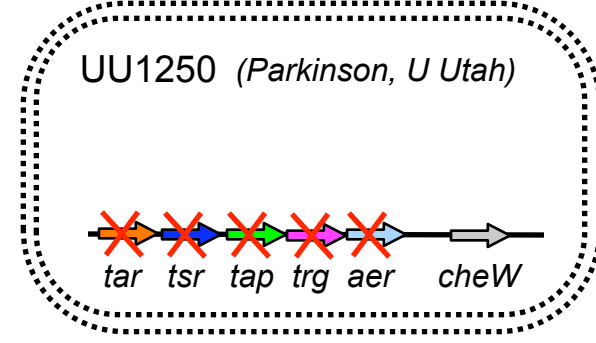
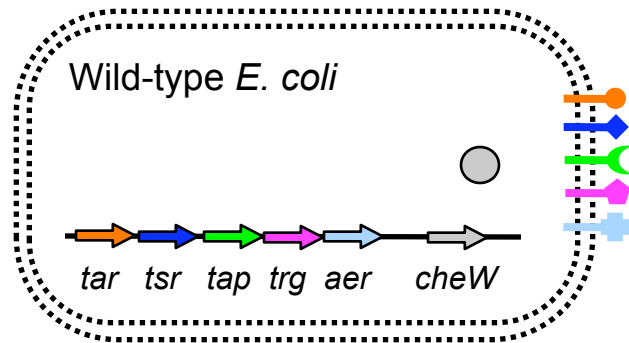
# Chassis Engineering

- Build *E. coli* strain lacking all sensors and CheW, but retaining the other chemotaxis proteins



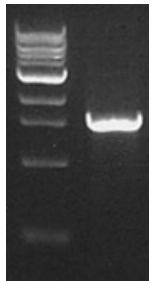
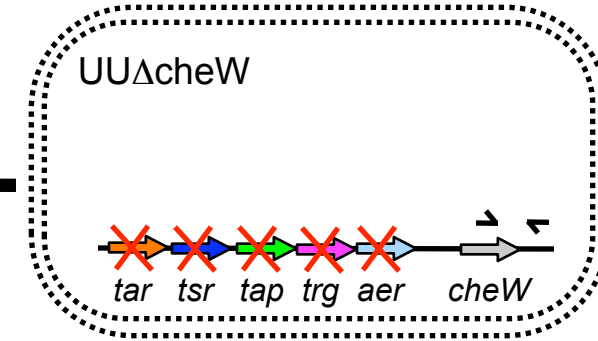
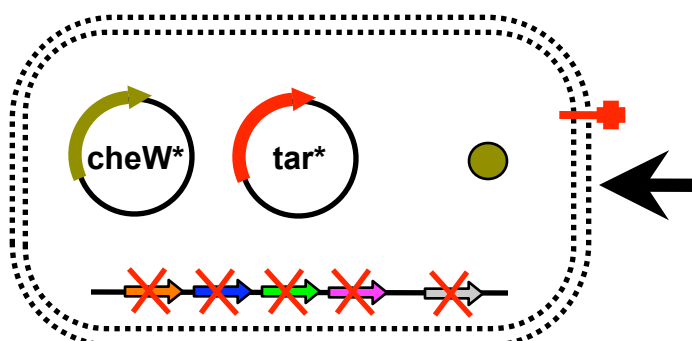
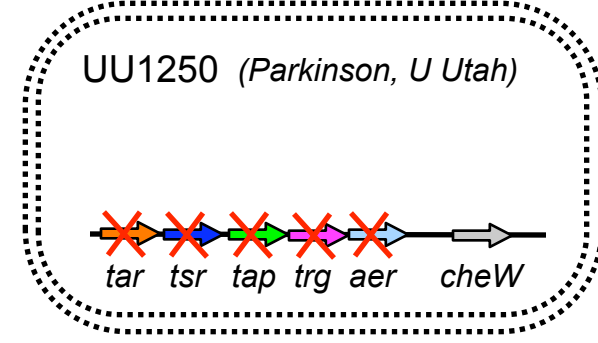
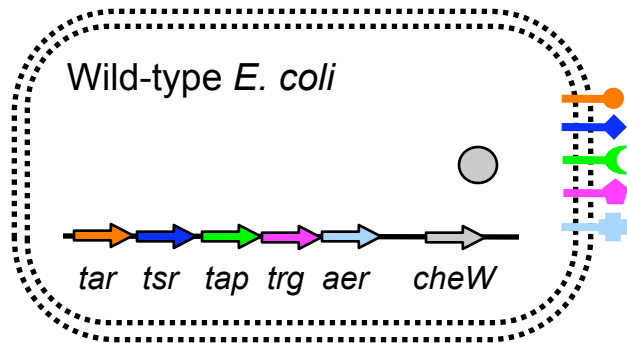
# Chassis Engineering

- Build *E. coli* strain lacking all sensors and CheW, but retaining the other chemotaxis proteins

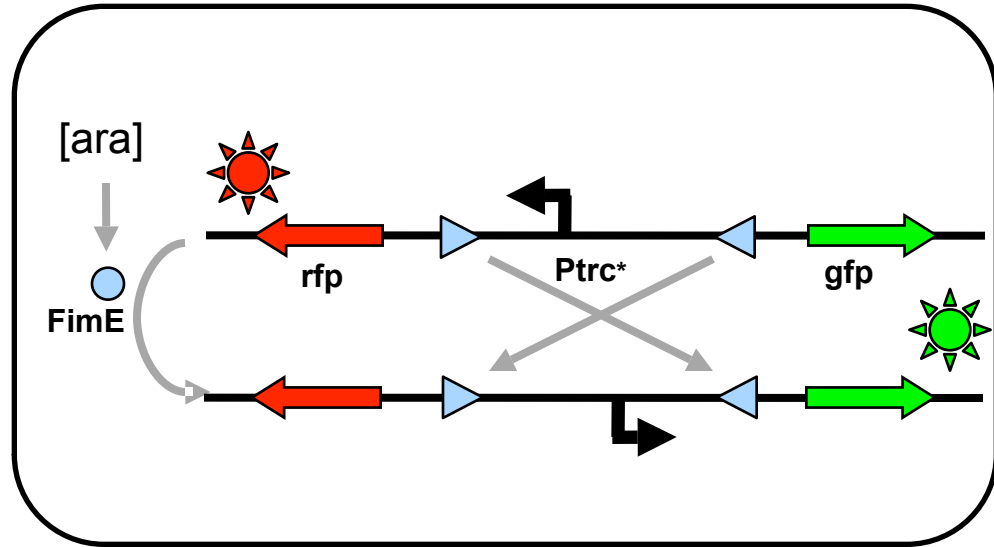
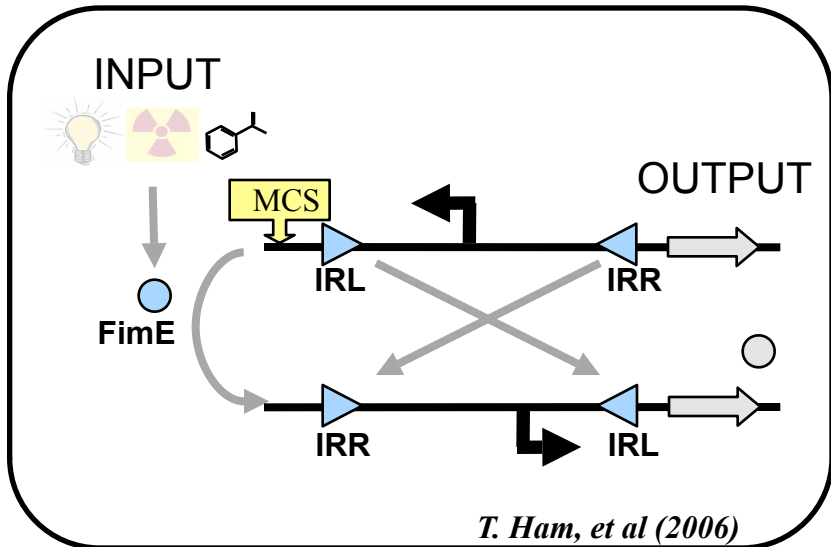


# Chassis Engineering

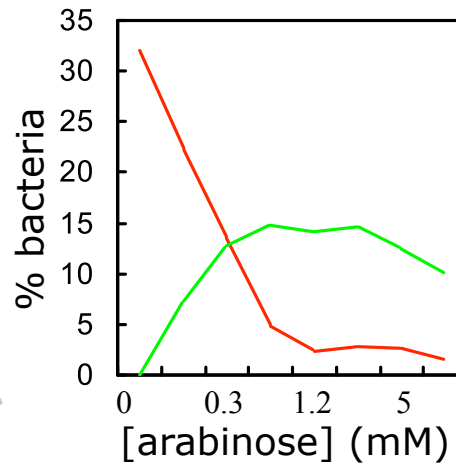
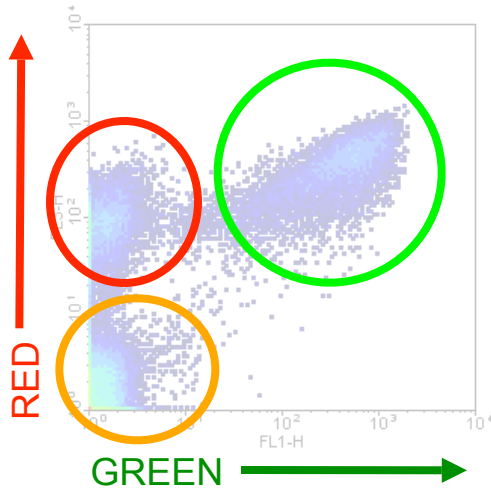
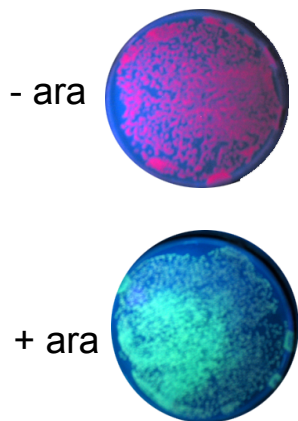
- Build *E. coli* strain lacking all sensors and CheW, but retaining the other chemotaxis proteins



# Device Characterization: The Fim-Switch



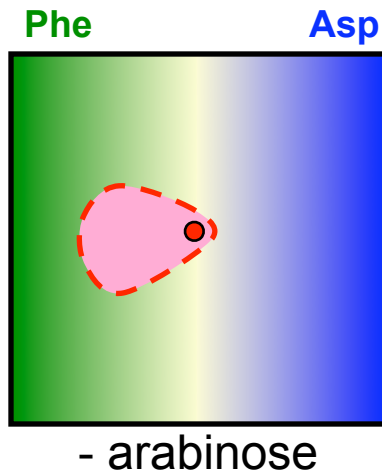
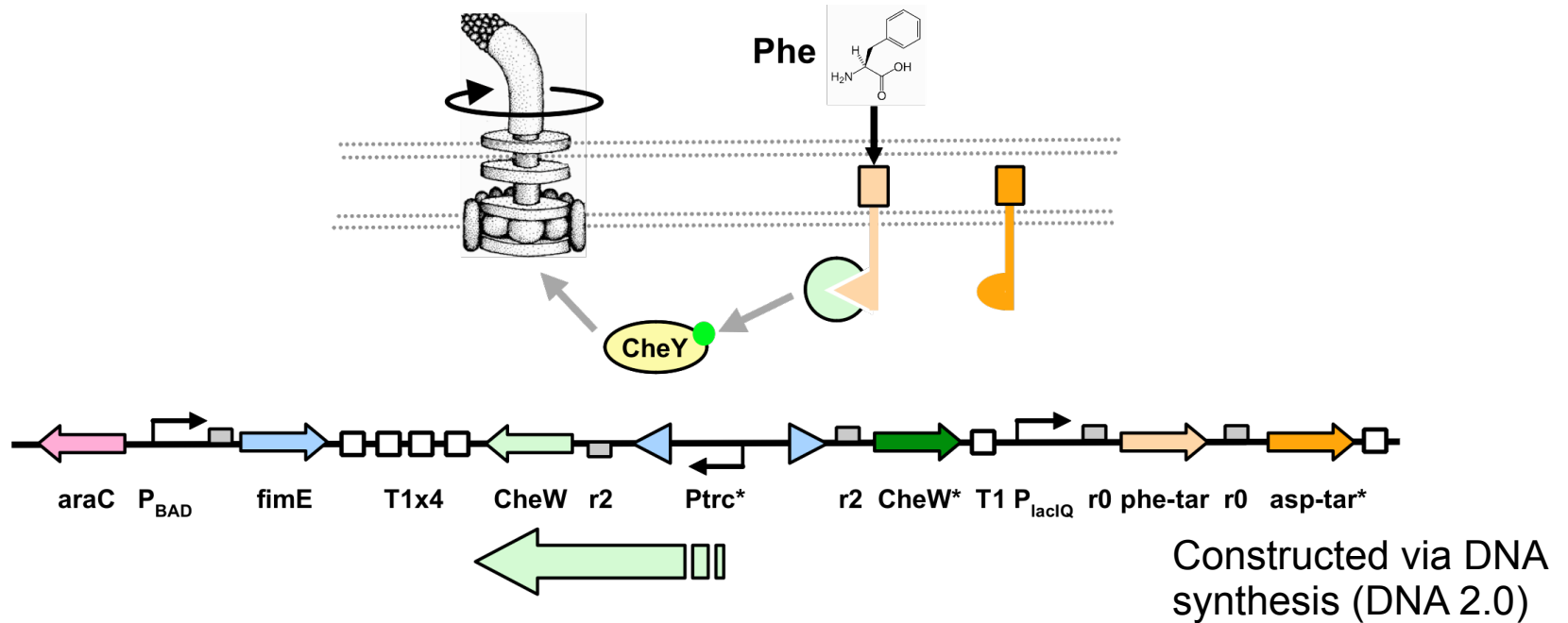
## Tested in UU1250



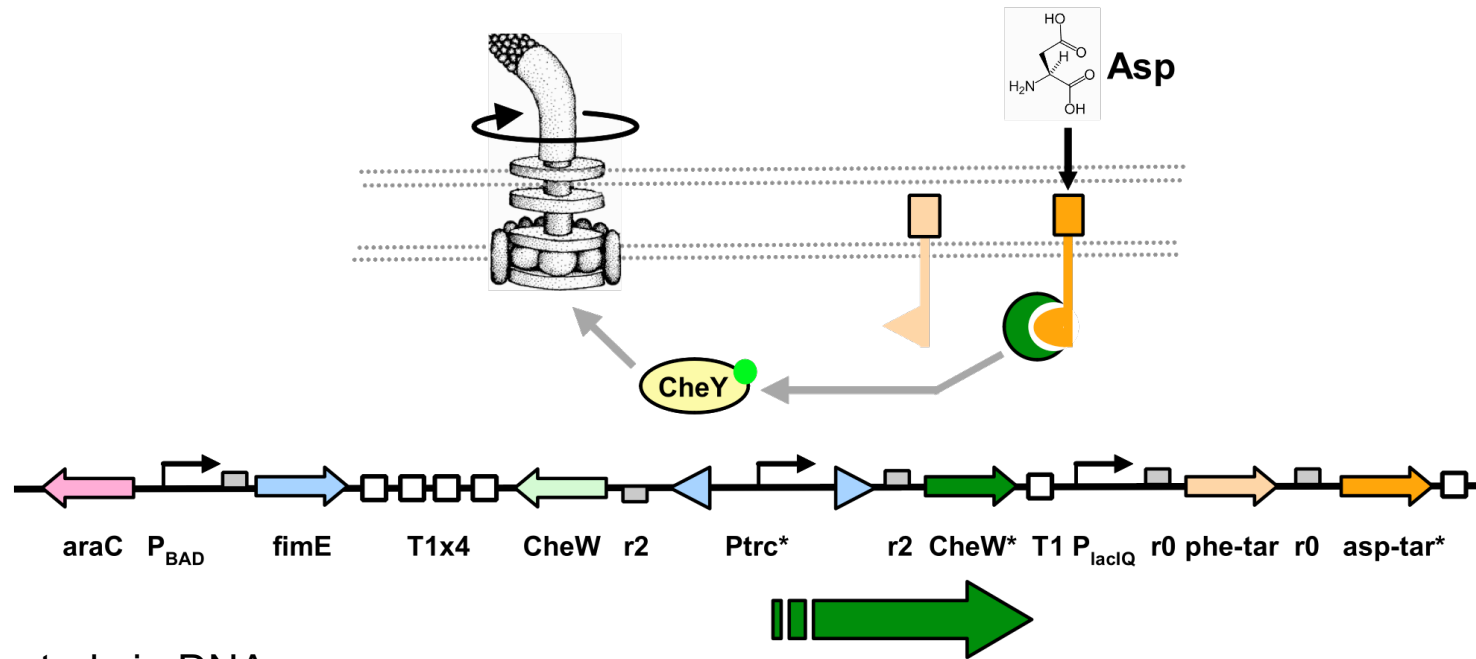
### 2 Possible Problems

1. The plasmid is unstable
2. The strain contains FimE/ FimB

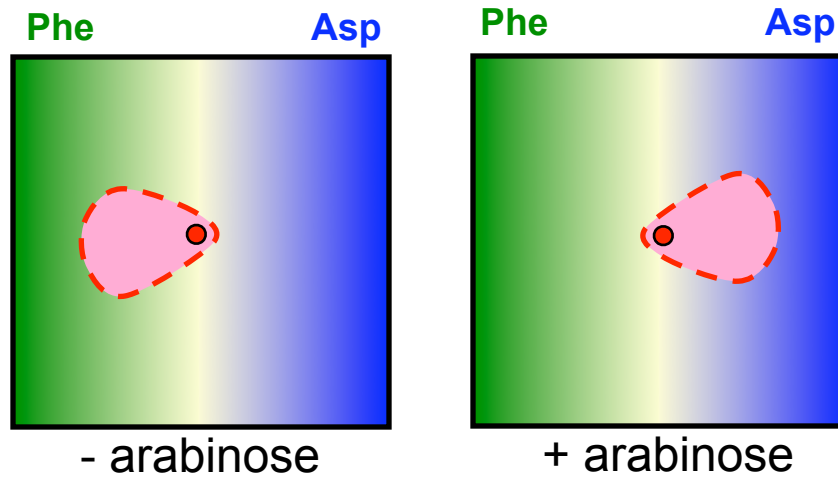
# System Design (11 kB)



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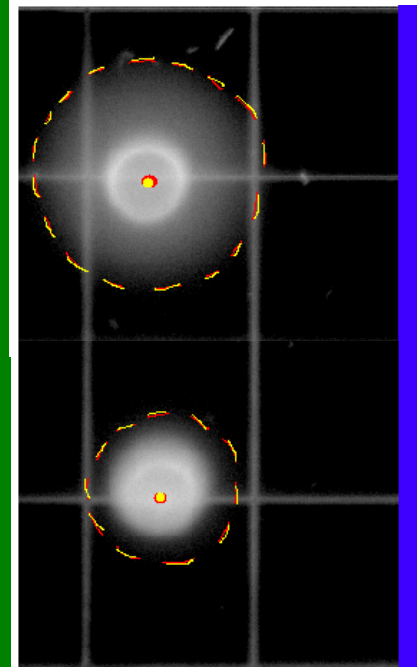
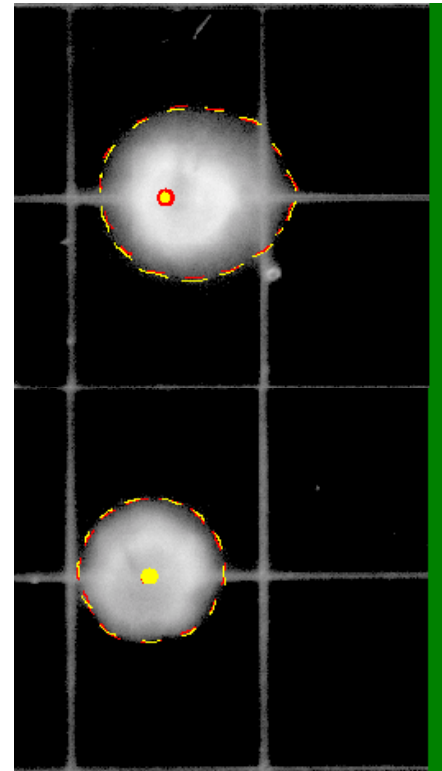
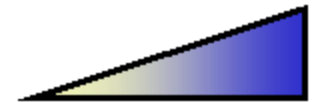
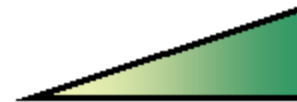
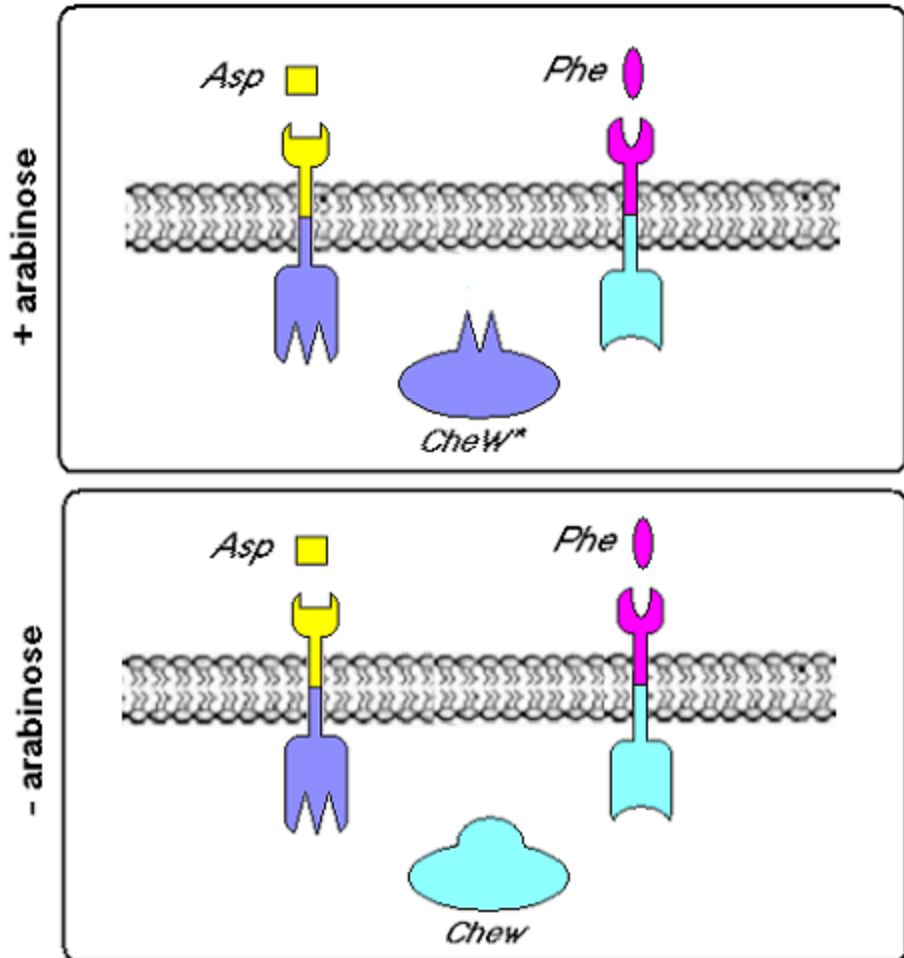
Constructed via DNA synthesis (DNA 2.0)



# Results

Gradient  
Aspartate

Gradient  
Phenylalanine



# Conclusions

- Orthogonal pairs are a rapid method to built signaling pathways that can operate simultaneously
- Chassis developed to rapidly screen for new protein-protein interactions
- Limited by the switch performance in this chassis
- Directional control could also be achieved by non-chemical inputs (light, radiation, etc)