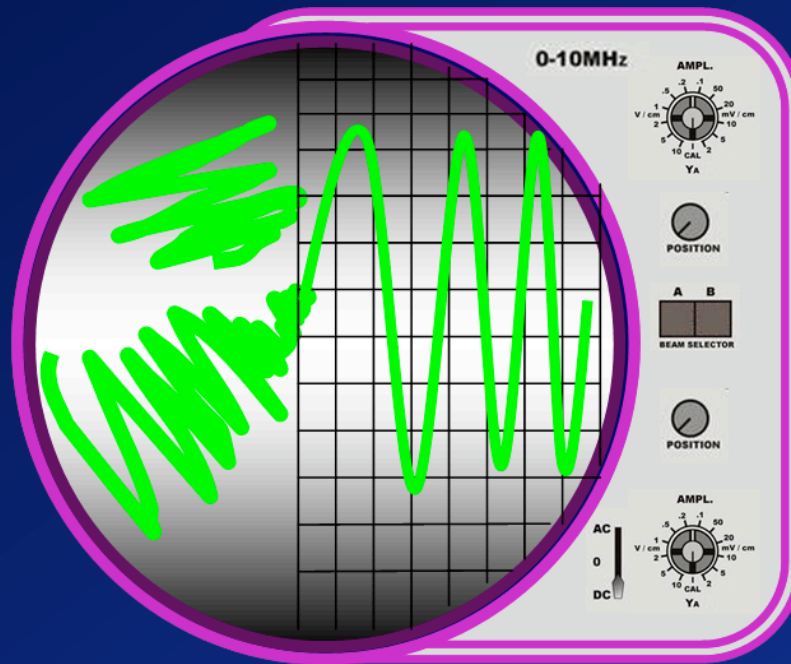
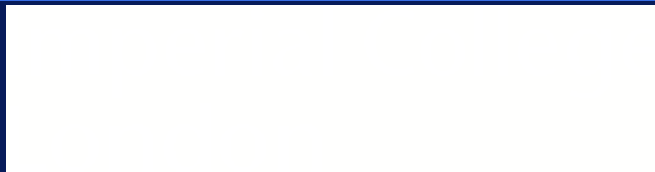


iGEM Jamboree  
MIT – Nov 4th 2006





# Engineering a Molecular Predation Oscillator



# iGEM 2006 @ Imperial



Biomedical Engineers



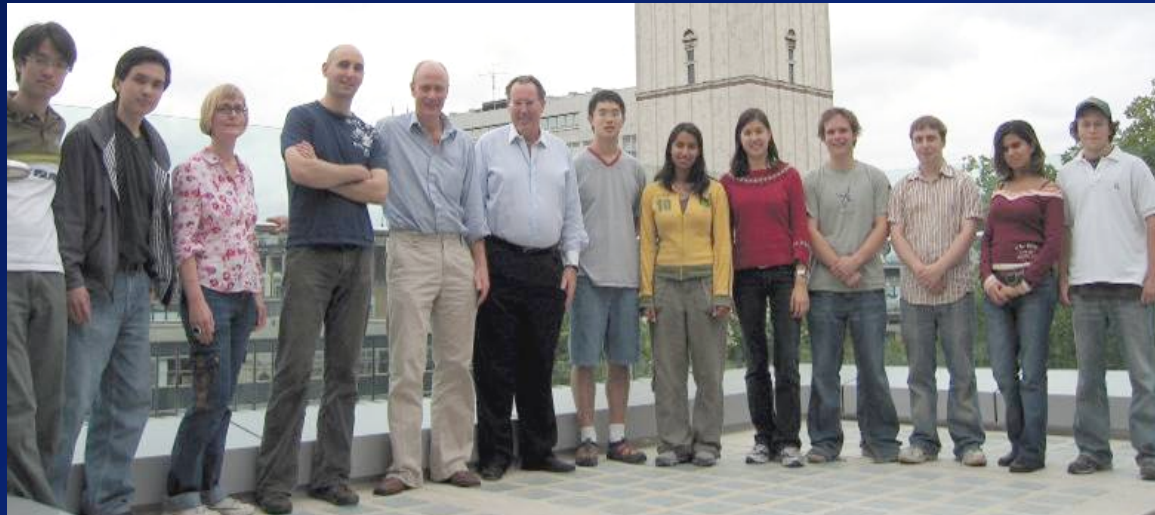
Electrical Engineer



Biochemist



Biologists



Biomedical Engineers

Biochemists



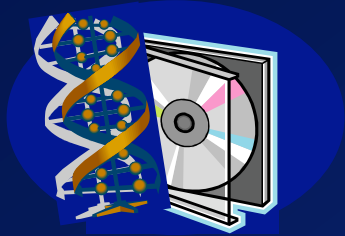
Imperial College



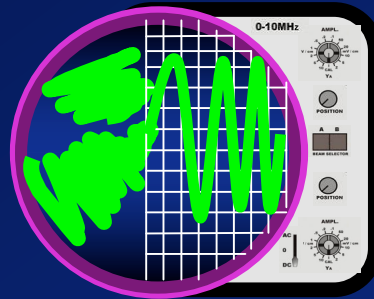
Dr Mann



# Project Ideas



Bio-Memory

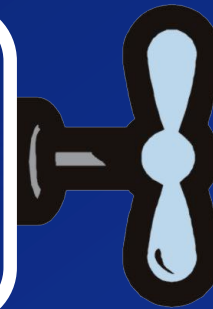


Oscillator



Bio-Clock

- Feasibility
- Originality of Design
- BioBrick Availability
- Future Impact



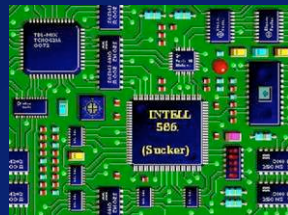


# What is an Oscillator?

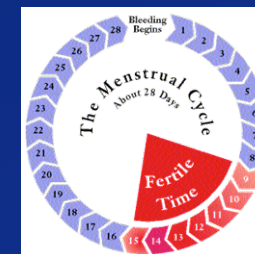
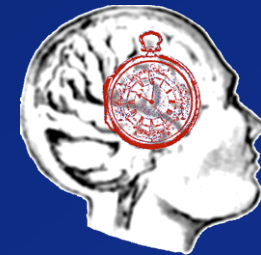
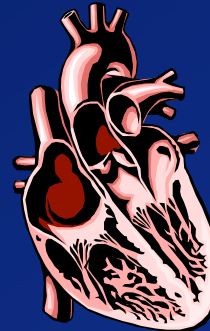
## Our Definition

Device producing a **periodic variation in time** of a measurable quantity, e.g. amplitude.

## Engineering



## Biology



# The Engineering Approach





# The Main Challenges

## Main challenges of past oscillators:

- Unstable
- Noisy
- Inflexible

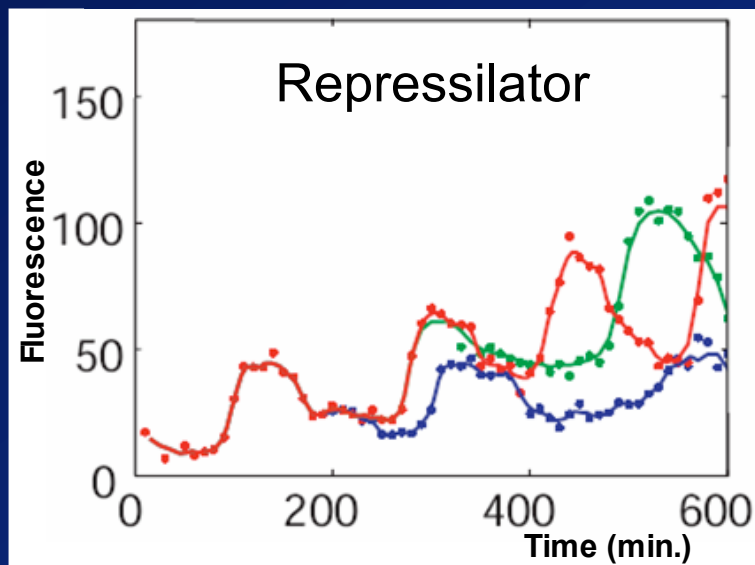


Figure Reference : Michael B. Elowitz & Stanislas Leibler *Nature* 2000

## Requirement for a Our Specifications: typical engineering oscillator

- Sustained Oscillations
- **Stability**:  $> 10$  periods
- **SINR**: Signal to Noise Ratio
- **Flexibility**: Controllable Amplitude and Frequency
- Standardized Device for **Modular Design**
- Easy Connectivity
- **Easy Connectivity**



# Our Initial Design Ideas

Based on

- Large populations of molecules to reduce influence of noise
- Oscillations due to population dynamics
- A well characterized model

## Molecular Predator - Prey





# The Lotka-Volterra Model

$$\frac{dN}{dt}$$

=

Prey Growth



—

Prey Killing  
by Predator



$$\frac{dP}{dt}$$

=

Predator Growth

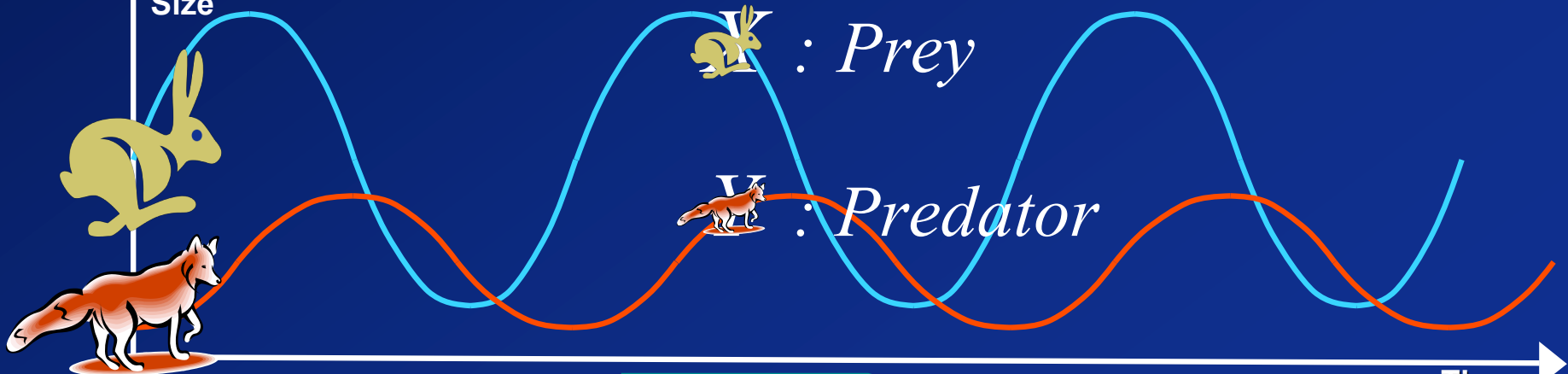


—

Predator Death



Population  
Size





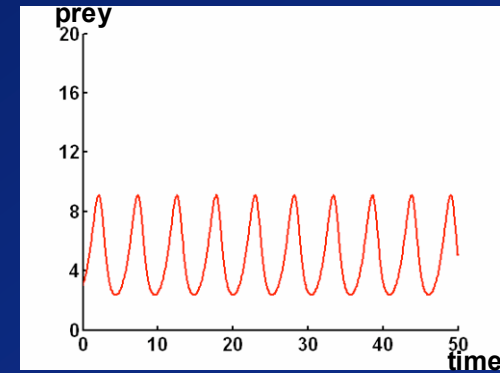
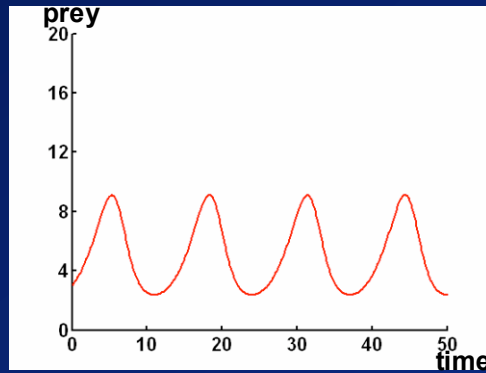
# Typical LV Simulations

Graph of  
Prey vs. Time

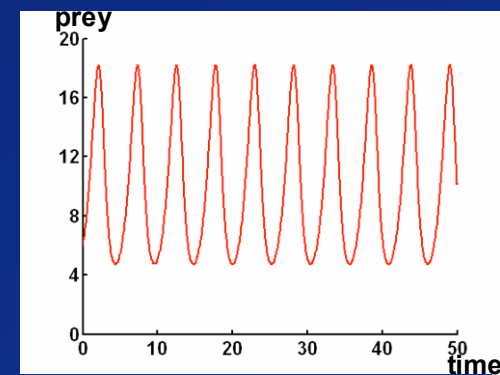
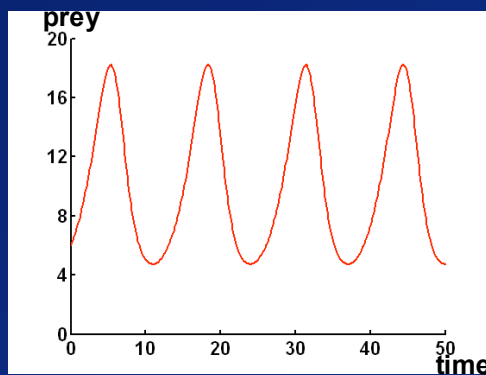
Low Frequency

High Frequency

Small  
Amplitude



Large  
Amplitude





# Required Biochemical Properties

$$\frac{dA}{dt}$$

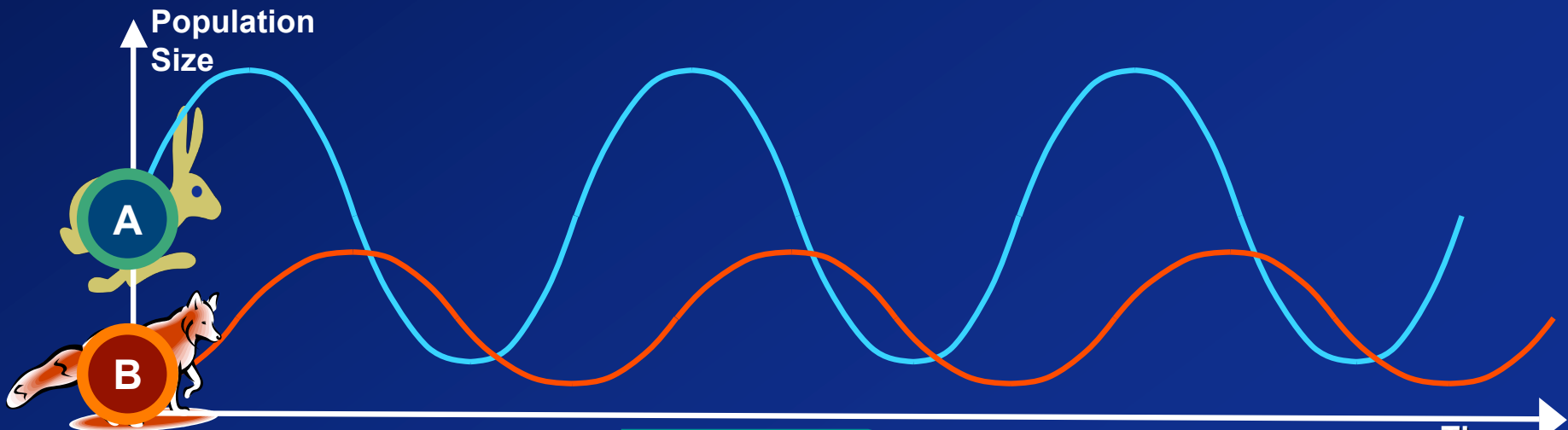
Self promoted  
expression of A

Degradation  
of A by B

$$\frac{dB}{dt}$$

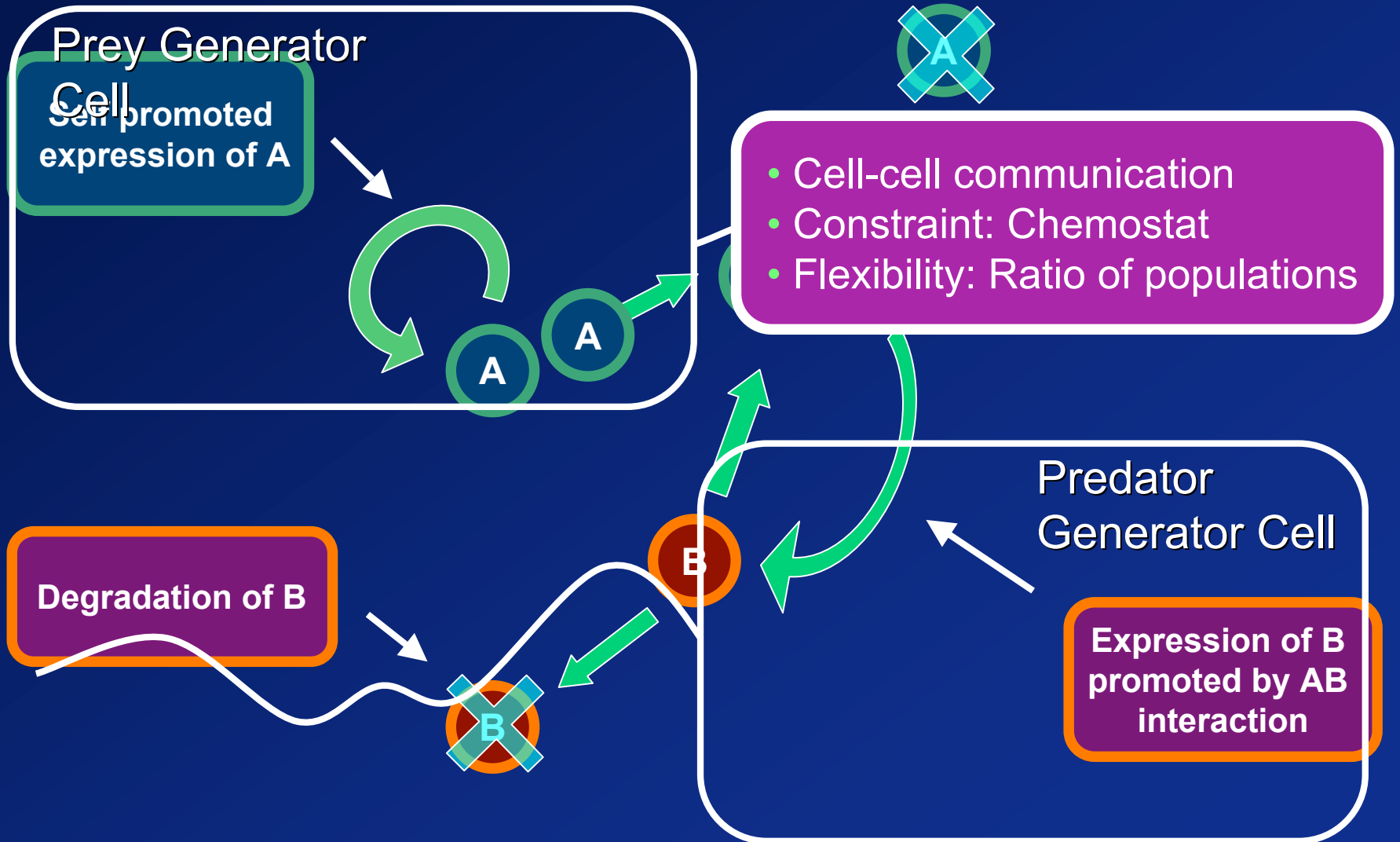
Expression of B  
promoted by  
AB interaction

Degradation  
of B



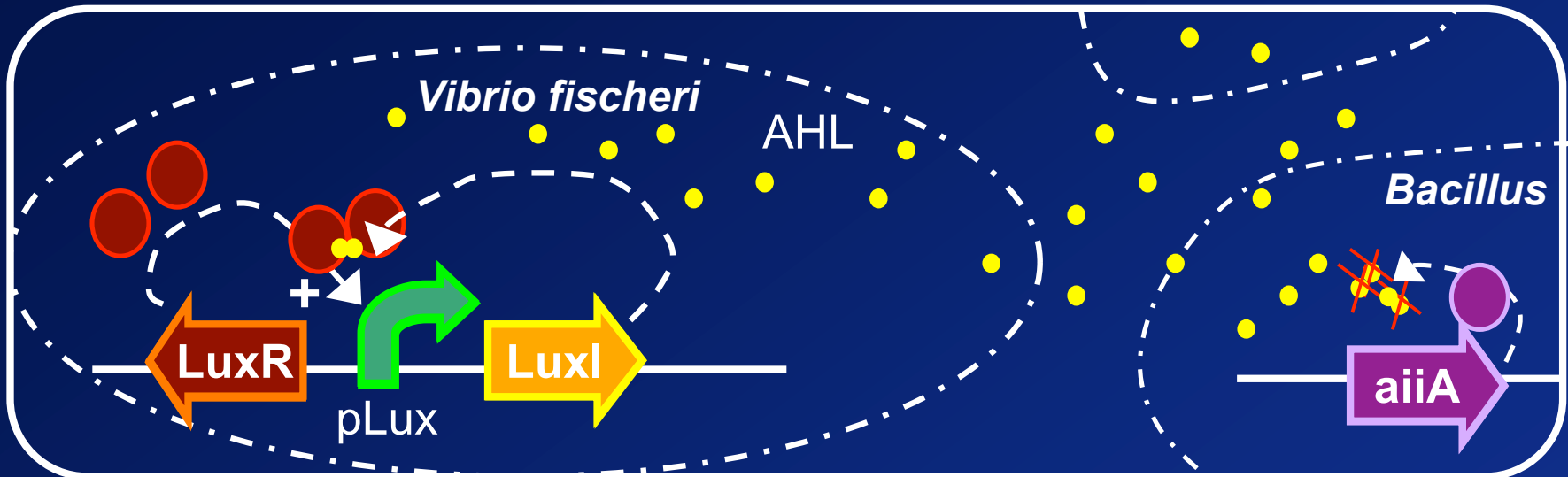


# Molecular System





# Quorum sensing/quenching



## BioBricks available

BBa_C0062	Forms a complex with AHL to activate pLux	BBa_C0061	Makes AHL
pLux BBa_R0062	pLux Promoter	BBa_C0160	Degrades AHL
<p>BBa_F2620</p>		AHL->Pops Receiver	



# Designing the Prey Generator

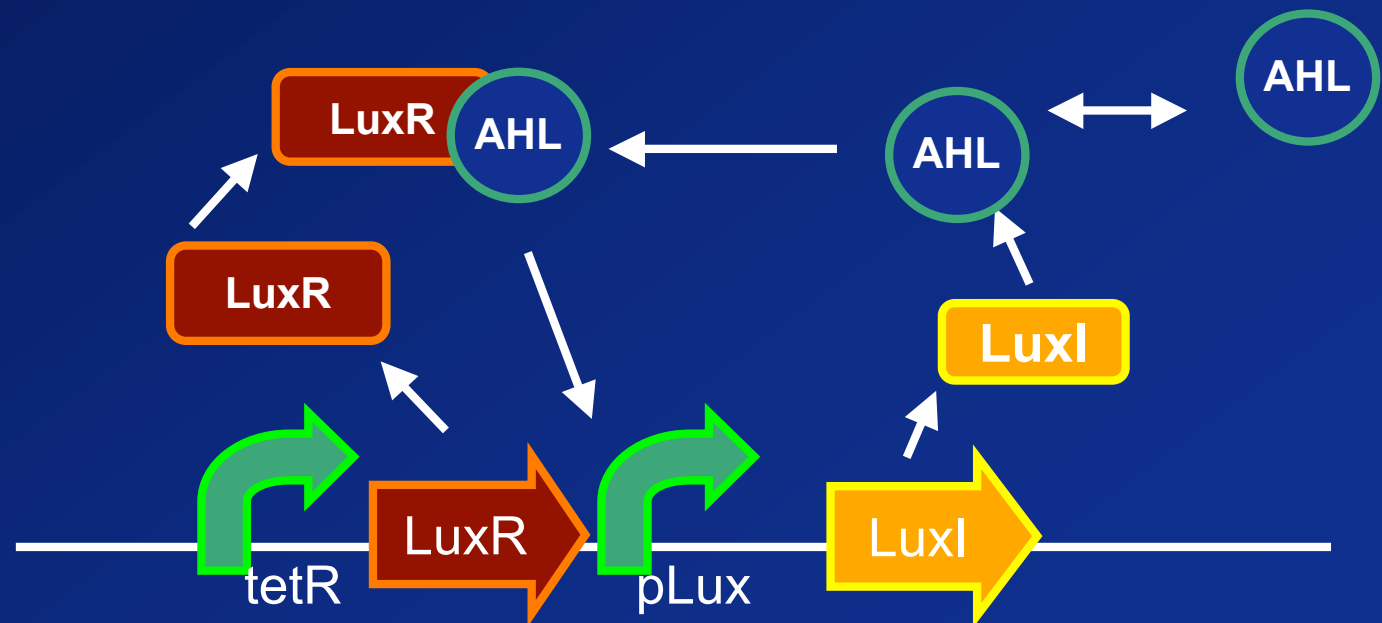
Required  
Dynamic

Self promoted  
expression of A ✓

Useful  
BioBricks



Final  
Construct





# Designing the Predator Generator

Required  
Dynamic

Expression of B  
promoted by  
AB interaction ✓

Degradation  
of A by B ✓

Degradation  
of B ✓

Useful  
BioBricks

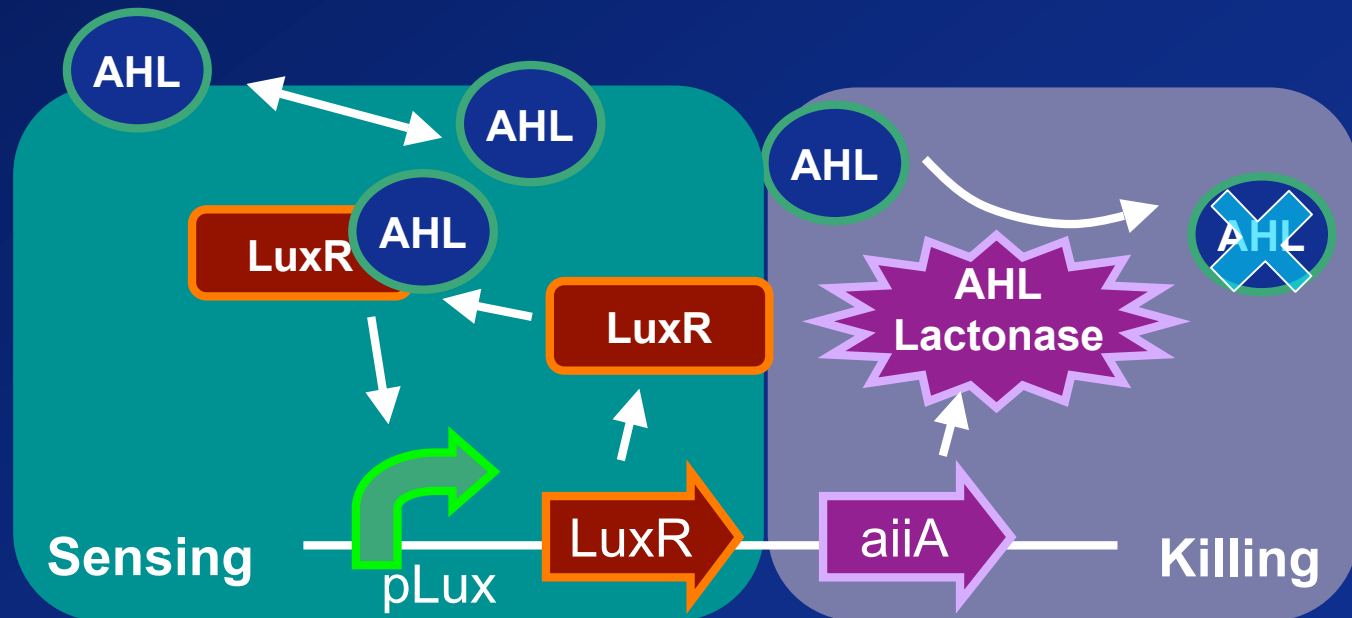
pLux  
R0062

LuxR  
C0062

aiiA  
C0060

Natural  
degradation

Final  
Construct

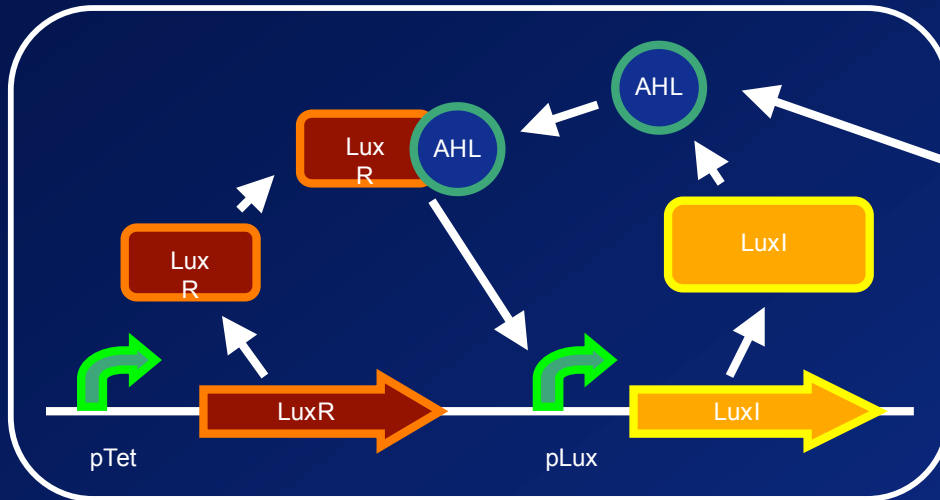




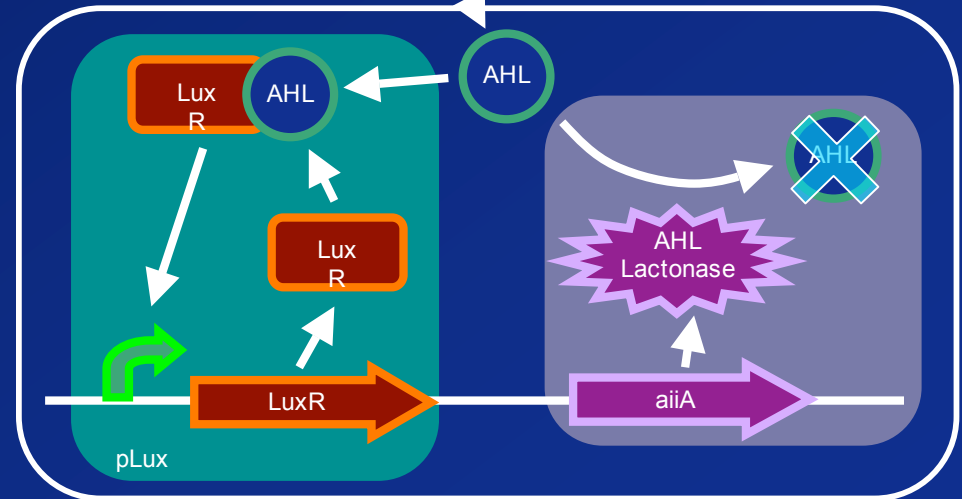
# System Overview

## Prey Generator Cell

Pool of AHL will oscillate

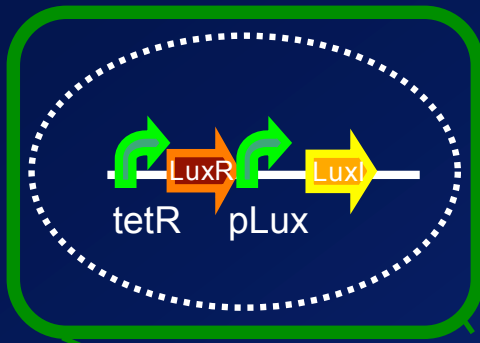


## Predator Generator Cell

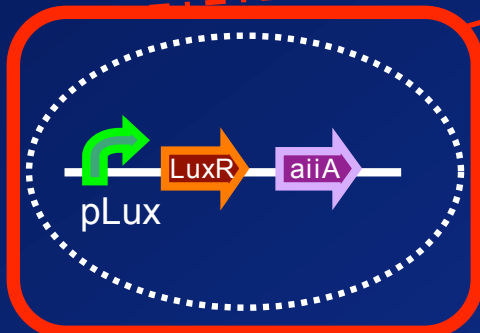




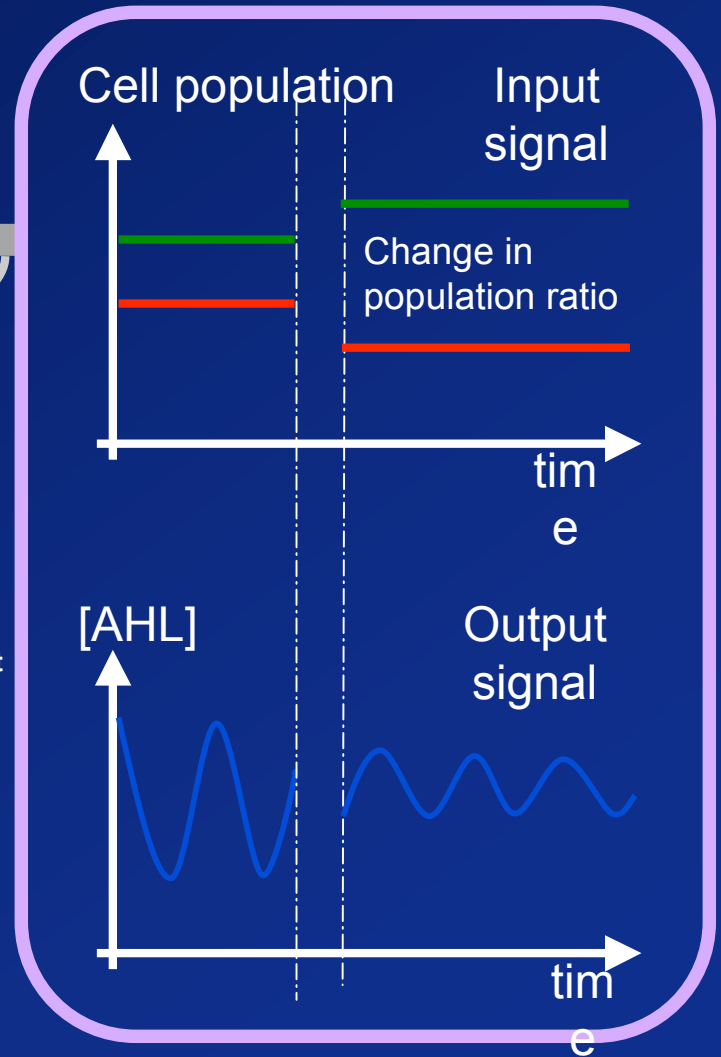
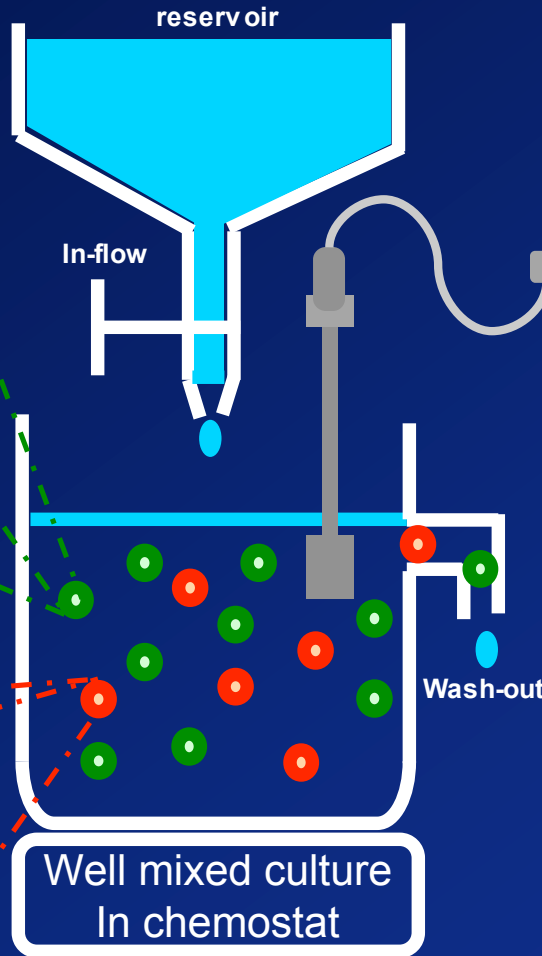
# Full System set-up



Prey molecule generator



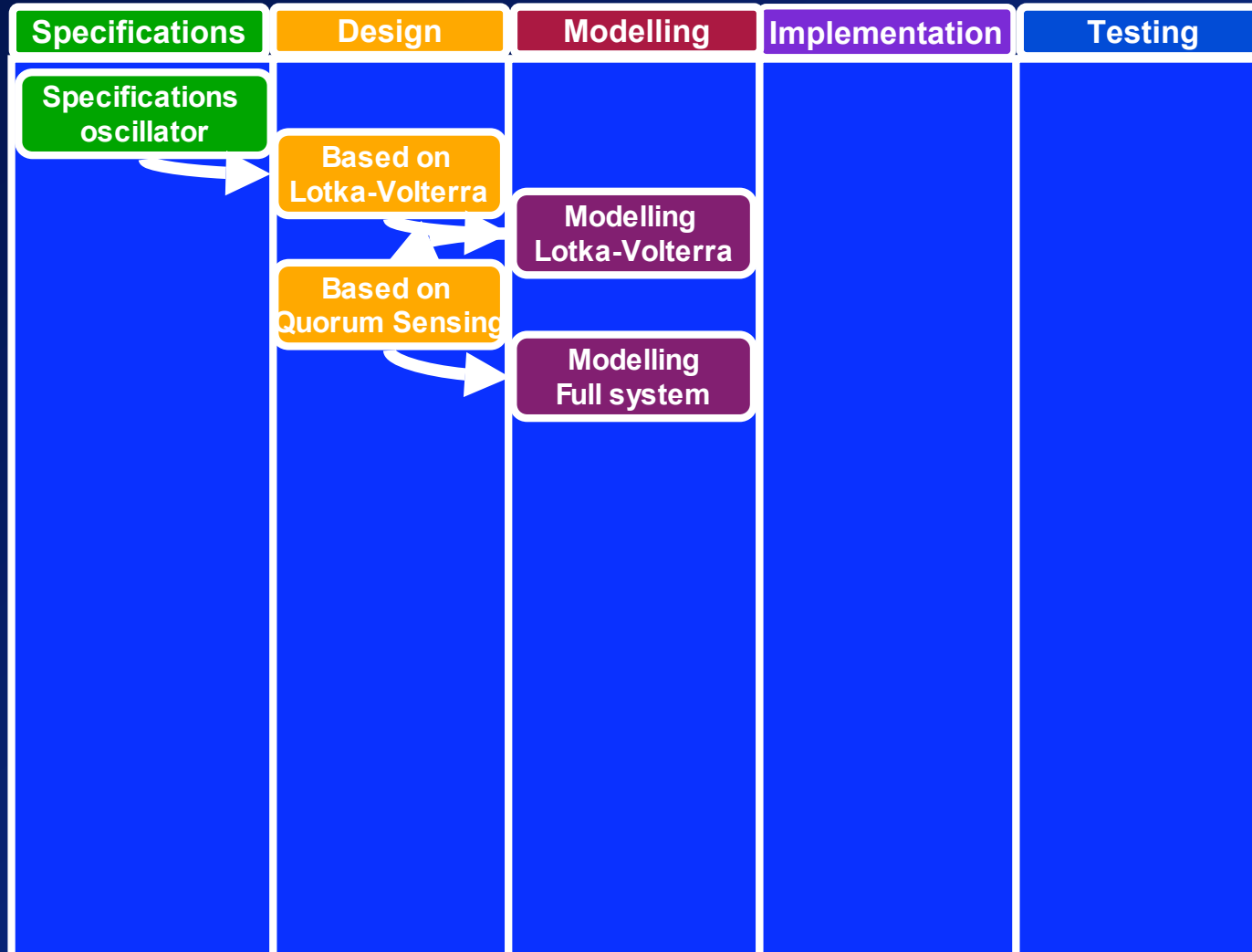
Predator molecule generator





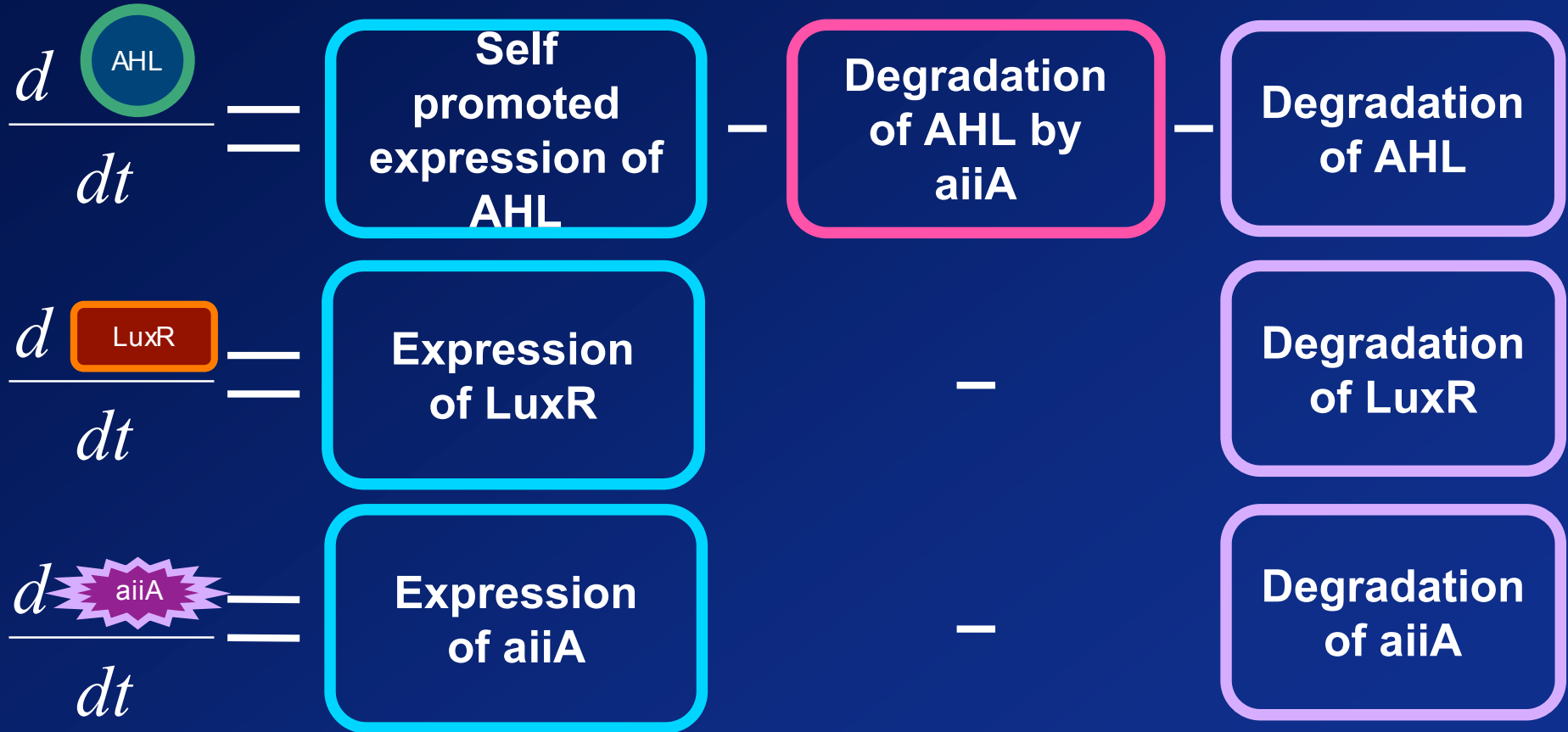
# Path to Our Goal

Start!  
→



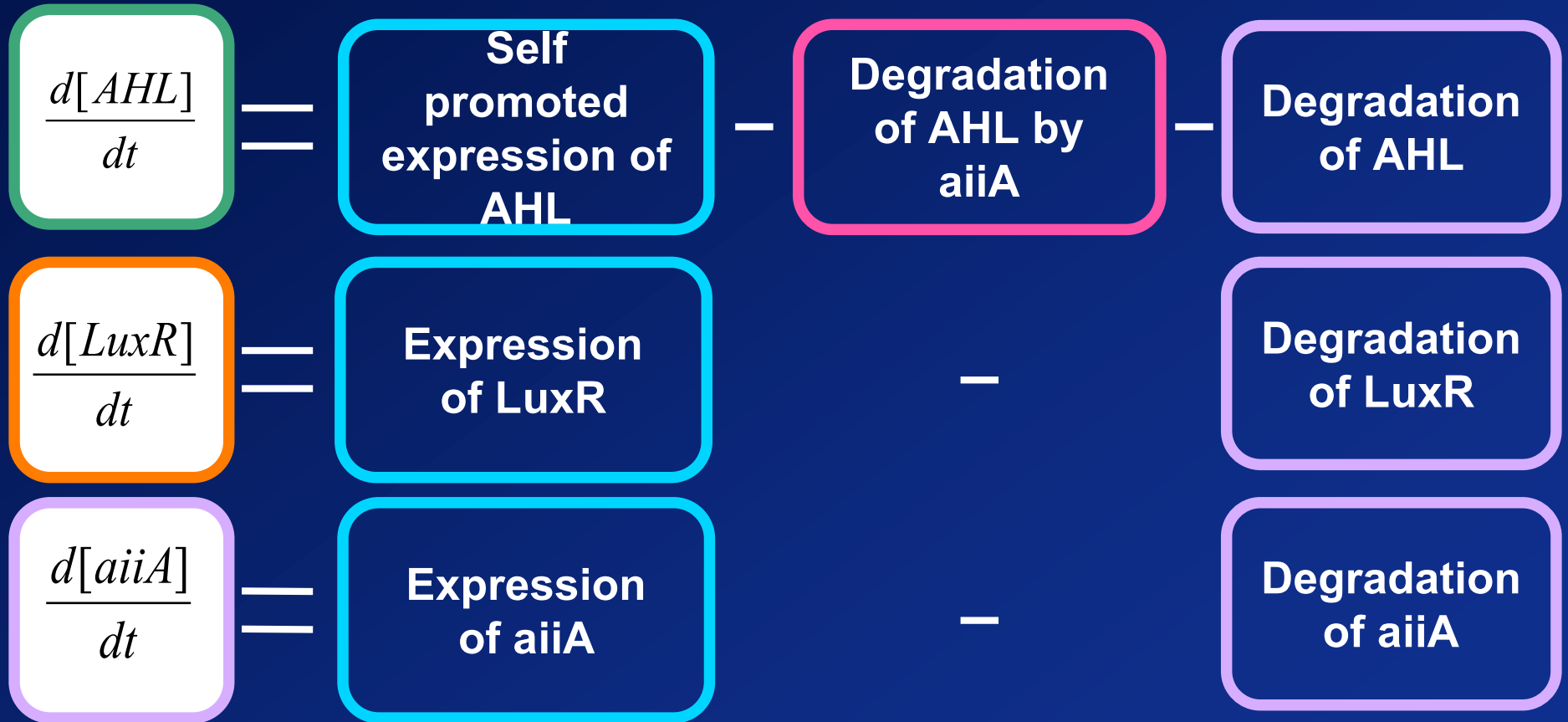


# Modelling the Full System





# Modelling the Full System





# Modelling the Full System

## Gene Expression

$$\frac{d[AHL]}{dt}$$

=

$$\frac{a[AHL]}{a_0 + [AHL]}$$

-

Degradation  
of AHL by  
aiaA

-

Degradation  
of AHL

$$\frac{d[LuxR]}{dt}$$

=

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

-

Degradation  
of LuxR

$$\frac{d[aiaA]}{dt}$$

=

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

-

Degradation  
of aiaA



# Modelling the Full System

Gene Expression

Enzymatic Reaction

$$\frac{d[AHL]}{dt}$$

$$\frac{a[AHL]}{a_0 + [AHL]}$$

$$\frac{b[aiiA][AHL]}{b_0 + [AHL]}$$

Degradation of AHL

$$\frac{d[LuxR]}{dt}$$

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

Degradation of LuxR

$$\frac{d[aiiA]}{dt}$$

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

Degradation of aiiA



# Modelling the Full System

Gene Expression

Enzymatic Reaction

Degradation

$$\frac{d[AHL]}{dt}$$

=

$$\frac{a[AHL]}{a_0 + [AHL]}$$

-

$$\frac{b[aiiA][AHL]}{b_0 + [AHL]}$$

-

$$e[AHL]$$

$$\frac{d[LuxR]}{dt}$$

=

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

-

$$d_1[LuxR]$$

$$\frac{d[aiiA]}{dt}$$

=

$$\frac{c[AHL][LuxR]}{c_0 + [AHL][LuxR]}$$

-

$$d_2[aiiA]$$

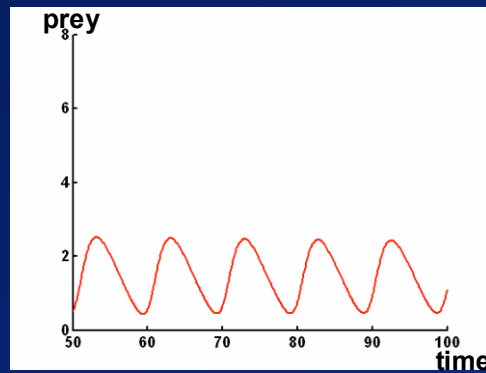


# Full System Simulations

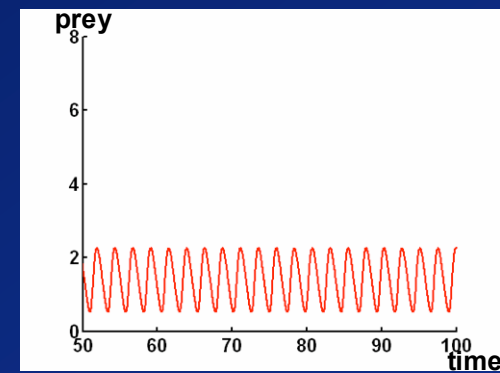
Graph of  
Prey vs.  
Time

Small  
Amplitude

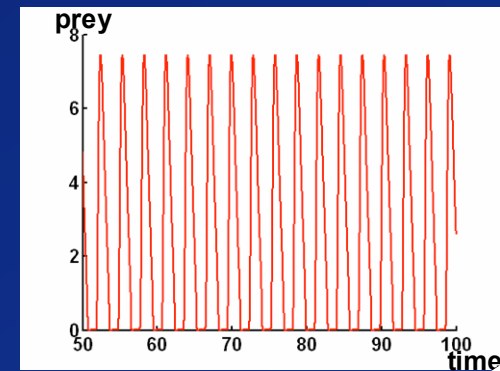
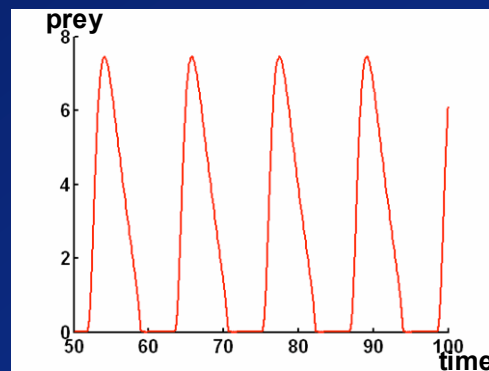
Low Frequency



High Frequency



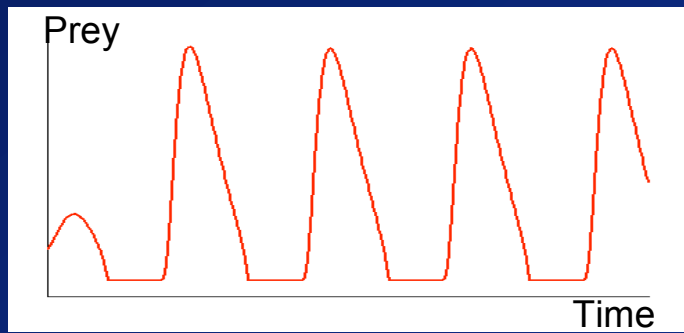
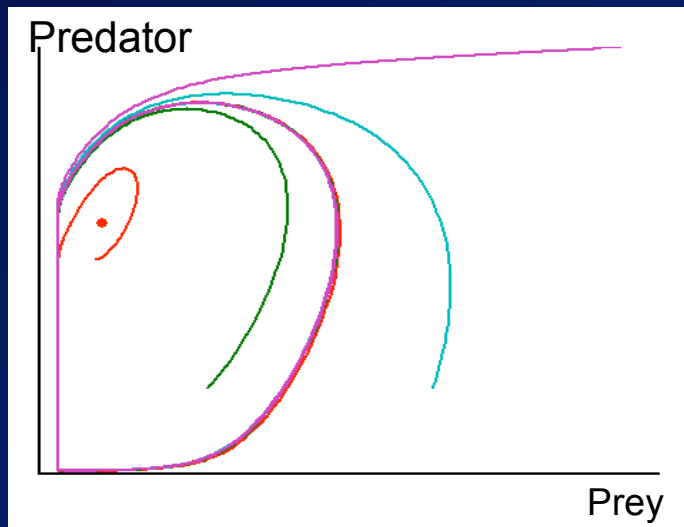
Large  
Amplitude



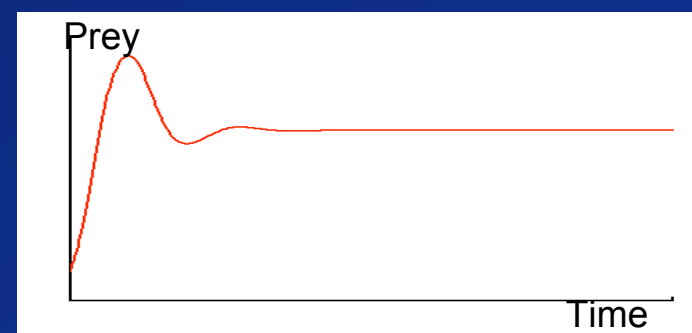
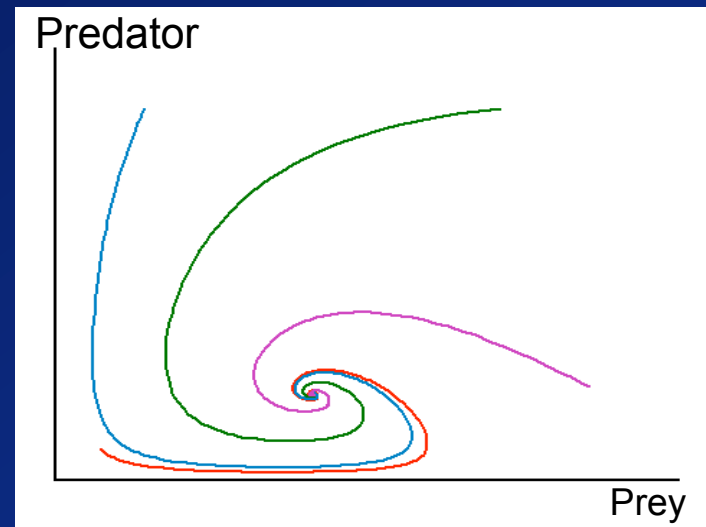


# Typical System Behaviours

## Oscillations with limit cycles

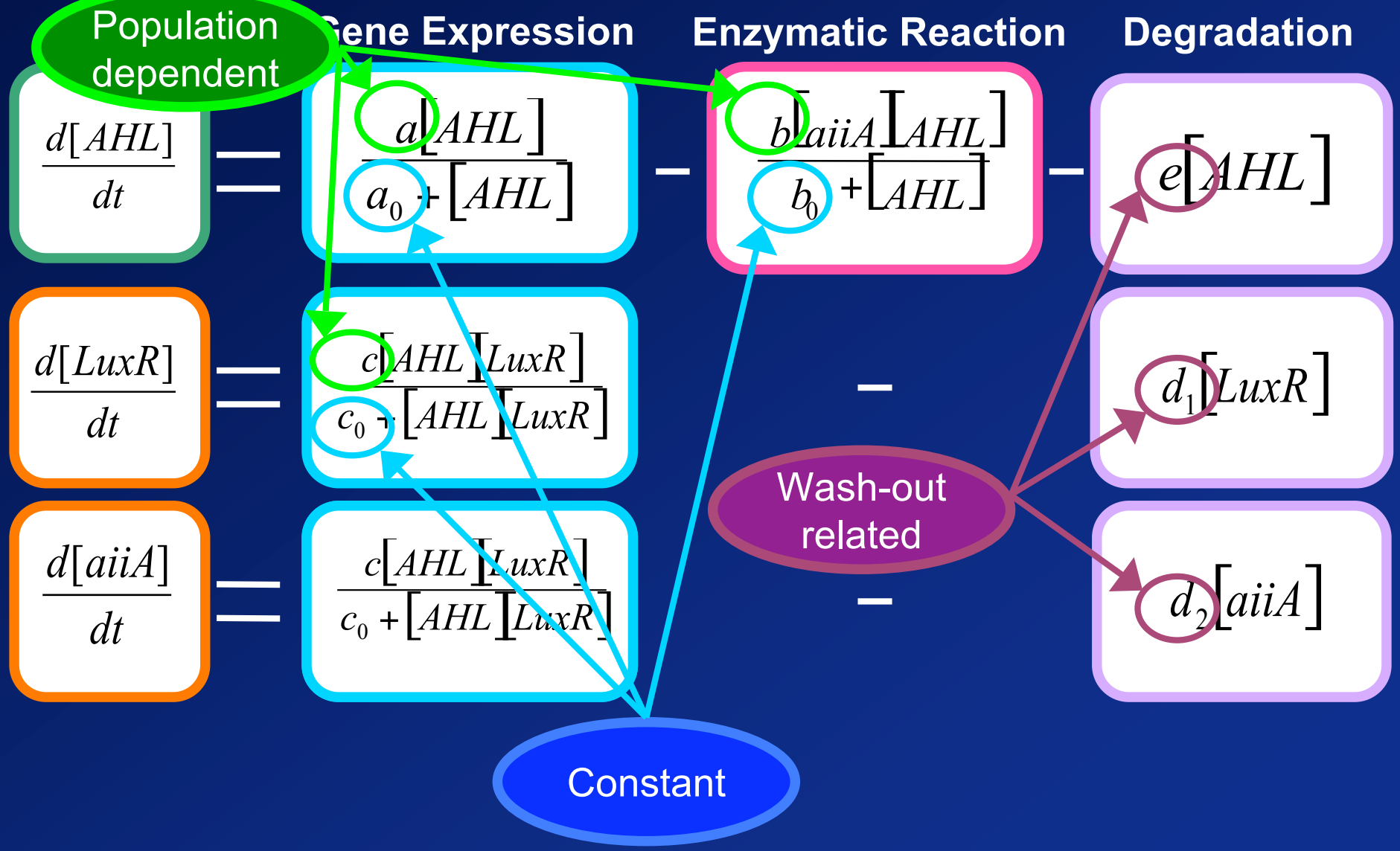


## No oscillations





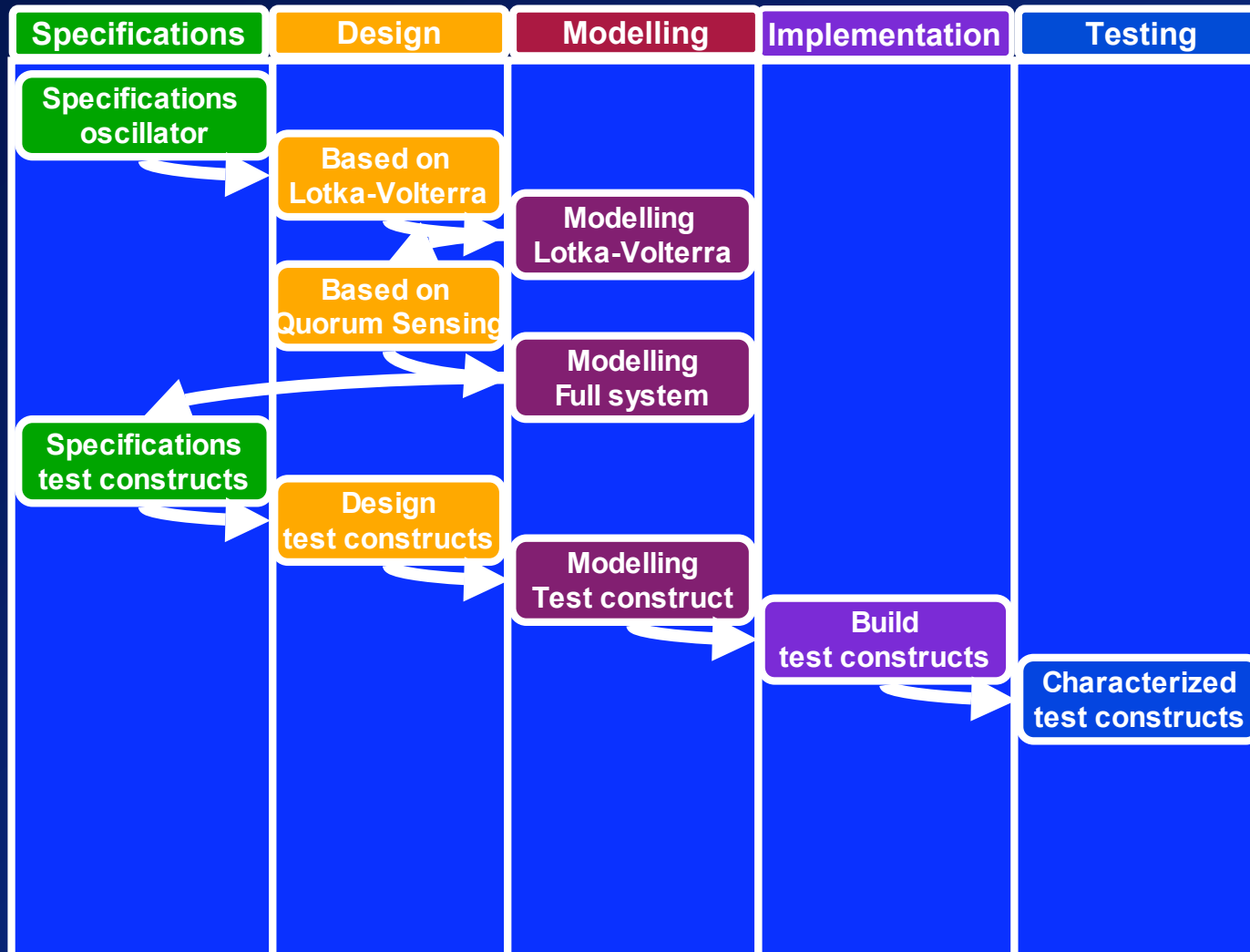
# Modelling the Full System





# Path to Our Goal

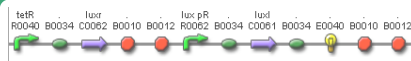
Start!  
→



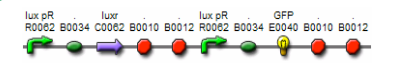


# Breaking Down the Complexity

Prey Generator



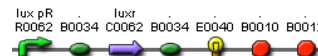
Prey Sensing



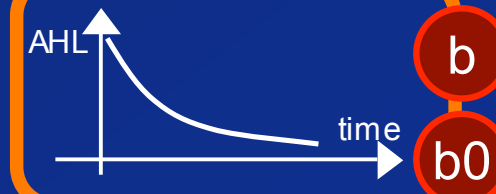
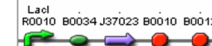
Predator Generator



Predator Sensing



Predator Killing





# Characterization Predator Sensing

Test part

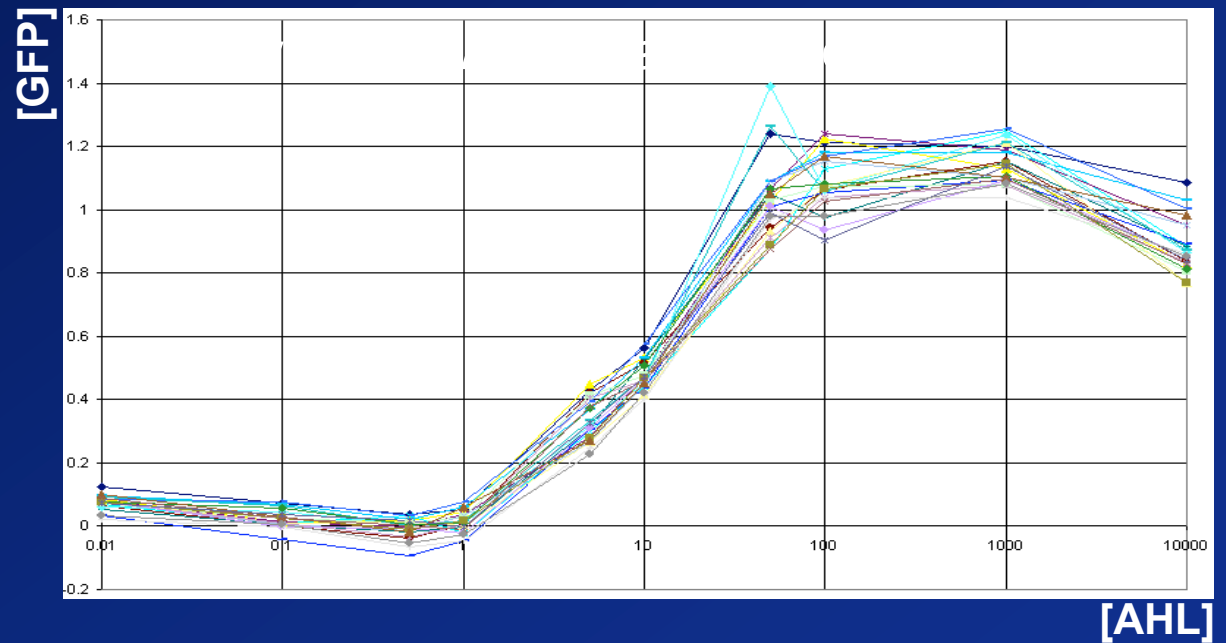


Predictive model  
transfer function



Experimental data

Experimental Data





# Characterization Predator Sensing

Test part



Predictive model  
transfer function



Experimental data

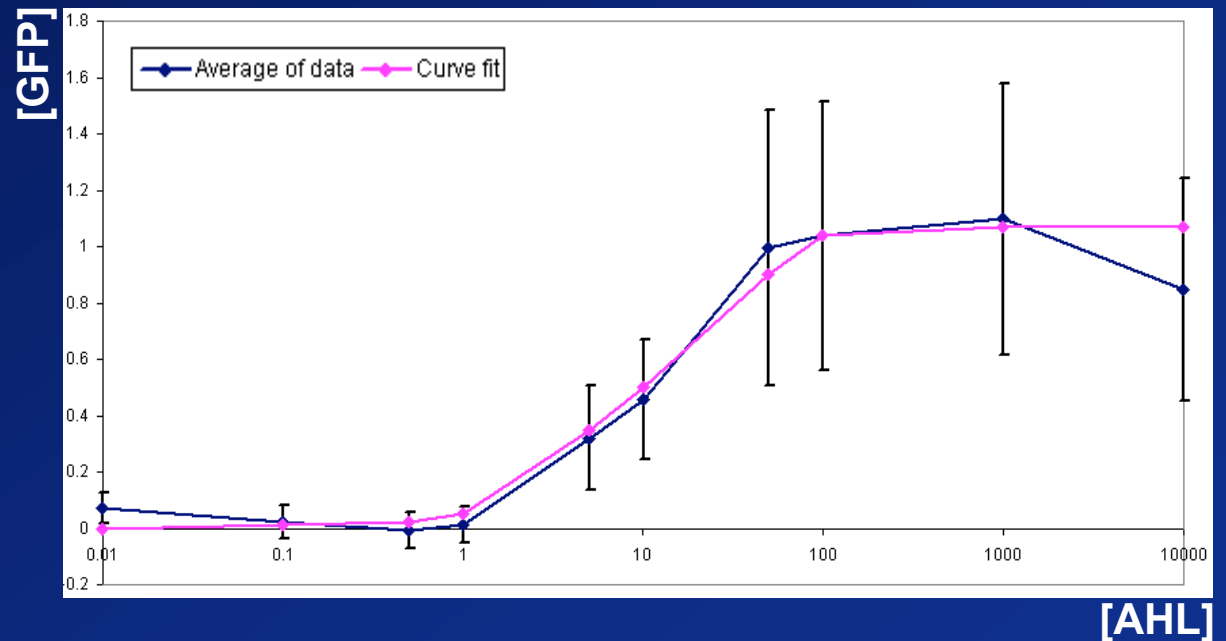


Fitting model to data



Parameter extractions

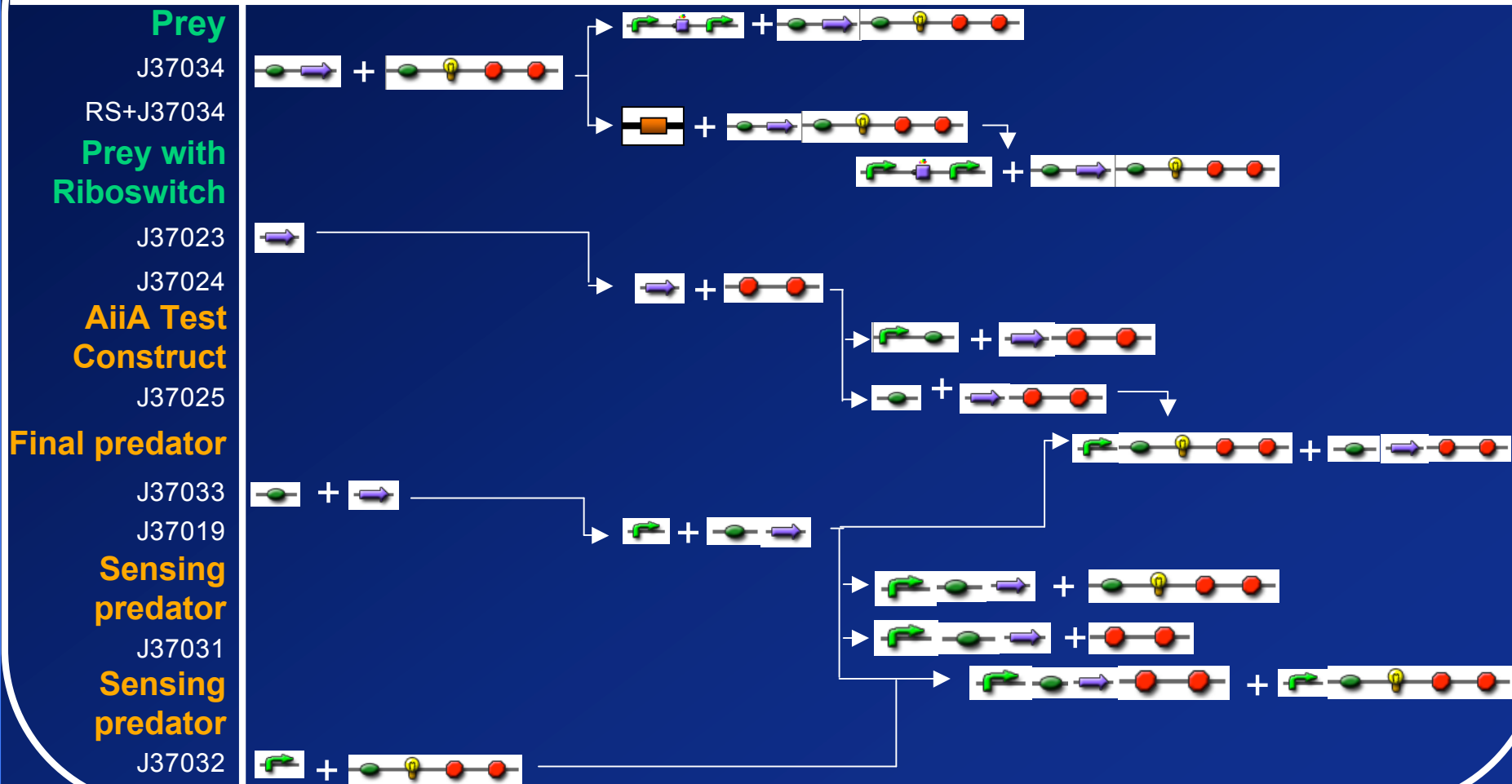
Average with variance and curve fitting



# Implementation

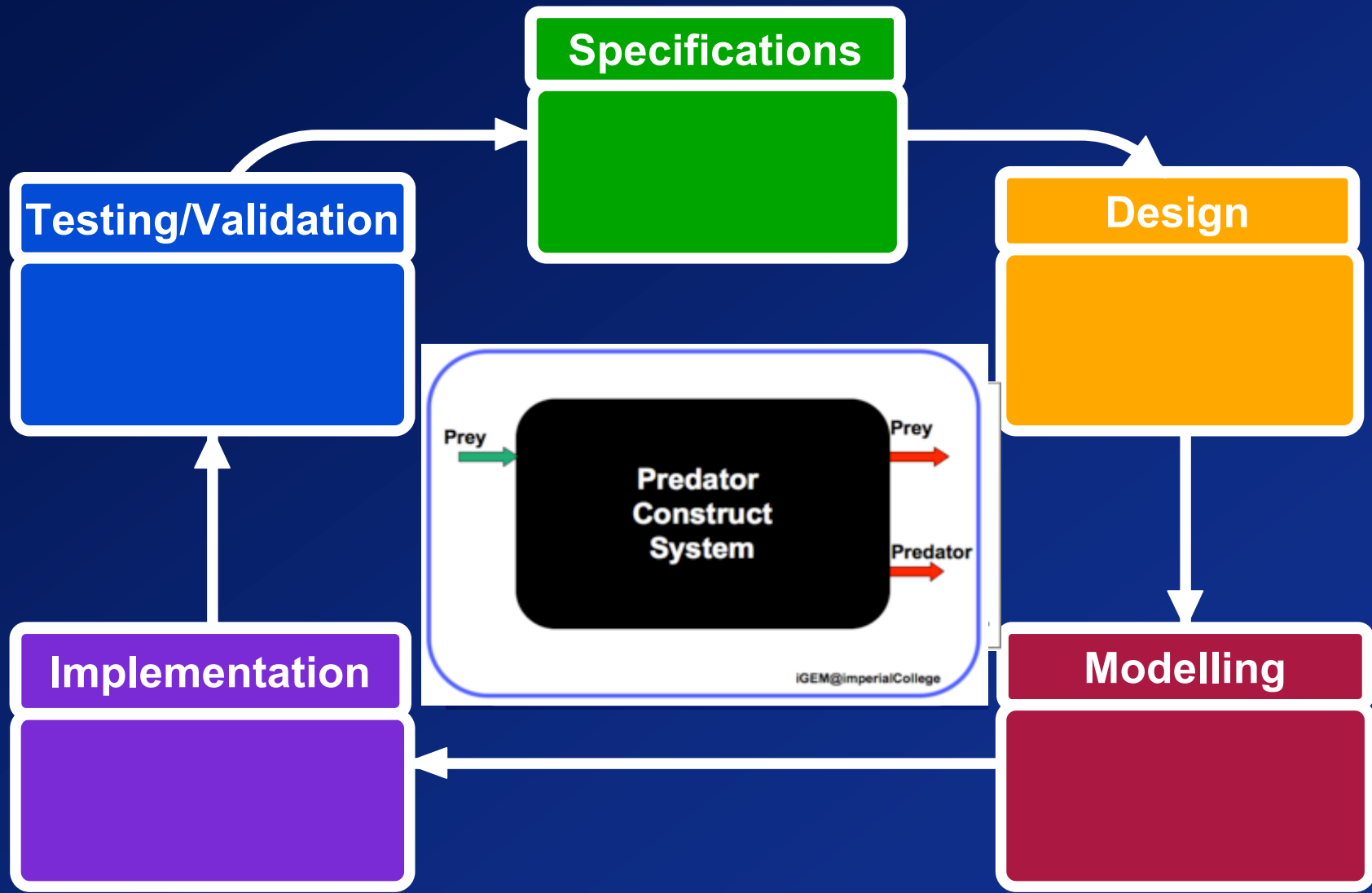
## Registry Catalogue Parts

## Assembly process





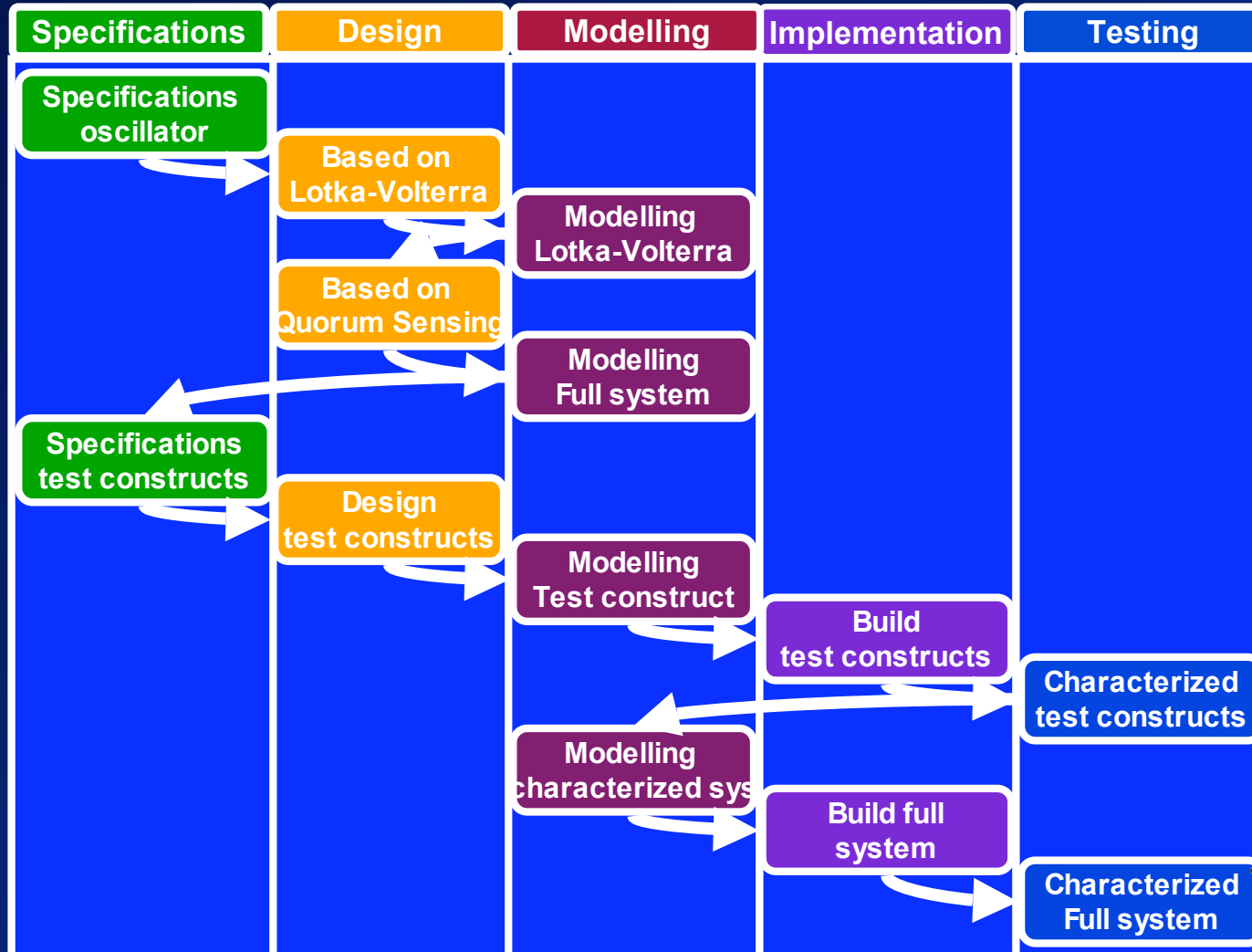
# On Our Experience





# Path to Our Goal

Start!

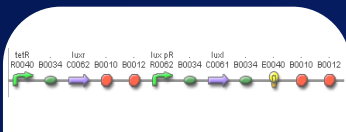
Our Goal!



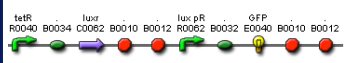
# Contributions to the Registry

## Functional Parts

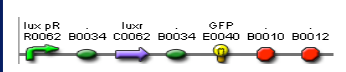
**Final Prey**  
J37015



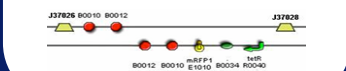
**Sensing Prey**  
T9002



**Sensing Predator**  
J37016



**Cre/Lox**  
J37027

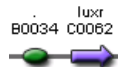


Built Sequenced Tested Characterized Documented

✓	✓	✓	✓	✓
		✓		✓
✓	✓	✓	✓	✓
✓	✓	✓		✓

## Intermediate Parts

J37033



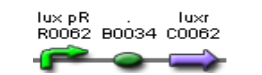
J37034



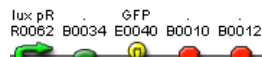
Built Sequenced

✓	✓
✓	✓

J37019



J37032



J37023



Built Sequenced

✓	✓
✓	✓
✓	✓



# Our Wiki

Imperial College London

**THE**  **REPORTER** 

The newsletter of Imperial College London iGEM 2006

[Our Contributions](#) [To Do List](#) [Biological Oscillator Parts](#) [Modelling](#) [Protocols](#) [Resources](#)

**Breaking News** [\[edit\]](#)

- 30 Oct 2006: 300+ pages documentation on OWW. New iGEM page
- 27 Oct 2006: Team Poloshirts arrived.
- 26 Oct 2006: BioBricks sent to M.I.T.
- 25 Oct 2006: Presentation of the iGEM 2006 project at the Bioengineering Departmental Seminar.
- News Archive ↗

Location of our visitors  
Visits since 6 Aug 2006



Updated daily

**Imperial College London iGEM Project(s) 2006** [\[edit\]](#)

**Engineering a Molecular Predation Oscillator.** [↗](#)



**PoPs Blocker** [↗](#) **Biological to Electrical Interface** [↗](#)

**Celebrities** [\[edit\]](#)



Get to know the I.Coli team and their advisors:

[The Undergrads](#) [\[show\]](#)

[The Advisors](#) [\[show\]](#)

[Acknowledgements](#)

**Activities** [\[edit\]](#)

- Event Calendar
- MIT Jamboree
- BioSysBio Conference, UK
- UK iGEM Teams Meeting in Cambridge

**In the Wetlab** [\[edit\]](#)

- Lab Notebook
- Protocols
- Lab Status
- Testing
- Sequencing of Parts
- Primers

**Education** [\[edit\]](#)

- Brainstorming
- Journal Club
- Lecture Notes
- Bibliography/Papers
- Resources

**Entertainment** [\[edit\]](#)

- Photos
- Logo & Team Shirts
- Inspirational Quotes
- Project Discussion
- Sandbox

- Documentation
- Communication
- Organization

<http://openwetware.org/wiki/IGEM:IMPERIAL/2006>



# Thank You

**Imperial College  
London**

## Acknowledgements:

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- Dr. Mauricio Barahona
- Dr. Danny O'Hare
- Dr. Geoff Baldwin
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- David Featherbe
- Ciaran Mckeown
- James Mansfield

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- Faculty of Engineering
- Faculty of Natural Sciences

